Serum levels of insulin-like growth factor system components and relationship to bone metabolism in Type 1 and Type 2 diabetes mellitus patients

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Abstract

Osteopenia has been ascribed to diabetics without residual insulin secretion and high insulin requirement. However, it is not known if this is partially due to disturbances in the IGF system, which is a key regulator of bone cell function.

To address this question, we performed a cross-sectional study measuring serum levels of IGF-I, IGF-binding protein-1 (IGFBP-1), IGFBP-3, IGFBP-4 and IGFBP-5 by specific immunoassays in 52 adults with Type 1 ($n=27$) and Type 2 ($n=25$) diabetes mellitus and 100 age- and sex-matched healthy blood donors. In the diabetic patients, we further determined serum levels of proinsulin, intact parathyroid hormone (PTH), 25-hydroxyvitamin D₃, 1,25-dihydroxyvitamin D₃ and several biochemical bone markers, including osteocalcin (OSC), bone alkaline phosphatase (B-ALP), carboxy-terminal propeptide of type I procollagen (PICP), and type I collagen cross-linked carboxy-terminal telopeptide (ICTP). Urinary albumin excretion was ascertained as a marker of diabetic nephropathy. Bone mineral density (BMD) of hip and lumbar spine was determined by dual-energy X-ray absorptiometry. Data are presented as means $\pm$ S.E.M. Differences between the experimental groups were determined by performing a one-way analysis of variance (ANOVA), followed by Newman-Keuls test. Correlations between variables were assessed using univariate linear regression analysis and partial correlation analysis.

Type 1 diabetics showed significantly lower IGF-I (119 ± 8 ng/ml) and IGFBP-3 (2590 ± 104 ng/ml) but higher IGFBP-1 levels (38 ± 10 ng/ml) compared with Type 2 patients (170 ± 13 , 2910 ± 118 , 11 ± 3 respectively; $P<0.05$) or healthy controls (169 ± 5 , 4620 ± 192 , 3.5 ± 0.4 respectively; $P<0.01$). IGFBP-5 levels were markedly lower in both diabetic groups (Type 1, 228 ± 9 ;

Type 2, 242 ± 11 ng/ml) than in controls (460 ± 7 ng/ml, $P<0.01$), whereas IGFBP-4 levels were similar in diabetics and controls. IGF-I correlated positively with IGFBP-3 and IGFBP-5 and negatively with IGFBP-1 and IGFBP-4 in all subjects. Type 1 patients showed a lower BMD of hip ($83 \pm 2\%$, Z-score) and lumbar spine ($93 \pm 2\%$) than Type 2 diabetics ($93 \pm 5\%$, $101 \pm 5\%$ respectively), reaching significance in the female subgroups ($P<0.05$). In Type 1 patients, BMD of hip correlated negatively with IGFBP-1 ($r = -0.34$, $P<0.05$) and IGFBP-4 ($r = -0.3$, $P<0.05$) but positively with IGFBP-5 ($r = 0.37$, $P<0.05$), which was independent of age, diabetes duration, height, weight and body mass index, as assessed by partial correlation analysis. Furthermore, biochemical markers indicating bone loss (ICTP) and increased bone turnover (PTH, OSC) correlated positively with IGFBP-1 and IGFBP-4 but negatively with IGF-I, IGFBP-3 and IGFBP-5, while the opposite was observed with bone formation markers (PICP, B-ALP) and vitamin D₃ metabolites. In 20 Type 2 patients in whom immunoreactive proinsulin could be detected, significant positive correlations were found between proinsulin and BMD of hip ($r = 0.63$, $P<0.005$), IGF-I ($r = 0.59$, $P<0.01$) as well as IGFBP-3 ($r = 0.49$, $P<0.05$). Type 1 and Type 2 patients with macroalbuminuria showed a lower BMD of hip, lower IGFBP-5 but higher IGFBP-4 levels, suggesting that diabetic nephropathy may contribute to bone loss by a disturbed IGF system.

In conclusion, the findings of this study support the hypothesis that the imbalance between individual IGF system components and the lack of endogenous proinsulin may contribute to the lower BMD in Type 1 diabetics.

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Introduction

Bone remodeling is regulated by systemic hormones and locally produced factors acting in concert to maintain bone

mass. Insulin-like growth factor-I (IGF-I) is an important anabolic regulator of bone cell function (e.g. decreasing collagen degradation, increasing bone matrix deposition, and increasing osteoblastic cell recruitment; McCarthy

et al. 1989, Mohan 1993). The effects of IGF-I are modulated by at least six different IGF-binding proteins (IGFBPs) and IGFBP proteases (Jones & Clemmons 1995, Rajaram *et al.* 1997). The key role of the IGF system for bone formation is demonstrated by the finding that approximately 50% of basal bone cell proliferation can be blocked by inhibiting the actions of endogenously produced IGFs by bone cells in serum-free culture (Mohan *et al.* 1989, Mohan 1993). In bone cells, locally produced IGF system components (e.g. IGF-I) may act as potential endogenous mediators of parathyroid hormone (PTH) and calcitriol, and may modulate IGF actions both in a positive (e.g. IGFBP-5) and negative (e.g. IGFBP-4) manner (McCarthy *et al.* 1989, Mohan *et al.* 1989, Mohan 1993). Studies in patients with osteoporosis suggest that IGFBP-1 and IGFBP-4 are possible 'inhibitors' of IGF action, whereas IGFBP-3 may play a 'stimulatory' role (Rosen *et al.* 1992, Wüster *et al.* 1993).

A number of endocrine disorders, such as Type 1 diabetes mellitus, hypogonadism, hyperthyroidism, hyperparathyroidism and hypercorticism, may lead to osteopenia and osteoporosis. In juvenile-onset diabetics, a greater than 10% loss in forearm bone mass, characteristic of cortical bone, was measured in more than 50% of the subjects studied (Levin *et al.* 1976). Osteopenia has been ascribed to diabetics without residual insulin secretion and high insulin requirement (McNair *et al.* 1978), whereas a large population-based study reported an increased bone mineral density (BMD) in Type 2 diabetes (Van Daele *et al.* 1995). A long-term study investigating 16 Type 1 and 19 Type 2 patients over approximately 12 years (Krakauer *et al.* 1995) suggested that, in Type 1 patients, low bone formation retards bone accumulation during growth, metabolic effects of poor glycemic control lead to increased bone resorption and bone loss in young adults, and low bone turnover retards age-related bone loss.

There has been no study of the relationship between bone metabolism and circulating levels of IGF system components in the two types of diabetes. In Type 1 diabetics, decreased serum levels of IGF-I have been reported which are attributed to inadequate insulinization (Dills *et al.* 1995). Another study reported a diminished osteoblast function (Bouillon *et al.* 1995). In prepubertal Type 1 diabetics, it has been shown that the low circulating IGF-I levels contribute to impaired growth (Strasser-Vogel *et al.* 1995). Based on these findings, we proposed the hypothesis that bone metabolism in diabetic subjects may be influenced by circulating IGF system components and β -cell function. As a means of testing this hypothesis, we have measured, in a cross-sectional protocol, serum levels of IGF-I and IGFBPs-1, -3, -4 and -5 in patients with Type 1 and Type 2 diabetes and in non-diabetic control subjects. In addition, BMD of hip and lumbar spine, biochemical markers (Power & Fottrell 1991, Eriksen *et al.* 1993) indicating bone formation (bone alkaline phosphatase (B-ALP) isoenzyme and procollagen

carboxy-peptide (PICP)), bone turnover (osteocalcin; OSC), and bone resorption (type I collagen carboxy-terminal telopeptide; ICTP), as well as serum levels of proinsulin, insulin and C-peptide, were determined and correlated to circulating levels of IGF system components. The findings of this study are consistent with a potential role of circulating IGF system components and β -cell function in regulating bone metabolism in diabetic patients.

Subjects and Methods

Patients and study design

In this cross-sectional study we investigated 52 diabetic patients (27 Type 1 and 25 Type 2) who were recruited from 65 diabetics who had been screened in the outpatient clinic of the University Hospital Ulm during a 6-month period. Patients with bone fractures during the last year, degenerative spondylosis, Paget's disease, metastatic bone lesions, immobilization, or other severe illnesses (e.g. liver, heart and kidney failure) were excluded from the study. The normal ranges of IGF-I and IGFBP levels were obtained from 100 age- and sex-matched healthy blood donors (mean age: 57 ± 2.6 years; female/male: 49/51) who were randomly recruited from the University Hospital Ulm. Patients' characteristics are summarized in Table 1. All subjects gave informed consent to participate in this study. Body mass index (BMI) was calculated as weight (kg)/height² (m²). Six female Type 1 patients and twelve female Type 2 patients were postmenopausal (median 10 and 12 years). One woman with Type 1 diabetes was treated with sodium fluoride; one female Type 2 diabetic received transdermal estrogen. The other patients did not receive any drugs to prevent or treat osteoporosis. All Type 1 patients studied were C-peptide negative and were on intensified conventional insulin therapy with neutral protamin hagedorn (NPH) insulin twice daily to cover their basal insulin requirements and regular insulin before meals three times per day. In the Type 2 group, seventeen patients (female/male: 9/8) injected insulin twice daily (NPH insulin/regular insulin: $70 \pm 10\%/30 \pm 8\%$), two patients (female/male: 1/1) combined insulin (NPH insulin at bedtime) with glibenclamide in the morning, and six patients (female/male: 3/3) were treated with glibenclamide only.

Assays

All blood samples were non-fasting morning samples. Serum creatinine, calcium and phosphate were measured by standard biochemical methods. Haemoglobin A1c (HbA1c) was determined by HPLC to assess diabetes control (normal below 6.1%). As a marker of diabetic nephropathy, albumin excretion was measured in 24-h

Table 1 Clinical data and serum parameters of bone metabolism in Type 1 and Type 2 diabetes patients (mean values $\pm$ S.E.M.)

	Type 1 (n=12F/15M)	Type 2 (n=13F/12M)	P (ANOVA)
Clinical data			
Age (years)	48 $\pm$ 4 ^{§§} /54 $\pm$ 3 [#]	60 $\pm$ 2/64 $\pm$ 2	**
BMI (kg/m ²)	22.3 $\pm$ 0.5 ^{§§} /24 $\pm$ 0.6	28.3 $\pm$ 1.4 ^{§§} /24.5 $\pm$ 0.7	*
Duration of diabetes (years)	23 $\pm$ 2.3 [§] /20 $\pm$ 3.4	14 $\pm$ 2.1/16 $\pm$ 3.5	*
HbA1c (%)	8.1 $\pm$ 0.4 ^{§+} /7.4 $\pm$ 0.2	9.1 $\pm$ 0.4/8.2 $\pm$ 0.4	*
Injected insulin (IU/kg body weight)	0.55 $\pm$ 0.04/0.57 $\pm$ 0.05	0.53 $\pm$ 0.06/0.44 $\pm$ 0.07	
Creatinine (37–110 μ mol/l)	84 $\pm$ 6/90 $\pm$ 4	76 $\pm$ 4/95 $\pm$ 5	
Excretion of albumin in urine (μ g/min)	221 $\pm$ 177 ^{§§++} /6.3 $\pm$ 2.6 ^{##}	18 $\pm$ 10 ^{§§} /341 $\pm$ 145	**
Serum parameters of bone metabolism (reference values)			
Calcium (2.0–2.8 mmol/l)	2.4 $\pm$ 0.03/2.4 $\pm$ 0.02	2.4 $\pm$ 0.02/2.4 $\pm$ 0.03	
Phosphate (0.8–1.5 mmol/l)	1.1 $\pm$ 0.06/1.0 $\pm$ 0.05	1.1 $\pm$ 0.06/1.0 $\pm$ 0.05	
PTH (10–65 pg/ml)	26 $\pm$ 5.5/16 $\pm$ 3.6 [#]	25 $\pm$ 4.7/43 $\pm$ 3.2	*
25-OHD ₃ (16–74 ng/ml)	22.2 $\pm$ 3.5/17.6 $\pm$ 2	26.1 $\pm$ 3.5/24.2 $\pm$ 3.4	
1,25-(OH) ₂ D ₃ (10–60 pg/ml)	45 $\pm$ 4/54 $\pm$ 5	45 $\pm$ 5.3/40 $\pm$ 2	
OSC (F: 11–39 ng/ml) (M: 13–48 ng/ml)	17.5 $\pm$ 1.9/14.6 $\pm$ 1.7	14.7 $\pm$ 1.8/13.3 $\pm$ 1.2	
B-ALP (F: 7.5–15.7 ng/ml) (M: 8–16.8 ng/ml)	8.4 $\pm$ 0.6/11.7 $\pm$ 1	9.8 $\pm$ 1.3/9.6 $\pm$ 1	
PICP (F: 50–170 ng/ml) (M: 38–202 ng/ml)	137 $\pm$ 11/134 $\pm$ 9	115 $\pm$ 9/109 $\pm$ 10	
ICTP (F: 1.8–5 ng/ml) (M: 1.8–5 ng/ml)	3.9 $\pm$ 0.3/3.9 $\pm$ 0.7	3.1 $\pm$ 0.2/4.4 $\pm$ 0.5	
PICP versus ICTP	35.1/34.4 [#]	37.1 [§] /24.8	*

ANOVA: Type 1 vs Type 2: *= P <0.05; **= P <0.005; Newman–Keuls test: female Type 1 vs female Type 2: §= P <0.05, §§= P <0.005; male Type 1 vs male Type 2: #= P <0.05, ##= P <0.005; female Type 1 vs male Type 1: += P <0.05, ++= P <0.005; female Type 2 vs male Type 2: §= P <0.05; §§= P <0.005. M, male; F, female.

urine (Turbitimer; Behring, Marburg, Germany; normal below 20 μ g/min; microalbuminuria: 21–300 μ g/min; macroalbuminuria >300 μ g/min). To correct for sampling errors, urine was examined on at least two occasions.

To assess bone metabolism we determined, by RIA: B-ALP (bone-specific alkaline phosphatase; Hybritech, Köln, Germany), OSC (osteocalcin; CIS, Dreieich, Germany), ICTP (type I collagen cross-linked carboxy-terminal telopeptide; Pharmacia, Freiburg, Germany), and PICP (carboxy-terminal propeptide of type I procollagen; Pharmacia). 25-Hydroxyvitamin D₃ (25-OHD₃) and 1,25-dihydroxyvitamin D₃ (1,25-(OH)₂D₃) were measured by specific RIAs (Incstar, Stillwater, MN, USA) with a less than 2.5% cross-reactivity to other vitamin D metabolites. Intact PTH was measured using a chemiluminescence immunometric assay (Nichols, California, CA, USA) with intra- and interassay coefficients of variation of less than 7%.

The components of the IGF system were measured according to published protocols (Bang *et al.* 1994, Blum & Breier 1994, Mohan *et al.* 1995a, Honda *et al.* 1996). IGF-I was determined by a IGFBP-blocked specific RIA (Mediagnost, Tübingen, Germany) with a cross-reactivity to IGF-II of less than 0.05% and intra- and interassay coefficients of variation of less than 4% and 8%. As given by the manufacturer, the 5th, 50th, and 95th percentiles

for healthy subjects aged 30–40 years (n =82) were 109, 188 and 324 ng/ml; for ages 40–50 years (n =83): 103, 178 and 307 ng/ml; for ages 50–60 years (n =61): 97, 169 and 294 ng/ml; for ages 60–70 years (n =32): 91, 161 and 284 ng/ml respectively. IGFBP-3 was determined by a specific RIA with intra- and interassay coefficients of variation of less than 3.5% and 7.5%, and no cross-reaction with IGFBP-1 or IGFBP-2 (Mediagnost). As given by the manufacturer, the 5th, 50th, and 95th percentiles for healthy subjects aged 30–40 years (n =28) were 2080, 3180 and 4860 ng/ml; for ages 40–50 years (n =37): 2010, 3080 and 4700 ng/ml; for ages 50–60 years (n =32): 1960, 3020 and 4650 ng/ml; for ages 60–70 years (n =19): 1900, 2980 and 4680 ng/ml respectively. IGFBP-1 was assessed by enzyme immunoassay (Mediagnost) with intra- and interassay coefficients of variation of less than 4% and 8%. IGFBP-4 and IGFBP-5 were measured by specific RIAs with intra- and interassay coefficients of variation of less than 10%, as previously described (Mohan *et al.* 1995a, Honda *et al.* 1996). These studies revealed an increase in mean IGFBP-4 serum levels (healthy adults: aged 23–40 years (n =24): 404 $\pm$ 146 (s.d.) ng/ml; aged 41–60 years (n =37): 447 $\pm$ 87 ng/ml; aged 61–87 years (n =41): 546 $\pm$ 135 ng/ml), and a decrease in mean IGFBP-5 serum levels (healthy prepubertal girls: aged 12 $\pm$ 1.3 years (n =20):

Table 2 Serum levels of IGF-I and IGFBPs (mean values ± s.e.m.) in Type 1, Type 2 and control subjects (100 age- and sex-matched healthy blood donors, mean age: 57 ± 2.6 years; female/male: 49/51)

	Controls (n=49F/51M)	Type 1 (n=12F/15M)	Type 2 (n=13F/12M)	P (ANOVA)
Females				
IGF-I	167 ± 6	116 ± 12 [§]	159 ± 15	*
IGFBP-1	3.6 ± 0.4	47 ± 11 ^{§§+}	13 ± 5 [#]	**
IGFBP-3	4700 ± 257	2728 ± 171 ^{§§+}	3038 ± 135 ^{###}	**
IGFBP-4	379 ± 17	373 ± 46	368 ± 30	
IGFBP-5	445 ± 11	231 ± 12 ^{§§}	264 ± 12 ^{##}	**
Males				
IGF-I	170 ± 6	122 ± 10 ^{§+}	187 ± 21	*
IGFBP-1	3.2 ± 0.3	31 ± 14 ^{§§+}	10 ± 3 [#]	**
IGFBP-3	4560 ± 227	2474 ± 108 ^{§§}	2664 ± 167 ^{##}	**
IGFBP-4	370 ± 18	324 ± 27	340 ± 29	
IGFBP-5	477 ± 8	241 ± 13 ^{§§}	219 ± 14 ^{##}	**

ANOVA: Type 1 vs Type 2 vs Control: *=*P*<0.05, **=*P*<0.005; Newman–Keuls test: Type 1 vs Control; §=*P*<0.05, §§=*P*<0.005; Type 2 vs Control: #=*P*<0.05, ##=*P*<0.005; Type 1 vs Type 2: +=*P*<0.05. M, male; F, female.

664 ± 109 ng/ml; healthy adult women: aged 51 ± 31 years (*n*=30): 417 ± 194 ng/ml).

Serum insulin was measured by a specific enzyme immunoassay (DRG Instruments, Marburg, Germany) which did not cross-react with proinsulin. In Type 2 subjects, serum proinsulin and C-peptide were determined as markers of residual β-cell function in Type 2 diabetes by specific enzyme immunoassays (DRG Instruments). All three assays showed intra- and interassay coefficients of variation of less than 8% and 10%.

Measurement of BMD

To measure BMD we chose dual-energy X-ray absorptiometry (DEXA; HOLOGIC QDR 1000; Siemens Medical Systems, Erlangen, Germany) for its high precision (0.2%–2%), its high accuracy (3%–5%), its low radiation hazard with an absorbed dose of radiation of 10–30 µSv, and its short examination time of about 10 min (Kellie 1992). BMD was measured at hip (femoral neck, Ward’s triangle, trochanter region) and lumbar spine (L₁–L₄) in anterior–posterior projection. All measurements were corrected for superimposition artifacts. The BMD of each patient was calculated as the percentage of the mean value (Z-score %) obtained from a healthy control group matched for sex and age (HOLOGIC database).

Statistical analyses

Data are presented as individual values or as means ± s.e.m. except where otherwise indicated. Differences between the experimental groups were determined by performing a one-way analysis of variance (ANOVA), followed by Newman–Keuls test. The analysis was separated for additional parameters (e.g. sex). Correlations between variables were assessed using univariate linear

regression analysis. Partial correlation analysis was used to predict the correlations between IGF system components and BMD after correction for age, BMI and diabetes duration. *P*<0.05 was accepted as statistically significant.

Results

Clinical data and serum parameters

The clinical data and the serum parameters are given in Table 1. Type 1 patients were of younger age but showed a longer duration of the disease compared with the Type 2 group. All subjects showed normal serum values for creatinine, calcium, phosphate, PTH, 25-OHD₃, 1,25-(OH)₂D₃ and biochemical markers of bone turnover. Male Type 2 diabetics showed the highest PTH levels and a significantly lower ratio of PICP (indicating bone formation) to ICTP (a marker of bone loss).

IGF-I and IGFBP serum levels and interrelations

Type 1 patients revealed significantly lower IGF-I and IGFBP-3 but higher IGFBP-1 serum levels than Type 2 or control subjects (Table 2). IGF-I correlated negatively with IGFBP-1 (Type 1: *r*=−0.57, *P*<0.01; Type 2: *r*=−0.37, *P*<0.05) and IGFBP-4 (Type 1: *r*=−0.38, *P*<0.05; Type 2: *r*=−0.14, n.s.) but positively with IGFBP-3 (Type 1: *r*=0.56, *P*<0.01; Type 2: *r*=0.65, *P*<0.01) and IGFBP-5 (Type 1: *r*=0.76, *P*<0.005; Type 2: *r*=0.39, *P*<0.05). Figure 1 demonstrates these correlations for both diabetic groups. In the healthy control subjects, similar correlations between IGF-I and the IGFBPs were observed (female/male): IGF-I vs IGFBP-1 (*r*=−0.43/−0.42, *P*<0.01), IGF-I vs IGFBP-3 (*r*=0.47/0.49, *P*<0.005), IGF-I vs IGFBP-4 (*r*=−0.21/−0.22,

Figure 1 Relationship between IGF-I and IGFBPs in 52 diabetic patients. IGF-I correlated negatively with (A) IGFBP-1 and (C) IGFBP-4 and positively with (B) IGFBP-3 and (D) IGFBP-5.

$P < 0.05$) and IGF-I vs IGFBP-5 ($r = 0.21/0.41$, $P < 0.05/0.01$).

Relationship of IGF-I and IGFBPs to age and diabetes duration

Diabetes duration showed strong negative correlations with IGF-I (Type 1: $r = -0.64$, $P < 0.005$), IGFBP-3 (Type 1/2: $r = -0.44/-0.38$, $P < 0.05$) and IGFBP-5 (Type 1: $r = -0.6$, $P < 0.005$). Similar correlations were obtained with age, which was further positively correlated with IGFBP-1 (Type 1/2: $r = 0.39/0.32$, $P < 0.05$). The correlations between IGF-I and IGFBPs (Fig. 1) were still significant after correction for the variables age, diabetes duration, height, weight and BMI by partial correlation analysis.

BMD and relationship to IGF system, bone metabolism and clinical data

The BMDs of hip (Fig. 2A) and lumbar spine (Fig. 2B) were lower in Type 1 than in Type 2 patients, reaching statistical significance in female subjects. Male Type 2 diabetics showed a lower BMD of lumbar spine than females ($P < 0.005$). BMD of hip correlated positively with BMD of lumbar spine (Type 1: $r = 0.41$, $P < 0.05$; Type 2: $r = 0.76$, $P < 0.001$). In Type 1 patients, BMD of hip

correlated negatively with IGFBP-1 (Fig. 3A) and IGFBP-4 (Fig. 3B), and positively with IGFBP-5 (Fig. 3C). Furthermore, BMD of hip correlated positively with $1,25-(\text{OH})_2\text{D}_3$ ($r = 0.56$, $P < 0.01$) and negatively with microalbumin in urine ($r = -0.46$, $P < 0.05$), serum levels of PTH ($r = -0.33$, $P < 0.05$) and the bone resorption marker ICTP ($r = -0.31$, $P < 0.05$). On the other hand, in Type 2 patients, IGFBP-3 was positively correlated with BMD of hip ($r = 0.38$, $P < 0.05$) and lumbar spine ($r = 0.45$, $P < 0.05$). In all subgroups, there was a significant positive relationship between BMD and BMI. The group with the highest BMI, female Type 2 patients, also showed the strongest correlation (BMI vs BMD of hip: $r = 0.59$; and lumbar spine: $r = 0.49$, $P < 0.05$). In contrast, female Type 1 patients not only showed the lowest BMD, lower levels of IGF-I, IGFBP-3 and IGFBP-5 and higher levels of IGFBP-1 and IGFBP-4, but also the lowest BMI and the longest diabetes duration (Table 1). Therefore, we performed partial correlation analysis to further assess whether circulating IGF system components contribute independently of age, diabetes duration, height, weight and BMI to BMD in both types of diabetes mellitus. In Type 1 subjects, BMD of hip still correlated negatively with IGFBP-4 ($r = -0.33$, $P < 0.05$) and positively with IGFBP-5 ($r = 0.57$, $P < 0.01$), while in Type 2 patients the positive relationship between IGFBP-3 and BMD of hip or lumbar spine lost its statistical significance ($r = 0.21$ and 0.23 respectively).

Figure 2 BMD (Z-score %) measured at (A) hip and (B) lumbar spine in 27 Type 1 and 25 Type 2 diabetes patients. The reference values of age- and sex-matched healthy controls were obtained from the HOLOGIC database and are defined as 100% (shaded bar=1 S.D.).

Relationship between IGF system components, markers of bone metabolism and diabetes therapy

In Type 1 patients, IGF-I correlated positively with the bone formation marker B-ALP ($r=0.33$, $P<0.05$) and showed a negative correlation with ICTP ($r=-0.42$, $P<0.05$), an acknowledged marker of bone loss. IGFBP-1 showed strong positive correlations with ICTP ($r=0.8$, $P<0.001$) and OSC ($r=0.64$, $P<0.005$), IGFBP-3 correlated positively with the bone formation marker PICP ($r=0.3$, $P<0.05$), and IGFBP-4 showed significant positive correlations with both ICTP ($r=0.42$, $P<0.05$) and PTH ($r=0.68$, $P<0.005$), whereas the opposite was found with IGFBP-5 ($r=-0.5$ and -0.31 respectively, $P<0.05$). Among the vitamin D₃ metabolites, 25-OHD₃ correlated positively with IGFBP-3 ($r=0.51$, $P<0.05$) and 1,25-(OH)₂D₃ with IGFBP-5 ($r=0.34$, $P<0.05$). The

Figure 3 Relationship between BMD of hip and IGFBPs in Type 1 patients. BMD of hip correlated negatively with (A) IGFBP-1 and (B) IGFBP-4 and positively with (C) IGFBP-5.

dose of injected insulin correlated positively with IGF-I ($r=0.37$, $P<0.05$) and B-ALP ($r=0.31$, $P<0.05$) but negatively with OSC ($r=-0.30$, $P<0.05$) and ICTP ($r=-0.36$, $P<0.05$). In Type 2 diabetics, there was a positive relationship between ICTP and PICP ($r=0.45$, $P<0.05$) which was not observed in Type 1 diabetics, indicating that the increased rate of bone resorption was not associated with a compensatory increase in the rate of bone formation in the latter group (i.e. negative

Table 3 BMD, IGF system components and 1,25-(OH)₂D₃ levels (mean values ± S.E.M.) in diabetic subjects without microalbuminuria (–microA), with microalbuminuria (+microA; incipient diabetic nephropathy) and with macroalbuminuria (+macroA; manifest diabetic nephropathy)

	–microA (n=37 (22/15)) [†]	+microA (n=10 (2/8)) [†]	+macroA (n=5 (2/3)) [†]	P (ANOVA)
BMD of hip (Z-score %)	90 ± 3	88 ± 9	82 ± 12	
IGFBP-4 (ng/ml)	341 ± 183	332 ± 108	532 ± 19 ^{§#}	*
IGFBP-5 (ng/ml)	239 ± 51	223 ± 62	210 ± 25	
IGFBP-5 vs IGFBP-4	0.7	0.67	0.39 ^{§#}	*
1,25-(OH) ₂ D ₃ (pg/ml)	52 ± 17	57 ± 15	28 ± 5 ^{§#}	*

ANOVA: (–microA vs +microA vs +macroA): * = $P < 0.05$, Newman–Keuls test: (–microA vs +macroA): § = $P < 0.05$, (+microA vs +macroA): # = $P < 0.05$. [†]Number of subjects in each group (Type 1/Type 2).

uncoupling of bone formation to resorption). In the insulin-treated Type 2 patients, the dose of injected insulin revealed significant negative correlations with IGFBP-4 ($r = -0.44$, $P < 0.05$), OSC ($r = -0.35$, $P < 0.05$) and ICTP ($r = -0.34$, $P < 0.05$).

Relationship between residual β -cell function, IGF system and BMD in Type 2 diabetes

In 20 Type 2 subjects (female/male: 11/9) proinsulin ($85 \pm 22/50 \pm 16$ pg/ml) and C-peptide ($2.2 \pm 0.5/2.8 \pm 0.7$ ng/ml) could be detected in serum. In these patients, proinsulin levels showed stronger positive correlations with BMD of hip ($r = 0.63$, $P < 0.005$) and lumbar spine ($r = 0.58$, $P < 0.005$) than C-peptide, whereas insulin levels did not correlate. Furthermore, proinsulin showed significant positive correlations with IGF-I ($r = 0.59$, $P < 0.01$), IGFBP-3 ($r = 0.49$, $P < 0.05$) and BMI ($r = 0.35$, $P < 0.05$).

Relationship of renal function to IGF system and bone metabolism

Serum creatinine levels correlated positively with age (Type 1: $r = 0.44$, $P < 0.05$), IGFBP-1 (Type 1: $r = 0.52$, $P < 0.05$), IGFBP-4 (Type 1/2: $r = 0.64/0.5$, $P < 0.005/0.05$) and markers indicating bone loss such as ICTP (Type 1/2: $r = 0.72/0.65$, $P < 0.005$), or bone turnover such as OSC (Type 1: $r = 0.71$, $P < 0.005$) and PTH (Type 1: $r = 0.61$, $P < 0.005$). On the other hand, serum creatinine correlated negatively with IGF-I (Type 1: $r = -0.45$, $P < 0.05$), IGFBP-5 (Type 1/2: $r = -0.38/-0.31$, $P < 0.05$) and 1,25-(OH)₂D₃ (Type 2: $r = -0.8$, $P < 0.001$). The excretion of albumin in urine, an acknowledged marker of diabetic nephropathy, was measured within the normal range (< 20 μ g/min) in 37 patients. In 10 patients microalbuminuria was observed (117 ± 9 μ g/min), while 5 patients showed macroalbuminuria (1309 ± 330 μ g/min), indicating manifest diabetic nephropathy. Linear regression analysis showed that the excretion of albumin in urine correlated positively with IGFBP-4 (Type 1/2: $r = 0.47/0.37$, $P < 0.05$) and negatively with the BMD of

hip (Type 1: $r = -0.46$, $P < 0.05$). As demonstrated in Table 3, the macroalbuminuric patients showed a lower BMD of hip, a lower molar ratio of IGFBP-5 vs IGFBP-4 and lower 1,25-(OH)₂D₃ levels.

Discussion

The current study provides the first analysis of immuno-reactive serum levels of IGF-I, IGFBPs-1, -3, -4 and -5 and proinsulin together with biochemical markers of bone metabolism, BMD and renal function in a group of Type 1 and Type 2 diabetic patients. Type 1 diabetics showed markedly lower IGF-I and higher IGFBP-1 levels compared with Type 2 patients and control subjects, consistent with several previous reports (Crosby *et al.* 1992, Dills *et al.* 1995, Hilding *et al.* 1995, Strasser-Vogel *et al.* 1995). Compared with published normal ranges for the assays, IGF-I levels in Type 1 diabetics were below the 20th percentile. IGFBP-3 levels were below the 50th percentile in Type 1 patients and in male Type 2 patients. The imbalance in circulating IGF-I, IGFBP-1 and IGFBP-3 levels appeared to be characteristic of insulin-deficient Type 1 diabetics rather than of Type 2 subjects with residual β -cell function. This is pathophysiologically plausible since insulin inhibits IGFBP-1 production but stimulates the synthesis of IGF-I (Bereket *et al.* 1995) and both IGFBP-1 and glucose are significant predictors for the levels of IGF-I (Hong *et al.* 1997). In line with these reports, we observed a negative relationship between diabetes duration and serum levels of IGF-I and IGFBP-3, as well as a positive correlation between serum IGF-I and the amount of injected insulin in Type 1 diabetics. IGFBP-3 levels were below the 50th percentile in Type 1 patients and in male Type 2 patients. For the first time, our study demonstrated that IGFBP-5 serum levels are substantially lower in both types of diabetes mellitus patient than in age- and sex-matched healthy controls. Like IGF-I and IGFBP-3, IGFBP-5 levels decrease with age (Mohan *et al.* 1995a). Thus, higher IGFBP-5 levels should be expected in the younger Type 1 diabetics. However, the strong negative relationship between

diabetes duration and IGFBP-5 suggests that diabetes plays an important role. In all subjects studied, serum IGF-I showed negative correlations with IGFBP-1 and positive correlations with IGFBP-3, which is in good accordance with previous reports (Argente *et al.* 1993, Blum & Breier 1994, Hilding *et al.* 1995). Our data further show that IGF-I correlated negatively with IGFBP-4 but positively with IGFBP-5. Taken together, this supports the concept that IGFBPs may predominantly stimulate (e.g. IGFBP-3, IGFBP-5) or inhibit (e.g. IGFBP-1, IGFBP-4) IGF action; however, the same IGFBP may exert stimulatory and inhibitory effects depending on the tissue type or the local conditions (Mohan *et al.* 1989, Jones & Clemmons 1995, Mohan *et al.* 1995a,b, Rajaram *et al.* 1997).

The finding that BMD is lower in Type 1 compared with Type 2 patients is in accordance with several other reports (Levin *et al.* 1976, McNair *et al.* 1978, Krakauer *et al.* 1995, Van Daele *et al.* 1995). We further observed a lower BMD of lumbar spine in male than in female Type 2 diabetics. The higher PTH levels and the lower ratio of PICP to ICTP may indicate a mild secondary hyperparathyroidism which is typically found in the early stages of declining renal function (*vide infra*). Considering the positive relationship between BMD and BMI, it is likely that the nutritional status may contribute considerably to bone formation. Several studies indicated that protein intake is especially closely related to the IGF system and osteoporosis (Bonjour *et al.* 1997). Furthermore, it is important to address how alterations in circulating IGFBPs may influence the bioavailability of IGFs for target tissues such as bone. Since the modulation of IGF action is rather complex (Rajaram *et al.* 1997), the key question beyond the present study is whether individual IGF system components may contribute to bone loss (Rosen *et al.* 1994). Although our cross-sectional study cannot prove etiological relationships, our data suggest an association between circulating IGF system components and bone metabolism. The significant correlations between circulating IGF system components and BMD in Type 1 diabetics persisted after statistical correction for the variables BMI, age and diabetes duration. Similar to our results, decreased serum levels of IGF-I and IGFBP-3 and significant positive correlations between these IGF system components and BMD have been reported in patients with osteoporosis (Wüster *et al.* 1993). Another study demonstrated that serum levels of IGF-I showed a significant positive correlation with the osteoblastic surface measured by histomorphometric techniques (Reed *et al.* 1995). The role of circulating IGF-I in stimulating bone growth and metabolism is clearly supported by studies showing that (1) the restoration of the markedly reduced serum IGF-I levels by injection of recombinant human (rh)IGF-I dramatically stimulates longitudinal growth in patients with Laron syndrome (growth hormone (GH) receptor defect; Laron & Klinger 1994), and (2) the subcutaneous application of rhIGF-I stimulated bone turnover in healthy young adults

(Mauras *et al.* 1996) as well as in postmenopausal women (Eberling *et al.* 1993). Consistent with the anabolic role of IGF-I in bone (Mohan 1993, Narusawa *et al.* 1995, Reed *et al.* 1995, Ono *et al.* 1996), we suggest that in Type 1 patients the lower IGF-I levels may contribute to diminished stimulation of bone formation, leading to a fall in BMD with time. In this context it should be considered that not only IGF-I but also IGFBPs may directly act on bone cells. *In vitro* studies have shown that IGFBP-4 is a potent inhibitor of IGF actions on bone cells by preventing the IGFs from binding to their receptors (Mohan *et al.* 1989). On the other hand, IGFBP-5 may exert anabolic effects on osteoblasts by an IGF-independent mechanism involving IGFBP-5-specific binding sites (Mohan *et al.* 1995b). For the first time, our study has demonstrated that BMD of hip correlates negatively with circulating IGFBP-4 and positively with IGFBP-5 in Type 1 diabetics. Although the exact functional role of serum IGFBP-4 and IGFBP-5 is not clear at this time, increased IGFBP-4 serum levels have been previously reported in patients with elevated serum PTH levels and hip fractures (Rosen *et al.* 1992). Further studies are needed to determine whether the measurement of circulating IGF system components can be used as a suitable tool for the diagnosis and therapeutic monitoring of osteopenia and osteoporosis in Type 1 diabetes.

The finding that OSC and B-ALP are in the low normal range in both diabetic groups indicates an impaired bone turnover and formation which has been previously reported in both types of diabetes mellitus (Bouillon *et al.* 1995, Krakauer *et al.* 1995). In a recent study, it has been demonstrated that GH treatment of GH-deficient children increases serum levels of IGF-I, IGFBP-3, IGFBP-5 and B-ALP (Ono *et al.* 1996). Consistent with these data, we observed that IGF-I, IGFBP-3 and IGFBP-5 showed positive correlations with B-ALP, an acknowledged marker of bone formation, and also with 25-OHD₃ and 1,25-(OH)₂D₃. In contrast, IGFBP-1 and IGFBP-4 showed positive correlations with PTH and with ICTP, a marker of bone resorption, as well as with OSC, a marker of bone turnover (Power & Fottrell 1991). The finding that the age of Type 1 patients revealed a stronger direct correlation with ICTP than with PICP supports the well-established concept that bone resorption increases but matrix formation decreases with age (Parfitt *et al.* 1983, Eriksen *et al.* 1993, Bouillon *et al.* 1995). In contrast to Type 2 patients, in Type 1 there was no significant positive correlation between ICTP and PICP, indicating that the increased rate of bone resorption was not associated with a compensatory increase in the rate of bone formation (i.e. negative uncoupling of bone formation to resorption).

For the first time, our study also suggests a role for proinsulin in the regulation of bone formation in Type 2 diabetics. Earlier reports in Type 2 diabetes described a relationship between obesity and elevated proinsulin levels

(Nagi *et al.* 1990), and positive correlations between urinary C-peptide, BMD and BMI (Wakasugi *et al.* 1993). We observed that proinsulin not only showed stronger positive correlations with BMD than C-peptide, but that it also correlated positively with IGF-I and IGFBP-3. Thus, it can be speculated that, in Type 2 diabetics, residual proinsulin secretion may protect from bone loss. Whether proinsulin may stimulate bone formation via the IGF system, a changed nutritional status or by an independent mechanism (e.g. via specific receptors; Jehle *et al.* 1996) remains to be investigated.

Although all subjects showed normal serum creatinine levels, even early diabetic nephropathy may contribute to bone loss. In Type 1 patients, we observed a significant negative correlation between the extent of microalbuminuria and BMD of hip. In the Type 2 group, the male subjects showed significantly higher levels of urinary microalbumin and a lower BMD of hip and lumbar spine compared with females. Compared with patients with microalbuminuria, diabetic patients with macroalbuminuria showed a lower ratio of IGFBP-5 to IGFBP-4, which was mainly due to elevated IGFBP-4 levels. Thus, we conclude that diabetic nephropathy further disturbs the balance between circulating IGF system components and thereby may act as a co-factor for the development of osteopenia in diabetes mellitus.

In summary, patients with Type 1 diabetes showed a lower bone mass, lower IGF-I serum levels and alterations in the pattern of IGFBPs compared with Type 2 diabetics and controls. In Type 2 diabetics, pancreatic proinsulin secretion and obesity may protect from bone loss, possibly via the IGF system.

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