Hypersecretion of corticotrophin-releasing hormone and arginine vasopressin in hypothyroid male rats as estimated with push–pull perfusion

A Tohei, G Watanabe and K Taya

Laboratory of Veterinary Physiology, Tokyo University of Agriculture and Technology, Fuchu, Tokyo 183, Japan

(Requests for offprints should be addressed to A Tohei, Laboratory of Veterinary Physiology, Tokyo University of Agriculture and Technology 3–5–8, Saiwai-cho, Fuchu, Tokyo 183, Japan)

Abstract

The relationship between hypothyroidism and disturbance of the hypothalamo–hypophysial–adrenal axis was investigated using adult male rats. Hypothyroidism was produced by administration of 4-methyl-2-thiouracil (thiouracil) in the drinking water for 2 weeks. Hypothyroidism decreased adrenal weights to 57% of controls and plasma concentrations of corticosterone to 48% of controls. The changes in the weight of adrenals recovered to control levels by administration of thyroxine. The pituitary responsiveness to corticotrophin-releasing hormone (CRH) and arginine vasopressin (AVP) for ACTH release markedly increased in hypothyroid rats as compared with euthyroid rats. In vivo release of CRH and AVP in median eminence significantly increased in hypothyroid rats as compared with euthyroid rats. There were no significant differences in hypothalamic concentrations of CRH and AVP.

These results indicate that hypothyroidism causes adrenal dysfunction directly and results in hypersecretion of ACTH mediated by increases in synthesis of CRH and AVP in the hypothalamus.


Introduction

Thyroid hormone is important for the growth, development and metabolism of many tissues. It has been shown to play an important role in adrenal function. Hypothyroidism has been reported to reduce the weight of adrenals (McCarthy et al. 1959, Kamilaris et al. 1991) and the plasma concentration of corticosterone, and to affect circadian adrenocortical rhythm (Murakami et al. 1984). In contrast, excess amounts of thyroid hormone increases the weight of adrenals (Boler & Moore 1982). Thyroidectomy decreases plasma and pituitary adrenocorticotropic hormone (ACTH) levels (Murakami et al. 1984), and causes a reduction in corticotrophin–releasing hormone (CRH) gene transcription (Shi et al. 1994) in the hypothalamic paraventricular nucleus (PVN). On the other hand, recent reports show that the pituitary content of ACTH increases (Tohei et al. 1997) and that ACTH responses to exogenous CRH are exaggerated in hypothyroid rats (Kamilaris et al. 1991, Tohei et al. 1997). Although it is obvious that thyroid hormone affects the hypothalamo–hypophysial–adrenal axis, the site of action of thyroid hormone on this axis is not clearly understood.

In order to clarify the hypothalamo–hypophysial–adrenal axis activity in hypothyroid rats, the pituitary responsiveness to CRH and (AVP) for ACTH release were examined in the present study. CRH and AVP release in median eminence (ME) were also measured directly using a push–pull perfusion method in adult male hypothyroid rats.

Materials and Methods

Animals

Adult male rats (350–400 g) of the Wistar strain were used throughout the present study. Animals were maintained with a ratio of 14 h light:10 h darkness (lights on at 0500 h). The room was kept between 23 and 26 °C. Rats received a standard laboratory diet and water and were allowed to feed ad libitum. Five animals of each group were used in the present experiment. Hypothyroidism was induced by administration of 0.03% 4-methyl-2-thiouracil (thiouracil; Wako Pure Chemical Industries Ltd, Osaka, Japan) in the drinking water for 2 weeks. Thyroid hormone replacement was performed daily by intraperitoneal (i.p.) injection of l-thyroxine (T4; 15 μg per rat, Sigma Chemical Co., St Louis, MO, USA) for 1 week before the experiment. Twenty-four hours before each experiment, a cannula (Dow Corning, Midland, MI, USA) was inserted into the right atrium via the external jugular vein in each rat for drawing blood samples. Blood (200 μl) was
withdrawn through the atrial cannula into a heparinized syringe without anaesthesia. After separation of the plasma by centrifugation at 1700g for 15 min at 4 °C, the red blood cells were resuspended in the same volume of 0.85% (w/v) NaCl solution (saline) and returned to the animal through the cannula.

Effects of thiouracil on the thyroid gland and pituitary–adrenal axis

To confirm the effects of hypothyroidism on the thyroid and adrenal gland, animals were killed by decapitation at 0900 h 2 weeks after administration of thiouracil. Blood was collected and centrifuged for the determination of plasma concentrations of tri-iodothyronine (T3), T4, thyroid stimulating hormone (TSH), ACTH and corticosterone. After decapitation, the adrenal glands and the hypothalami were removed and weighed. The hypothalami were homogenized and centrifuged at 25,000 g for 30 min at 4 °C. Supernatants were stored at −50 °C until assayed for CRH and AVP.

To investigate the pituitary responsiveness to CRH and AVP, CRH and AVP challenges were performed 2 weeks after administration of thiouracil. CRH (1 or 10 µg, Peptide Institute Inc., Osaka, Japan) and AVP (0·1 or 1 µg, Peptide Institute Inc.) dissolved in 0·2 ml saline were injected through the cannula at 0900 h. Control animals received the same volume of vehicle. Blood samples for plasma ACTH determinations were drawn from the cannula immediately before injection and 0·5, 1, 2 and 3 h after injection of CRH and 5, 10, 15, 30, 45 and 60 min after injection of AVP.

Push–pull perfusion protocol

Animals were anaesthetized with sodium pentobarbital (40 mg/kg) and placed in a stereotaxic apparatus to implant the guide cannula with a removable stylette into the ME. Implantation coordinates (Paxinos & Watson 1986) were −3·14 mm anterior, −0·1 mm lateral and −10·2 mm ventral from bregma for ME. The cannula was fixed onto the skull with anchor screws and dental cement. Animals were given a minimum recovery period of 7 days. On the day of push–pull perfusion, the inner stylette was removed and replaced with the inner cannula perfusion assembly. Artificial cerebrospinal fluid (Watanobe & Takebe 1994) was administered through the push cannula at a flow rate of 15 µl/min. Perfusion fractions (300 µl) were collected every 20 min over a total period of 180 min (1830–2130 h). The perfusates were immediately frozen on dry ice and lyophilized. After completion of experiments, each animal was anaesthetized with ether and subjected to vascular perfusion with 0·9% (w/v) NaCl, followed by 10% formal–saline. The brain was removed and soaked in 10% formol–saline for 3 days. Frozen (−20 °C) serial 40-µm sections were cut along the plane of the cannula tract and stained with cresyl violet to confirm the position of the cannula.

Radioimmunoassay (RIA)

Concentrations of TSH were measured using NIDDK rat RIA kits for rat TSH. Hormone for iodination was rat TSH–I–9 and the antiserum used were anti-rat TSH–S–5. Results were expressed in terms of NIDDK rat TSH–RP–2. The intra- and interassay coefficients of variation were 6·6 and 7·9% respectively.

T3, T4 (Tohei et al. 1997), ACTH (Tomabechi et al. 1994), corticosterone (Kanesaka et al. 1992) and CRH (Suda et al. 1987) were measured by double-antibody RIAs using 125I-labelled radioligands as described previously. Antisera to T3 and T4 were kindly provided by Dr M Suzuki (Gunma University, Maebashi, Gunma, Japan). Antiserum to corticosterone was kindly provided by Dr G.D. Niswender (Colorado State University, Fort Collins, CO, USA). Antiserum to CRH was kindly provided by Dr T Suda (Hirosaki University, Aomori, Japan). The intra- and interassay coefficients of variation were 7·2 and 17·4% for T3, 9·4 and 10·9% for T4, 11·3 and 19·4% for ACTH, 9·8 and 17·5% for corticosterone, and 5·5 and 11·2% for CRH respectively. AVP was measured by a commercial RIA kit purchased from Mitsubishi Chemical Corporation (Tokyo, Japan). The detection limits (defined as the amount of hormone that reduced binding to 85% of that occurring in the absence of unlabelled hormone) were 1 pg/tube for CRH and 0·1 pg/tube for AVP.

Statistical analyses

All results are expressed as means ± s.e.m. The data of plasma concentrations of T3, T4, TSH and hypothalamic concentrations of CRH and AVP were analyzed by Student’s t-test but when more than two means were compared, an ANOVA was carried out and significance of the difference between means was determined by Duncan’s multiple range test. The data of organ weight and plasma concentration of corticosterone were analyzed using one-way analysis of variance (ANOVA) followed by Fisher’s protected least significant difference (PLSD) test. The effects of CRH and AVP challenges and the hypothalamic secretion of CRH and AVP were analyzed using two-way ANOVA followed by Fisher’s PLSD test; a value of P<0·05 was considered significant.

Results

Effects of thiouracil on thyroid gland and pituitary–adrenal axis

Plasma concentrations of T3 and T4 were markedly suppressed (T3, from 464 to 76·8 pg/ml; T4, 37·2 to
9.8 ng/ml) in male rats 2 weeks after administration of thiouracil. On the other hand, the plasma concentration of TSH in hypothyroid rats was significantly increased (from 0.76 to 7.26 ng/ml) as a result of administration of thiouracil (Fig. 1).

Adrenal weights and plasma concentrations of corticosterone were significantly lower in hypothyroid rats (35.62 mg and 35 ng/ml) as compared with intact rats (61.34 mg and 72.6 ng/ml). These changes recovered to control levels (57.28 mg and 84.6 ng/ml) after administration of T4 (Fig. 2).

In response to two doses of CRH, plasma levels of ACTH increased and reached a peak by 15 min after the injection in both groups of animals. The pituitary response to CRH for ACTH release was higher at high dose (10 µg) in hypothyroid than euthyroid rats, although there was no significant difference in low dose (1 µg) CRH-injected groups. In response to two doses of AVP, plasma levels of ACTH increased and reached a peak by 5 or 10 min after the injection in both groups of animals. The pituitary response to AVP was higher with both doses (0.1 and 1 µg) in hypothyroid than in euthyroid rats (Fig. 3).

Hypothalamic secretion of CRH and AVP as estimated with push-pull perfusion

In vivo release of CRH and AVP in ME by push–pull perfusion significantly increased in hypothyroid rats as compared with euthyroid rats (Fig. 4). There were no significant differences in hypothalamic concentrations of CRH and AVP (Fig. 5).

Discussion

In the present study, we observed decreases in adrenal weights and basal levels of plasma corticosterone in hypothyroid male rats at 2 weeks after administration of thiouracil. These changes in adrenal weights recovered to control levels by administration of T4. It has been reported that adrenal responsiveness to ACTH also decreased in hypothyroid rats (Tohei et al. 1997). Our present findings are supported by previous reports (McCarthy et al. 1959, Murakami et al. 1984).

The pituitary responsiveness to CRH and AVP for ACTH release significantly increased in hypothyroid rats as compared with control rats. We have previously reported that pituitary contents of ACTH also increased in hypothyroid male rats and the change recovered to control levels after administration of T4 (Tohei et al. 1997). Previous reports regarding the effect of hypothyroidism on ACTH secretion are conflicting. Murakami et al. (1984) reported that thyroidectomy resulted in a decrease in plasma and pituitary levels of ACTH in female rats. On the other hand, most recent reports have shown that ACTH responses to exogenous CRH were exaggerated, while corticosterone responses to ACTH were reduced in hypothyroid male rats (Kamilaris et al. 1991, Tohei et al. 1997). Other reports have shown that under conditions of stress, a marked increase in plasma levels of ACTH in
hypothyroid rats was observed, whereas the increase in plasma level of corticosterone in response to immobilization stress was much smaller in hypothyroid than in control rats (Tohei et al. 1997). In the present study, decreases in adrenal weights and plasma concentrations of corticosterone, and increases in the plasma concentrations of ACTH were observed in hypothyroid male rats. These results suggest that hypothyroidism causes adrenal dysfunction directly and results in hypersecretion of ACTH.

In the present study, it is also demonstrated that the secretion of CRH and AVP in ME increased in hypothyroid rats compared with euthyroid rats using the push–pull perfusion technique, though the hypothalamic contents of CRH and AVP were not different between the two groups. These results suggest that the synthesis of CRH and AVP in the hypothalamus probably increases in hypothyroid rats as compared with euthyroid rats. It has been known that CRH is a major physiological mediator of the hypothalamic control of ACTH secretion (Antoni 1986). In addition, it is also demonstrated that AVP amplifies the effect of CRH about 2– to 3-fold, and hence the slope of a dose–response curve for CRH or AVP alone will be less than that of AVP applied in combination with CRH (Gillies et al. 1982, Rivier & Vale 1983, Vale et al. 1983). Several laboratories reported that CRH and AVP are colocalized in parvicellular neurons of the PVN (Roth et al. 1982, Tramu et al. 1983, Mouri et al. 1993).
co-expression of CRH and AVP in the neurons have been demonstrated in adrenalectomized rats (Kiss et al. 1984, Sawchenko et al. 1984) as a model of adrenal dysfunction. Adrenalectomy markedly increases the levels of CRH and AVP immunoreactivity (Antoni et al. 1983, Paul & Gibbs 1983, Kiss et al. 1984), and the amount of mRNA transcripts of CRH and AVP gene (Wolfson et al. 1985, Antoni 1986, Davis et al. 1986) in the neurons of the PVN which contains glucocorticoid receptors in rats (Reul & De Kloet 1985). It has also been reported that adrenalectomy increases the concentration of CRH and AVP in hypophysial portal blood (Oliver et al. 1983, Kooy et al. 1990) and the median eminence (Suda et al. 1983, Holmes et al. 1986). In the present study, it is clearly demonstrated that adrenal dysfunction causes the hypersecretion of CRH, AVP and ACTH at the hypothalamus–pituitary level in hypothyroid male rats. In contrast to our results, a previous report by Shi et al. (1994) has shown that hypothyroidism causes a reduction in CRH gene transcripts in the PVN of male rats, with a concomitant decrease in both pro-opiomelanocortin (POMC) gene expression in the anterior pituitary gland and circulating corticosterone, though the plasma levels of POMC-related peptides were not reported. Although we cannot explain the differences between our results and those of Shi and colleagues we believe that the push–pull perfusion technique provides us with direct evidence that CRH and AVP release are increased from at least the ME.

In conclusion, hypothryoidism causes adrenal dysfunction directly, and results in hypersecretion of ACTH. The adrenal dysfunction leads to the hypersecretion of CRH and AVP from the hypothalamus in hypothyroid male rats. Therefore, reduced levels of thyroid hormone probably act on adrenal glands directly, and not at the hypothalamus–pituitary level.

Acknowledgements

We are grateful to the Rat Pituitary Hormone Distribution Program, NIDDK, NIH, Bethesda, MD, USA for providing RIA materials; Dr G D Niswender, Animal Reproduction and Biotechnology Laboratory, Colorado State University, Fort Collins, CO, USA for providing antisera to corticosterone (GDN B377); Dr M Suzuki, Gunma University, Maebashi, Japan for antisera to T3 and T4; Dr T Suda, Hirosaki University, Aomori, Japan for antisera to CRH; Dr R Hokao, The Imamichi Institute for Animal Reproduction, Ibaraki, Japan for Wistar rats and Dr J L Voogt, Department of Physiology, The University of Kansas Medical Center, KS, USA for the helpful suggestions. This work was supported by a grant-in-aid for Scientific Research from the Ministry of Education of Japan no. 0660375. A Tohei received a fellowship from the Japan Society for the Promotion of Science (JSPS).

References


Received 9 January 1997

Revised manuscript received 23 May 1997

Accepted 16 September 1997

---

*Journal of Endocrinology (1998) 156*, 395–400