Beneficial effect of 1,25 dihydroxyvitamin D₃ on cytokine-treated human pancreatic islets

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Abstract

We examined whether 1,25 dihydroxyvitamin D₃ (1,25 D₃), the active form of vitamin D involved in the regulation of the immune system, may also protect human pancreatic islet cells from destruction induced by cytokines. In this study, we specifically investigated the effect of 1,25 D₃ on oxidative stress and major histocompatibility complex (MHC) induction, both implicated in cytokine-induced islet cell dysfunction and destruction. We also investigated the effects of 1,25 D₃ on interleukin (IL)-6, a pleiotropic cytokine implicated in the pathogenesis of immunoinflammatory disorders.

Human pancreatic islets, isolated from heart-beating donors, were treated with a combination of three cytokines, IL-1β+tumor necrosis factor α+interferon γ, in the presence or absence of vitamin D, and compared with untreated control cells. Metabolic activity was assessed by cell viability and insulin content. Oxidative stress was estimated by heat shock protein 70 (hsp70) expression, cell manganese superoxide dismutase (MnSOD) activity and nitrite release, a reflection of nitric oxide (NO) synthesis. Variation of immunogenicity of islet preparations was determined by analysis of the MHC class I and class II transcripts. Inflammatory status was evaluated by IL-6 production. After 48 h of contact with cytokines, insulin content was significantly decreased by 40% but cell viability was not altered. MHC expression significantly increased six- to sevenfold as well as NO and IL-6 release (two- to threefold enhancement). MnSOD activity was not significantly induced and hsp70 expression was not affected by the combination of cytokines.

The addition of 1,25 D₃ significantly reduced nitrite release, IL-6 production and MHC class I expression which then became not significantly different from controls. These results suggest that the effect of 1,25 D₃ in human pancreatic islets cells may be a reduction of the vulnerability of cells to cytotoxic T lymphocytes and a reduction of cytotoxic challenge. Hence, 1,25 D₃ might play a role in the prevention of type 1 diabetes and islet allograft rejection.

Introduction

Type 1 diabetes is a multifactorial autoimmune disease in which insulin-producing beta cells are selectively destroyed. Impaired function and destruction of beta cells may result from direct contact with islet-infiltrating macrophages and T lymphocytes and/or exposure to inflammatory products of the islet-infiltrating cells, such as free radicals and cytokines (Mandrup-Poulsen et al. 1990). The inflammatory cytokines, interleukin (IL)-1β, tumor necrosis factor α (TNFα) and interferon γ (IFNγ), acting individually or more potently in combination, are cytotoxic to human islet beta cells *in vitro* by stimulating cytotoxic mediators such as other inflammatory cytokines, nitric oxide (NO), superoxide anion or major histocompatibility complex (MHC) (Iwahashi et al. 1998, Rabinovitch & Suarez-Pinzon 1998). This combination of cytokines may also activate pathways of gene regulation leading to induction of protective compounds such as stress proteins and antioxidant enzymes (Eizirik 1996). The modulation of such inductions may prove to be beneficial for beta cells and 1,25 dihydroxyvitamin D₃ (1,25 D₃), the active metabolite of vitamin D₃, may be a candidate able to fulfill some of these criteria.

Indeed, 1,25 D₃ has been shown to prevent autoimmune disease such as experimental encephalomyelitis (Lemire & Archer 1991), autoimmune thyroiditis (Fournier et al. 1990) or experimental rat nephritis (Hattori 1990). Other experiments carried out in the non-obese diabetic (NOD) mouse, a model for type 1 diabetes,
demonstrated that 1,25 D₃ also prevented insulitis and diabetes (Mathieu et al. 1992, 1994, Casteels et al. 1998). In fact, the incidence of insulitis was reduced by 1,25 D₃, a known powerful immunosuppressive compound which inhibits lymphocyte proliferation and cytokine production in vitro including IL-6 secretion (Müller et al. 1991, Lemire et al. 1995).

On the other hand, pancreatic beta cells taken from newly diagnosed patients with type 1 diabetes have been shown to express MHC class II molecules in addition to NOD mice (Bottazzo et al. 1987) and a previous report from Hahn et al. (1997) showed that, in islet cells of bio-breeding (BB) rats, MHC antigen expression stimulated by IFNγ was reduced by 1,25 D₃ and an analog ZXY 1106.

Besides immunomodulatory properties, 1,25 D₃ might also exhibit beta cell-protective properties since, in rat pancreatic islet cells, Sandler et al. (1994) demonstrated that 1,25 D₃ and some of its structural analogs counteracted the suppressive effect of IL-1β on accumulated insulin release.

No demonstration of the potential protective action of 1,25 D₃ in human pancreatic islet cells has been achieved at this time. We therefore studied the effect of 1,25 D₃ on cytokine-induced damage to human pancreatic islets and particularly the effect on oxidative stress, IL-6 and MHC induction.

**Materials and Methods**

**Human islet processing**

Human pancreases (mean age 36 ± 15 years, n = 10) were harvested from adult brain-dead donors in accordance with French regulations and with the local Institutional Ethical Committee. Pancreatic islets were isolated after ductal distension of the pancreas and digestion of the tissue with Liberase (Roche Diagnostics, Meylan, France) according to the automated method of Ricordi et al. (1988) with modifications (Kerr-Conte et al. 1994). Semi-purification was achieved with Euro–Ficoll discontinuous density gradient using a cell separator (Lake et al. 1989). Islet number was determined on samples of each preparation after dithizone staining and expressed as equivalent number of islets (IE) (Ricordi et al. 1990). Preparations used in this study exhibited an 85 ± 7% purity and an average yield of 2150 ± 545 IE/g pancreas. Semi-purified islets were cultured for 24 h in CMRL 1066 medium (Gibco BRL, Life Technologies, Cergy-Pontoise, France) with 2% Ultroser G (Gibco BRL), then processed in batches of 500 IE in 4-well multilidish plates (Nunc; Gibco BRL) or 40 IE in 96-well multiscreen plates (Millipore, Saint Quentin Yvelines, France) (Vandewalle et al. 1999).

Islet processing consisted of a 48-h treatment at 37°C in different media, including control basal medium (Ham’s F10 containing 0-5% bovine serum albumin), cytokine-treated medium (basal medium with IL-1β (50 IU/ml)+TNFα (1000 IU/ml)+IFNγ (1000 IU/ml); cytokines from Biosource, Clinisciences, le Pont de Claiix, France) and cytokine-treated medium containing 1,25 D₃ dissolved in ethanol (10⁻⁸ and 10⁻⁶ mol/l, final concentrations). Alcohol concentration did not exceed 0-01% in the culture media and was without measurable effect on cell cultures. 1,25 D₃ was a generous gift from Hoffmann la Roche AG, Basel, Switzerland.

**Insulin content**

Islets were pelleted by centrifugation and homogenized by sonication (20 kHz, 30 W) for 15 s in 200 µl Tris–HCl, 1 mmol/l EDTA, pH 7-4 and stored frozen at −20°C.

The islet insulin content was measured by radioimmunoassay (CIS bio International, Gif-sur Yvette, France) after overnight extraction at 4°C of 50 µl aliquots of homogenates with 125 µl acid–ethanol (0-18 mol/l HCl in 95% ethanol).

**Estimation of islet cell viability**

We used the Alamar blue reagent (Alamar Blue TM; Biosource) which contains a non-toxic REDOX indicator. Reduction related to metabolic activity causes the REDOX indicator to change from an oxidized to a reduced fluorescent form (Ahmed et al. 1994). Alamar blue was added directly (10% v/v) to islet cells cultured in multiscreeen wells containing 40 IE per condition in 200 µl culture medium and incubated for 48 h and 144 h at 37°C in an atmosphere of air:CO₂ (95:5). After transfer of the reaction mixture to a 96-well microtiter plate, the fluorescence was read at 544 nm (excitation wavelength) and 590 nm (emission wavelength) (Vandewalle et al. 1999).

**Nitrite determination**

The nitrite assays were performed in the culture medium from each well as previously described (Vandewalle et al. 1999). This assay was adapted from the study of Misko et al. (1993) where nitrite determination is based upon the reaction of nitrite with 2,3-diaminonaphthalene (Sigma-Aldrich Chemicals, Saint Quentin Fallavier, France) to form the fluorescent product 1-(H)-naphtotriazole.

**Manganese superoxide dismutase activity**

To measure superoxide dismutase (SOD) activity, batches of 500 IE were homogenized by sonication (20 kHz, 30 W) for 10 s in 250 µl phosphate buffer, pH 7-8 (1/15 mol/l Na₂PO₄·KH₂PO₄·0-1 mmol/l EDTA). The activity of SOD was estimated as described by the manufacturer (Randox Laboratories Ltd, Crumlin, Co. Antrim, UK). In this method superoxide radicals, generated by the action of xanthine oxidase on xanthine, reacted with
Extraction of RNA

Total cellular RNAs were obtained by lysis of 500 IE per condition in 1 ml RNAzol® (Bioprobe Systems, Montreuil, France) and quantified spectrophotometrically (260 nm). The cDNAs were synthesized from 2 µg DNA-free total RNA with 1000 ng oligo(dT) 12–18 primer and 4 U RNAsin (Gibco BRL) first incubated for 10 min at 70 °C. Reverse transcription was then performed at 42 °C for 1 h by the addition of 200 U reverse transcriptase (M–MLV; Life Technologies) plus 2 mmol/l of each dNTP (Pharmacia, Saint Quentin Yvelines, France), 5 mmol/l potassium cyanide (Flohé & Otting 1984). Products were further incubated for 1 h at 42 °C using 100 U reverse transcriptase then the final reaction mixture was heated at 95 °C for 5 min and stored at −20 °C until use.

Heat shock protein 70 analysis by quantitative RT-PCR

Heat shock protein 70 (hsp70) mRNA was monitored in 96-well plates by quantitative RT-PCR of the target fragment and an 18S housekeeping gene sequence as an internal standard. Amplification was achieved with specific primers of the target sequence hsp70 (5′-GACCTT GACAAACAGGCTGTTGA-3′) and (5′-ACAGGGTC CTCTTGCCGCCC-3′) and 18S (5′-TCCAGTGTTGC GGATTCA-3′) and (5′-GATCCAGGGCCCTCA AAC-3′) (GENSET SA, Paris, France), resulting in 129 bp and 104 bp products respectively.

The template concentration per reaction represented one-tenth of the cDNA reaction performed on 2 µg total RNA. The amplification was achieved in 25 µl reaction mixture containing 3 µl cDNA, 15 pmol of each oligonucleotide primer and 2 × Sybr Green Master Mix (4309155; Perkin-Elmer Biosystems, Courtaboeuf, France).

After denaturation for 10 min at 95 °C, 40 cycles of amplification were performed in an ABI Prism 7700 Sequence Detection System (Perkin-Elmer Biosystems), amplification parameters included 30-s denaturation at 94 °C and 1-min annealing step at 60 °C. Direct detection of PCR products was monitored by measuring the increase in fluorescence caused by the binding of SYBR Green to double-stranded DNA; all quantitations were achieved with the comparative C_T method and normalized to the endogenous control 18S rRNA as described by the manufacturer (Perkin-Elmer Biosystems).

Analysis of MHC I and MHC II transcripts by semi-quantitative RT-PCR

Semi-quantitative, non-competitive RT-PCR was performed with AmpliTaqGold™ (Perkin-Elmer) which allowed simultaneous co-amplification of the housekeeping gene (β-actin) and each of our target genes.

Primer sets (Perkin-Elmer Biosystems) included the β-actin primers (position 405–722): sense 5′-ATCATGT TTGAGACCTCCTAA-3′, antisense 5′-CATCTCTTGTGC TCAGAAGTCCA-3′ resulting in a 314 bp product; MHC I primers (locus A.B.C): sense 5′-CTCCCCAGACG CCGAGA-3′, antisense 5′-GGGCTGCAGCTCCTC TT-3′ resulting in a 632 bp product; MHC II primers (locus DR β): sense (position 508–528 in exon 1) 5′-TC TCCAGCATGGTGTTCTGA-3′, antisense (position 1146–1166 in exon 2) 5′-CTCCGCCGCTGCAGTGTG AAGC-3′ resulting in a 390 bp product. PCR was performed with 1 µl cDNA from each sample in the presence of 200 µmol/l dNTP, 1.5 µmol/l MgCl2, 5 U AmpliTaq DNA polymerase and 25 pmol/l of each couple of primers. PCR (32 cycles) was carried out in a thermal cycler (2400; Perkin-Elmer Biosystems) with each cycle consisting of denaturation at 94 °C for 30 s, annealing at 54 °C for 1 min and polymerization at 75 °C for 30 s. The last PCR step was a final extension at 75 °C for 5 min. The PCR products were electrophoresed in 2% agarose gel and bands were scanned with an integration camera CDD (COHU 4912; Clara Vision, Orsay, France) and analyzed with GelAnalysts® 3-01 FR software (© GreyStone-Iconix; MEB Electronique, Paris, France). Band intensity was expressed in arbitrary units and MHC I or MHC II mRNA expression indexed to β-actin mRNA expression.

IL-6 determination

Concentrations of IL-6 released in culture medium were determined with commercial enzyme-linked immunosorbent assay kits (Immunotech, Marseille, France). Assays were performed as described by the manufacturer using monoclonal antibodies and acetylcholinesterase as the tracer enzyme (Brailly et al. 1994).

Presentation of results

Data analyses were performed using Statview (Abacus Concepts Inc., Berkeley, CA, USA). Given protocols were tested in islet cells from five to ten preparations from different donors. The statistical significance between means was assessed by Student’s t-test for paired values.

Results

After 48 h of contact of human islets with cytokines, insulin content was significantly decreased (Fig. 1), cell
viability was not altered at that time but became significantly lower than the control after 6-day exposure (Fig. 2). In both cases, 1,25 D3 addition resulted in a tendency to revert values to those of controls.

The effect of 1,25 D3 on the oxidative stress induced by cytokines was first estimated by the measurement of nitrite accumulation in the culture medium. Cytokines significantly enhanced nitrite levels (two- to threefold increase). The addition of 1,25 D3 resulted in a significant reduction of values that were no longer significantly different from controls (Fig. 3).

We also observed a stimulation of MnSOD by cytokines that did not, however, reach statistical significance (Fig. 4), while the expression of hsp70 was not modified at all by cytokines (data not shown). The addition of 1,25 D3 did not induce further significant modifications in these two latter results.

Figure 1 The insulin content of 48-h treated cells was assessed by immunoassay after extraction as described in Materials and Methods. Values are expressed as percentage of control cells (2.7 ng/IE). Data are means ± s.e.m. (n = 7). *P < 0.05 (Student’s t-test for paired values).

Figure 2 The metabolic activity of 48-h (solid bars) and 6-day (open bars)-treated cells was assessed using the REDOX indicator Alamar blue as described in Materials and Methods. Values are expressed as percentage of control cells. Data are means ± s.e.m. (n = 6). *P < 0.05 (Student’s t-test for paired values).

Figure 3 Nitrite released by cells in the culture medium during 48 h of treatment was assessed by fluorometric assay as described in Materials and Methods. Values are expressed as percentage of control cells (1.7 pmol NaNO2/IE). Data are means ± s.e.m. (n = 7). **P < 0.01 (Student’s t-test for paired values).

Figure 4 MnSOD activity in 48-h treated cells was assessed by a photometric assay as described in Materials and Methods. Values are expressed as percentage of control cells (74 µIU/IE). Data are means ± s.e.m. (n = 5).
Analysis of MHC expression in cytokine-treated islet cells highlighted a strong stimulation of both class I and II complexes (six- to sevenfold increase); 1,25 D3 significantly decreased MHC class I but not class II expression (Fig. 5).

Finally, cytokines significantly enhanced IL-6 release (three- to fourfold increase) and a significant reduction of values by 1,25 D3 was noticed (Fig. 6).

Discussion

In this study we have tried to assess, in human pancreatic islet cells, the effect of 1,25 D3 on cytokine-mediated inhibition of beta cell function in association with the expression of MHC antigens and the production of oxygen species. Indeed, important factors in cytokine-induced beta cell death may be NO-mediated cell damage and the toxic effects of oxygen-free radicals. Other factors are cytokine-stimulated class I and/or class II molecules which, when associated with antigens and recognized by T-Helper cells, enhance the targeting of cytotoxic T-lymphocytes against islet cells. In addition, in pancreatic beta cells, as in a variety of cell types including fibroblasts, endothelial cells and monocytes, IL-1β and TNFα also stimulate the production of other cytokines such as IL-6 which may act as a co-stimulator for autoreactive B and T lymphocytes in autoimmune diabetes (Campbell et al. 1989).

Simultaneously, potentially important defence and repair responses such as formation of heat shock proteins and superoxide dismutase are induced by cytokines. Human pancreatic islet cells have been shown to possess more defence molecules than murine islet cells (Welsh et al. 1995, Eizirik 1996) so we used a combination of three cytokines at the concentrations known to be required to induce damage in human islet cells (Rabinovitch et al. 1990, Eizirik et al. 1994).

We noticed, as described by others, that cytokines induce a significant decrease in insulin content. The addition of 1,25 D3 showed a clear tendency to revert values to control levels. After 48 h of contact with cytokines, the cell viability was not affected. Longer time-periods (5–6 days) have been reported to be necessary for the destruction of human islet cells (Eizirik et al. 1994, Delanay et al. 1997). In our experiments, a significant reduction in cell viability was also observed after 6 days of exposure; the addition of 1,25 D3 also reverted values to those of the control.

NO, induced by cytokines and assessed by nitrite accumulation in the culture medium, was significantly enhanced after 48 h of culture compared with control values. The addition of 1,25 D3 significantly reduced nitrite levels. NO is known to mediate physiological
processes via a reaction with superoxide or by complexing with a metal (e.g. iron/sulfur complexes) at the active sites of enzymes. In pancreatic beta cells, these NO-induced complexes result in an inhibitory effect on the Krebs cycle and, particularly, in an inhibition of the activity of aconitase, an enzyme which converts citrate to isocitrate, thus leading to a severe decrease in glucose metabolism and ATP production (Welsh et al. 1991). IL-1β-induced NO production has been reported to be mediated by the nuclear transcription factor NF-κB (Flodstrom et al. 1996). The action of 1,25 D3 on cytokine-induced NO production might lie at the level of NF-κB activation, since, in other systems, a modulation of NF-κB protein levels by 1,25 D3 has been reported (Yu et al. 1995, Larsen et al. 1998). Other transcription factors of potential relevance for inducible nitric oxide synthase (iNOS) activation are c-Jun N-terminal kinase (Bonny et al. 2000) and activator protein-1 (Xie et al. 1993) which might also represent potential targets for 1,25 D3.

Cytokine-induced iNOS mRNA expression and NO production is paralleled by increased MnSOD expression (Bigdeli et al. 1994) and NF-κB has also been shown to be required for cytokine-induced MnSOD expression in insulin-producing cells (Darville et al. 2000). A combination of three cytokines was also reported to be necessary to induce MnSOD protein expression in human pancreatic islets (Welsh et al. 1995). In our study, islets exposed to cytokines for 48 h showed a mean 2.8-fold greater activity of MnSOD than in control islets; however, this was not significant (P = 0.08) due to the great dispersion of values; moreover, the addition of 1,25 D3 did not significantly modify this enhancement. We concluded that 1,25 D3 prevents the induction of NO by cytokines in human islets, without affecting cytokine-induced MnSOD activity.

We also found that the combination of cytokines did not significantly increase the hsp70 expression. The high level of human hsp70 in basal conditions may explain this absence of a stimulatory effect of cytokines (Welsh et al. 1995, Eizirik 1996); in addition, 1,25 D3 did not modify hsp70 expression. However, an emerging body of evidence suggests that hsp70 participates in cytokine signal transduction (Moseley 1998) and that its protective effect on islet metabolic function may be associated with an inhibition of IL-1β-stimulated iNOS expression (Scarim et al. 1998). In our experiments, such a mechanism was not likely to be effective since hsp70 expression was unrelated to NO modifications.

In the pancreatic islets of rodents, BB rats and humans with autoimmune diabetes, over-expression of class I MHC (Allison et al. 1988, Kay et al. 1991) is observed. Class I MHC expression is up-regulated by cytokines, and especially INF-γ, and is induced in beta cells during destructive insulinitis (Campbell et al. 1986, Lafferty 1997, Thomas et al. 1998). INF-γ also induces MHC class II expression in many cell types, including fibroblasts and endothelial cells (Pujol-Borell et al. 1987). In human pancreatic islet cells, MHC II has been demonstrated in 5–10% of ductal cells; this percentage increases in the presence of INF-γ (Pavlovic et al. 1997). No expression, however, has been found in human endocrine beta and non-beta cells and conflicting results regarding the possible induction by INF-γ have been reported (Campbell et al. 1986, Pujol-Borell et al. 1987, Pavlovic et al. 1997).

In cytokine-stimulated human pancreatic islet cells, we demonstrated a significant increase in both MHC I and MHC II expression and a significant inhibition of MHC I induction by 1,25 D3 contrary to MHC II expression which remained statistically unchanged by 1,25 D3. Hyperexpression of MHC molecules, in response to certain cytokines, could be a primary event leading directly to beta cell destruction (Okamoto 1999) and suppression of MHC class I antigen hyperexpression has been shown to prevent insulitis and diabetes (Mashima et al. 1999). The counteracting effect of 1,25 D3 on MHC induction has also been described by Hahn et al. (1997) in rat pancreatic islet cells; nevertheless, the mechanisms implicated in these processes remain to be established.

The toxic effects of cytokines such as IL-1β, TNFα and INF-γ may also be linked to the production of other cytokines such as IL-6. Indeed, this inflammatory cytokine, which is mainly secreted by monocytes, may also be produced by isolated mouse islets or rat insulinoma cells after exposure to INF-γ and TNFα (Müller et al. 1991). This IL-6 production is of pathogenic importance since in NOD mice, animal models of insulin-dependent diabetes mellitus, the administration of antibodies directed against IL-6 reduces the incidence of diabetes (Campbell et al. 1991). In human islets, we found a significant threefold increase of IL-6 release after 48 h of contact with cytokines and a significant reduction of values by the addition of 1,25 D3. Likewise, in other tissues such as mice thymocytes (Müller et al. 1991) and human fibroblast cell lines (Srivastava et al. 1994), such modifications of the effects of IL-6 by 1,25 D3 have also been described. Our results demonstrate that production of IL-6 by the islets, which may enhance the activity of local autoreactive lymphocytes and thus play a role in the development of autoimmune disease, may be efficiently counteracted by 1,25 D3.

Identification of mediators of beta cell destruction and of mechanisms involved in beta cell repair and protection may help develop strategies favoring beta cell survival in the early stages of the disease where there is a negative balance between beta cell damage and beta cell repair. The ability of immune cells to synthesize 1,25 D3 indicates a possible paracrine/autocrine mode of action for this hormone which may influence a wide range of immune functions, including modulation of cytotoxic macrophage activity and antigen presentation by MHC molecules in immune as well as in somatic cells.

Suppression of NO production or inhibition of NO activity, in endocrine as well as in ductal cells (Pavlovic
et al. 1999), is also a potential strategy to prevent islet cell dysfunction and killing and to increase the early function and engraftment of transplanted islets in the clinical setting. Indeed, pancreatic islet preparations intended for allografting contain various types of cells which may contribute to triggering beta cell destruction. Although variable based on the purity of the preparations (85 ± 7% in this study), pancreatic islets are contaminated with 10–30% of exocrine cells, other minor contaminants being passenger leucocytes (1%). Purified islet fractions consist of ductal cells (20–30%) plus endocrine cells (40–50%) (Kerr-Conte et al. 1996) including beta cells accounting for only about 25% of total cells (Shapiro et al. 2000). With the exception of insulin, the parameters which were modified by cytokines may involve all cell types, in particular MHC I which is expressed on all nucleated cells. The beneficial effect exerted by 1,25 D3 may therefore affect the whole inflammatory status of the graft and thereby be beneficial to insulin-secreting cells.

To summarize, the overall effect of 1,25 D3 in human pancreatic islet cells may be a reduction of vulnerability of cells to allogenic cytotoxic T lymphocytes and a reduction of cytoxic challenge. Hence 1,25 D3 and its analogs might become useful both in the prevention or treatment of type I diabetes as well as in the management of transplant rejection.

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