The de novo synthesis of numerous proteins is decreased during vitamin D₃ deficiency and is gradually restored by 1,25-dihydroxyvitamin D₃ repletion in the islets of Langerhans of rats

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Abstract

Since both the release and de novo biosynthesis of insulin are severely decreased by vitamin D₃ deficiency and improved by 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃) repletion following a 6-h delay in the rat, the present experiments investigated the effects of vitamin D₃ deficiency on the biosynthesis of heavier molecular weight proteins using electrophoretic separation. Gel protein staining by Coomassie blue showed very different profiles for islets protein production from 4-week vitamin D₃-deficient rats compared with normal islets. The pattern was characterised by a decrease in high molecular weight proteins, concomitantly accompanied by an increase in low molecular weight proteins. This tendency was partially reversed in vivo by 1,25(OH)₂D₃ repletion treatment for 7 days and was evident after only 16 h of treatment.

In parallel with these in vivo observations, which represent a static index of islets protein production, a kinetic study was performed in vitro by a double-labelling method allowing us to measure the de novo synthesis of proteins in islets during a strong 16·7 mM glucose stimulation. Comparison of ³H and ¹⁴C labelled samples was achieved via coelectrophoresis to avoid experimental artefacts. The study of the ratio of d.p.m. ³H/d.p.m. ¹⁴C for each molecular weight protein in islets stimulated by 16·7 mM glucose (versus basal 4·2 mM glucose) showed an increase in the height of certain peaks: 150, 130 and 8·5 kDa.

Under the same conditions, islets from 4-week vitamin D₃-deficient rats (versus normal islets) presented a large deficit of numerous newly synthesised proteins and particularly those implicated in the response to glucose stimulation. In vitro repletion of 1,25(OH)₂D₃ tended to reverse, at least in part, the deleterious effect of vitamin D₃ deficiency on the de novo protein synthesis of islets but these effects were gradual. Indeed, there was no detectable effect at 2 h incubation, but 1,25(OH)₂D₃ increased the 60 to 65 kDa, 55 kDa, and 9 to 8 kDa molecular mass proteins at 4 h, and increased the level of most newly synthesised proteins at 6 h. These data support the hypothesis of a beneficial genomic influence of 1,25(OH)₂D₃ that occurs progressively within the islets of Langerhans and which may prepare the β cells for an enhanced response to glucose stimulation.


Introduction

In the rat, as in several other species, vitamin D₃ and 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃), its biologically active principal metabolite, play crucial roles in the maintenance of calcium homeostasis. It is generally accepted that 1,25(OH)₂D₃ acts via specific nuclear receptors, found in target organs such as intestine, bone and kidney (Norman et al. 1982), but also in many non-classical target tissues (Walters 1992). The rat endocrine pancreas is one of these non-classical target tissues presenting 1,25(OH)₂D₃ receptors (Johnson et al. 1994), even in vitamin D₃-deficient rats (Clark et al. 1980, Stumpf et al. 1981, Ishida & Norman 1988). Several studies have demonstrated a regulatory role for 1,25(OH)₂D₃ in improving the insulin release that is dramatically reduced by vitamin D₃ deficiency (Norman et al. 1980, Clark et al. 1981, Chertow et al. 1983, Labriji-Mestaghanni et al. 1988). 1,25(OH)₂D₃ acts, at least in part, as a steroid in numerous tissues. Indeed, specific intracellular receptors facilitate the nuclear action of 1,25(OH)₂D₃ (Pike 1985): binding to promoter sequences in the genome causing up- or down-regulation of the transcription of various genes (Minghetti & Norman 1988), activating mRNA production coding for several de novo synthesised proteins (Norman et al. 1982), and...
influencing the biosynthesis of many proteins in various tissues (Verhaeghe et al. 1989, Brunner & De Boland 1990, Chang & Price 1991, Mouland & Hendy 1991). The genomic actions of this steroid on β cells of the endocrine pancreas have been emphasised (Norman et al. 1982, Faure-Dussert et al. 1997). 1,25(OH)₂D₃ activates the β cell insulin response to glucose in vitro after a delay of 3 to 20 h (Ishida et al. 1983, Kodawaki & Norman 1985, Cadé & Norman 1987, Ozono et al. 1990), or after 6 h in vitro (Billaudel et al. 1990), via an improvement of calcium handling occurring after a 4-h delay (Billaudel et al. 1990), increasing both Ca²⁺ entry by voltage-dependent channels and Ca²⁺ mobilisation from Ca²⁺ stores (Billaudel et al. 1993). However, these beneficial influences of 1,25(OH)₂D₃ can be observed more rapidly (within 45 min) when the protein kinase C (PKC) pathway of the β cell is stimulated by acetylcholine (Billaudel et al. 1995) or by PKC activators such as phorbol esters 12-O-tetradecanoyl-phorbol-13-acetate (TPA) (Billaudel et al. 1997). The existence of variable delays for the actions of 1,25(OH)₂D₃ supports the hypothesis of a genomic action of the steroid on the biosynthesis of several β cell proteins implicated in the different steps of the insulin excitation-secretion coupling.

In a previous study, 1,25(OH)₂D₃ was shown to increase insulin mRNA levels (Ozono et al. 1990), and we also found that it can stimulate the biosynthesis of insulin and low molecular weight proteins (Bourlon et al. 1999). The latter protein biosynthesis studies were performed in non-denaturing and non-reducing conditions with column chromatography separation, which is better adapted for small proteins. In the present study, we used a polyacrylamide gel electrophoresis separation in order to study the biosynthesis of other proteins within the islets of Langerhans which could be influenced by vitamin D₃ deficiency and by 1,25(OH)₂D₃ in vitro repletion (1 day and 7 days treatments). To investigate whether 1,25(OH)₂D₃ selectively stimulated newly synthesised islet proteins, we performed double labelling experiments which were completed by a time-course study of the effects of 1,25(OH)₂D₃ in vitro.

**Materials and Methods**

**Animals and isolation of islets of Langerhans**

After weaning on the 21st post-natal day, Wistar rats (CERJ, Le Genest-Saint-Isle, France) received either a normal balanced diet (AO4, UAR, Epinay sur Orge, France) or a rachitogenic diet (US Biochemical Corporation, Cleveland, OH, USA) lacking vitamin D₃ but containing low calcium (0.50% w/w) and phosphate (0.30% w/w) for 4 weeks. Rats were housed in a dark room and had free access to food and water. As previously described (Labriji-Mestaghanni et al. 1988, Bourlon et al. 1996) such a 4-week vitamin D₃ deficiency induced rachitism, with a smaller body weight (× 0.3), hypoglycaemia (× 0.7), and relative hypocalcaemia (× 0.8). Pancreatic islets were isolated by collagenase digestion (Lacy & Kostianovsky 1967). In vitro experiments using islets could then be performed after a 30-min equilibration period.

All animal experiments were carried out in accordance with the guidelines laid down by the French Ministère de l’Agriculture et du Développement Rural.

In vivo 1,25(OH)₂D₃ administration

One group of 4-week vitamin D₃-deficient rats received 1,25(OH)₂D₃ in vivo (Hoffman-La Roche, Basel, Switzerland) for 7 days during the last week of vitamin D₃ deficiency. This treatment consisted of 50 µl i.p. injections of 1 µg/kg/day 1,25(OH)₂D₃ dissolved in ethanol and 0.9% NaCl (50% v/v), for 7 days. This group of rats was called the 1,25(OH)₂D₃-replete group (+D7). Another group of deficient rats received just a single i.p. injection of 1,25(OH)₂D₃ 16 h before the experiments, and was called 1-day treated group (+D1).

In vitro 1,25(OH)₂D₃ administration

1,25(OH)₂D₃ was added directly to isolated islets of Langerhans in the incubation medium for various times from 2 to 6 h at two concentrations: either 10⁻⁸ M which was the most commonly used, or 10⁻¹² M as a control as this dose is not considered to be biologically active (Billaudel et al. 1990). Previous work has shown an identical level of islets insulin release for such 10⁻¹⁲ M controls and other controls using the vehicle alone (Bourlon et al. 1999). The medium with freshly prepared 1,25(OH)₂D₃ was changed every 2 h in an attempt to limit 1,25(OH)₂D₃ degradation and the well known retroinhibition exerted by insulin in a closed medium (Iversen & Miles 1971). The final concentration of ethanol (the 1,25(OH)₂D₃ vehicle) in the medium was 1‰ v/v.

**Islet incubations**

Groups of 100 islets were distributed into microvials (gassed with 95% O₂-5% CO₂ to maintain pH 7.4 of the incubation medium) at 37 °C with mild shaking. The incubation medium consisted of a Krebs bicarbonate buffer (116 mM NaCl, 5 mM KCl, 1 mM MgCl₂, 24 mM NaHCO₃) containing 0.5 mM calcium and 0.5% bovine serum albumin (fraction V, RIA grade, Sigma, Aldrich Chimie, St Quentin Fallavier, France). It was enriched by an amino acid mixture (in mM: alanine 0.1; arginine 0.1; cysteine 0.05; histidine 0.05; isoleucine 0.2; leucine 0.2; lysine 0.2; methionine 0.05; threonine 0.2; tryptophan 0.02; tyrosine 0.1; valine 0.2) to favour protein biosynthesis. Leucine was absent in the medium of islets prepared for incorporation of labelled leucine. The medium was supplemented with 8.3 mM glucose. During the last two
hours of induction islets were either stimulated with 16·7 mM glucose or were not stimulated (4·2 mM) in the presence of labelled leucine. Aliquots for the total protein content of islets were assayed by protein dye binding (Bradford 1976) before polyacrylamide gel electrophoresis.

Labelling of newly synthesised proteins

A radio labelled amino acid, leucine, was incorporated into newly synthesised islet proteins during the last 2-h incubation in the presence of 1,25(OH)2D3 induction and 16·7 mM glucose stimulation. To compare the protein biosynthetic capacities of islets from different groups with rigorously similar conditions, we used a double labelling method (Drittanti et al. 1989), one group being labelled with 111 kBq l-[^3H]leucine, the other with 37 kBq l-[U-14C]leucine (Amersham International, Amersham, Bucks, UK). After washing with a cold medium containing 5 mM non-labelled leucine to eliminate free radioactivity (not incorporated into proteins), [3H]leucine- and [14C]leucine-labelled samples were mixed in equal amounts of non-labelled proteins, and determined by total protein assay before protein separation by coelectrophoresis.

Protein separation by polyacrylamide gel electrophoresis

Samples of islets were dissolved in electrophoresis buffer (0·05 M Tris, 4% sodium-dodecyl-sulphate (SDS-Page), 12% glycerol, 15 mM dithiothreitol, 0·01% Bromophenol blue from Bio-Rad, Richmond, CA, USA), homogenised with an ultrasonic probe (Sonics & Materials Inc., Danbury, CT, USA), and heated at 100 °C for 1 min. Aliquots containing 10 µg protein were applied onto 0·3% SDS polyacrylamide gel in 3 M Tris (pH 8·45). Electrophoretic runs (Laemmli 1970) were performed using 5 to 6 h of isoelectric focusing (40 mA constant amperage).

Gel protein staining by Coomassie blue

Gels were stained in a fixative solution of 10% acetic acid, 4% formaldehyde containing 50% methanol and 0·1% Coomassie blue R-250 (Sigma, Aldrich Chimie, St Quentin Fallavier, France) for 1 h and revealed by a destaining medium containing 30% methanol and 7·5% acetic acid applied for 24 h with shaking. Gels were calibrated with molecular weight markers; standards ranged from 6·5 to 200 kDa (Bio-Rad). The different fractions were analysed by optical densitometry.

Detection of labelled proteins

Gel lanes were sliced into 2-mm fractions and dissolved in 50% H2O2 (Prolabo, France). The [3H] and [14C] activities were measured in a liquid scintillator (Emulsifier Safe, Packard, Rungis, France) using a β spectrometer (Packard-Tricarb, Rungis, France). Both [3H] and [14C] radioactivities could be measured on the same slice, thus proteins of the same molecular weight from the two groups of islets could be compared.

Presentation of the results from double labelling experiments

The d.p.m. measured from the [3H]leucine-labelled or [14C]leucine-labelled material within islets can vary from one experiment to another one. Thus, the electrophoretic patterns are qualitative, and cannot be quantitatively compared. However the d.p.m. [3H]/d.p.m. [14C] ratio of values measured together from islets of different groups, run in coelectrophoresis, allow a rigorous analysis of the variations in the amounts of newly synthesised proteins.

Results

Electrophoresis of total islet proteins

Groups of 100 islets from each group of rats were isolated, dried, and replaced in a minimal distilled water aliquot for sonication with an ultrasonic probe. As previously described (Bourlon et al. 1999) the total protein content of islets was not statistically different between the various groups of rats. In any case, in order to avoid the interference of any such variations, the protein content of islets groups was measured using the Bio-Rad method in order to obtain equal amounts of total proteins within each well of the electrophoresis gel. After electrophoretic migration, proteins were revealed by Commassie blue and their molecular weights determined using standard markers as shown in Fig. 1 (St). The relative distribution of proteins in each lane was compared between islets from vitamin D3-deficient rats (−D) and islets from normal rats (N) (Fig. 1). This showed that −D islets proteins presented a significant deficit in most of the high molecular mass proteins over 15 kDa (versus N), except around 66 kDa. The intense staining observed at 15 kDa in N islets was also observed in −D islets, whereas the lower molecular weight proteins were more intensely stained in −D islets particularly between 12 and 7 kDa. In vivo repletion of 1,25(OH)2D3, as compared with the −D lane, tended to reverse these effects of vitamin D3 deficiency; the effect was seen as soon as the 1st day of treatment (+D1) and was more pronounced after 7 days (+D7). It enhanced the staining of most of the high molecular weight proteins over 15 kDa, did not change the staining of proteins around 15 kDa and decreased the staining of the bands around 66 and 12 to 7 kDa.

Newly synthesised proteins within islets: validation of the double labelling method in normal islets

In the present experiments, the method of coelectrophoresis which was previously applied to skeletal muscle
cells (Drittanti et al. 1989) was adapted to isolated islets of Langerhans. Two groups of 100 islets from normal rats were incubated for 2 h in the presence of basal 4·2 mM glucose and an amino acid mixture in which leucine was either $^3$H or $^{14}$C labelled, for incorporation into newly synthesised proteins. After washing and sonicating, $^3$H and $^{14}$C labelled samples, mixed in equal amounts of non-labelled protein content, were run in coelectrophoresis, thus generating rigorously identical experimental conditions. Analysis of radioactivity as a function of electrophoretic mobility (see Fig. 2A and B) showed that both groups of islets from normal rats, either labelled with $^3$H or $^{14}$C, presented the same electrophoretic pattern, with maximum leucine incorporation into proteins for which the molecular mass was around 60 kDa, 48 kDa and 24 kDa. Figure 2C presents the d.p.m. $^3$H/d.p.m. $^{14}$C ratio which showed a rather constant value with slight variations included between the two dotted lines of the Fig. 2C. These limits obtained for islets from normal rats were used as a reference for experimental variations on all the following figures presenting ratio studies.

**Influence of glucose stimulation on the incorporation of radiolabelled leucine into islets**

Two groups of 100 islets from normal rats were incubated for 2 h either in the presence of basal 4·2 mM glucose and $[^{14}$C]leucine labelling or in the presence of 16·7 mM glucose (providing a strong specific stimulation of biosynthesis) in the presence of $[^3$H]leucine labelling. As described above, equal protein samples were run in coelectrophoresis. Figure 3A showed the same variations in the d.p.m. patterns induced by glucose stimulation especially around 9 kDa and over 66 kDa. Analysis of the d.p.m. $^3$H/d.p.m. $^{14}$C ratio in Fig. 3B showed more precisely that glucose stimulation induced increases in several peaks of the electrophoresis pattern over the reference limits (dotted lines). This graph shows that glucose stimulation increased the neo-synthesis of some
3H labelled protein as compared with 14C labelled proteins of islets in basal conditions. These increments corresponded mainly to 150 kDa, 130 kDa, and 8.5 kDa, and to a lesser degree to 75 kDa, 45 kDa, 30 kDa, 22 kDa, 10 kDa and 5.5 kDa. The 8.5 kDa molecular mass species may correspond to proinsulin-like material.

Influence of vitamin D₃ deficiency on newly synthesised islet proteins

One group of 100 islets from normal rats (N) was incubated for 2 h in the presence of a 16.7 mM glucose stimulus and [3H]leucine. Another group of 100 islets from 4-week vitamin D₃-deficient rats (D) was also incubated for 2 h in the presence of 16.7 mM glucose but with [14C]leucine. Both protein samples, mixed equally as previously described, were run in coelectrophoresis. Examination of the d.p.m. ³H/d.p.m. ¹⁴C ratio in Fig. 4 showed a larger neo-synthetic activity in N islets than within D islets since many peaks appeared over the dotted reference lines. Thus vitamin D₃ deficiency was observed to impair the neo-synthesis of many islet proteins: those of 200 to 113 kDa, 92 kDa, 65 to 60 kDa, 36 kDa, 28 kDa, 17 kDa, 12 kDa, 9 to 8 kDa and 5.5 kDa molecular mass. Some of these impairments corresponded to the proteins which are highly solicited during a glucose stimulus, particularly 150 to 130 kDa and 9 to 8 kDa, as shown by the comparison of Figs 3 and 4 (note the very different scales). The higher peaks, representing the larger differences between N and D groups and thus the larger deficit in neo-synthesis, were around 200 kDa, 65 kDa and 8.5 kDa.

It was only for the low molecular mass range around 7 kDa that the experimental ratio curve was below the reference line, corresponding to an increment of ¹⁴C labelled material (D) as compared with the ³H labelled normal material (N). These data suggested an increase in de novo biosynthesis of the corresponding small proteins in islets from vitamin D₃-deficient rats as compared with normal rats.

Time-course of the effect of 1,25(OH)₂D₃ in vitro on islets from vitamin D₃-deficient rats

Three different periods of 1,25(OH)₂D₃ induction were studied (2 h, 4 h, 6 h) in three separate experiments. Groups of 100 islets from vitamin D₃-deficient rats
received either $10^{-8}$ M 1,25(OH)$_2$D$_3$ and $[^3]$Hleucine, or $10^{-12}$ M 1,25(OH)$_2$D$_3$ as control with $[^14]$Cleucine added directly to the incubation medium. Then, equivalent amounts of islet proteins were treated as previously described before coelectrophoresis. The 2-h induction study was performed in the presence of 16·7 mM glucose, 0·5 mM calcium and labelled leucine; the 4-h and 6-h induction studies were performed in the presence of 8·3 mM glucose for the first hours, followed by a 16·7 mM glucose stimulation for the last 2 h for the incorporation of labelled leucine as above.

The main observation as shown in Fig. 5 was that the influence of $10^{-8}$ M 1,25(OH)$_2$D$_3$ induction was gradual. Indeed, no influence of 1,25(OH)$_2$D$_3$ could be seen at 2 h since the radioactivity ratios stayed within the reference limits (Fig. 5A). Some newly synthesised proteins appeared after a 4-h induction with $10^{-8}$ M 1,25(OH)$_2$D$_3$ as shown by the d.p.m.$[^3]$H/d.p.m.$[^14]$C peaks over the reference (Fig. 5B) for 60 to 65 kDa, 55 kDa and 9 to 8 kDa molecular masses. On the other hand $10^{-8}$ M 1,25(OH)$_2$D$_3$ decreased the neo-synthesis of 7 kDa proteins, the ratio of which dropped below the reference curve. After 6 h of $10^{-8}$ M 1,25(OH)$_2$D$_3$ induction the activation of protein neo-synthesis was more pronounced and involved more numerous proteins. Indeed the d.p.m.$[^3]$H/d.p.m.$[^14]$C ratio (Fig. 5C) showed many peaks appearing over the reference curve: 120 to 110 kDa, 92 kDa, 70 kDa, 60 to 56 kDa, 40 kDa, 35 kDa, 27 kDa to 25 kDa, 22 kDa, 15 kDa, 12 to 8 kDa and 5·5 kDa molecular mass. The radioactivity ratio was below the reference level for 7 to 6·5 kDa molecular mass species suggesting a negative regulatory effect of $10^{-8}$ M 1,25(OH)$_2$D$_3$ on the biosynthesis of these small molecular mass proteins (which are increased during vitamin D$_3$ deficiency). This effect was more pronounced at 6 h than at 4 h. A control experiment was performed with $10^{-8}$ M 1,25(OH)$_2$D$_3$ for 4 h in the presence or not of 5·$10^{-4}$ M cycloheximide (Fig. 6). This protein synthesis inhibitor almost completely suppressed the incorporation of labelled leucine into the islets, thus confirming that 1,25(OH)$_2$D$_3$ influenced the neo-synthesis of proteins induced during a glucose stimulus.

Discussion

There is evidence that in the islets of Langerhans which contain 1,25(OH)$_2$D$_3$ receptors, 1,25(OH)$_2$D$_3$ may act, at least in part, as a steroid via a nuclear mechanism rendering the β cell more competent as concerns its insulin response to glucose. Indeed, in previous studies we found that the beneficial influence of 1,25(OH)$_2$D$_3$ on insulin release is only seen when the β cells are stimulated and not in basal conditions. It is observable only after 6 h of induction (Billaudel et al. 1990) and it cannot occur in the presence of cycloheximide, a transcriptional inhibitor (Bourlon et al. 1999). Moreover, in this recent study, we show that the total islets proteins is not statistically modified by vitamin D$_3$ deficiency or by 1,25(OH)$_2$D$_3$, whereas the amount of newly synthesised labelled proteins during a glucose stimulation exhibits some variation. The amount of tritiated tyrosine incorporated into the total islet proteins during a glucose stimulation is decreased during vitamin...
D₃ deficiency and can be re-activated by 1,25(OH)₂D₃ induction (Bourlon et al. 1999). The present study (Fig. 3) showed the numerous proteins required for insulin synthesis, maturation, storage and/or exocytosis implicated in β cell stimulation-secretion coupling by glucose. The strong specific stimulation exerted by 16·7 mM glucose increased the neo-synthesis of several proteins in islets in agreement with similar findings using insulin secretory granules (Guest et al. 1991): this involved two groups of proteins over 98 kDa and a group of proteins with a molecular mass around 7 kDa. The latter method with N and −D labelled islets in a coelectrophoresis excluded any artefact of an experimental degradation of heavy proteins into smaller fragments.

The in vivo administration of 1,25(OH)₂D₃ tended to reverse the influence of 4 weeks of vitamin D₃ deficiency on the relative distribution between heavy and low proteins, increasing the amount of heavier molecular weight proteins and lowering that of small molecular weight proteins, as revealed by Coomassie blue. Similar to this static observation, the kinetic study of the in vitro influence of 1,25(OH)₂D₃ on labelled islet proteins, which were thus newly synthesised during a glucose stimulation, demonstrated that this effect was progressive. It was detectable in vitro after either a 4 h or a 6 h induction period, but not as early as 2 h. Indeed, during the first 2 h of 1,25(OH)₂D₃ induction, the neo-synthesis of islet proteins was either not affected by the steroid, or was not detectable. However, this observation does not exclude rapid or intermediate effects of 1,25(OH)₂D₃ that may be likely to involve both membrane-initiated rapid actions and transcriptional effects on early genes that do not require the nuclear receptor, such as in osteoblasts (Farach Carson & Ridall 1998) or in islets (Billault et al. 1995, 1997).

During the later periods of 1,25(OH)₂D₃ induction, the steroid progressively increased the de novo synthesis of numerous islet proteins, some of these being already activated at 4 h (60 to 65 kDa, 55 kDa, 9 to 8 kDa). Among the numerous proteins whose synthesis is activated by 1,25(OH)₂D₃ in the islets of Langerhans, some of them (presenting an equivalent molecular weight) were also found to be increased by 1,25(OH)₂D₃ in skeletal muscle cells: a glycoprotein of 55 kDa molecular mass and several calcium binding components of 100, 40, 17 and 9 kDa (Drittanti et al. 1989); others such as those of 17, 20, 30, 38, 89 kDa were also found to be PKC substrates in islets (Howell 1994). Among the proteins whose synthesis is increased by the glucose stimulus some of them such as those with molecular masses of 150 to 130 kDa and 8·5 kDa were observed to be decreased during vitamin D₃ deficiency and re-activated by 1,25(OH)₂D₃. Thus, these proteins may play a crucial role in the events implicated in transcription or transduction or during the process of insulin exocytosis. The 1,25(OH)₂D₃-induced neo-synthesis of 8·5 kDa proteins may be proinsulin-like materials, but the experimental conditions used during the electrophoresis process did not allow the detection of insulin (6 kDa). In fact, the denaturing and reducing conditions are not adapted for proteins containing disulphide bridges, such as insulin, since the two A and B chains of insulin may be separated. However, in a previous column chromatography study we showed that protein migration on gel electrophoresis, decreasing the amount of most islet proteins, especially the heaviest proteins and enhancing certain low molecular weight proteins particularly around 7 kDa. The latter method with N and −D labelled islets in a coelectrophoresis excluded any artefact of an experimental degradation of heavy proteins into smaller fragments.
1,25(OH)₂D₃ increases both the amount of newly synthesised insulin and proinsulin-like materials, accelerating more particularly the neo-conversion of proinsulin into insulin when the β cell is highly solicited by glucose (Bourlon et al. 1999).

In conclusion, the present data lend support to the hypothesis of genomic effects of 1,25(OH)₂D₃ on islets from vitamin D₃-deficient rats, in agreement with observations on the 1,25(OH)₂D₃-induced increase in proinsulin mRNA found by other authors (Ozono et al. 1990), but this is the first time that these beneficial effects were shown to occur gradually, and on numerous neo-synthesised proteins which may prepare the β cells for an enhanced insulin response to glucose.

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