Decreased concentration of plasma leptin in periparturient dairy cows is caused by negative energy balance

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Abstract

Dairy cows suffer from an intense energy deficit at parturition due to the onset of copious milk synthesis and depressed appetite. Despite this deficit, maternal metabolism is almost completely devoted to the support of mammary metabolism. Evidence from rodents suggests that, during periods of nutritional insufficiency, a reduction in plasma leptin serves to co-ordinate energy metabolism. As an initial step to determine if leptin plays this role in periparturient dairy cows, changes in the plasma concentration of leptin were measured during the period from 35 days before to 56 days after parturition. The plasma concentration of leptin was reduced by ~50% after parturition and remained depressed during lactation despite a gradual improvement in energy balance; corresponding changes occurred in the abundance of leptin mRNA in white adipose tissue. To determine whether negative energy balance caused this reduction in circulating leptin, cows were either milked or not milked after parturition. Absence of milk removal eliminated the energy deficit of early lactation, and doubled the plasma concentration of leptin. The plasma concentration of leptin was positively correlated with plasma concentrations of insulin and glucose, and negatively correlated with plasma concentrations of growth hormone and non-esterified fatty acids. In conclusion, the energy deficit of periparturient cows causes a sustained reduction in plasma leptin. This reduction could benefit early lactating dairy cows by promoting a faster increase in feed intake and by diverting energy from non-vital functions such as reproduction.


Introduction

During the periparturient period, high yielding dairy cows experience major changes in energy metabolism (Bauman & Currie 1980, Bell 1995, Barber et al. 1997, Bauman 2000). With the onset of lactation, the added metabolic activities of the mammary gland increase the total energy requirements by approximately fourfold. A pronounced energy deficit develops because voluntary feed intake is insufficient to meet this increased energy expenditure, and is met by mobilizing lipids from white adipose tissue (WAT) (Bauman & Currie 1980, Bell 1995, Barber et al. 1997, Bauman 2000). Despite this shortfall, partitioning of nutrients to the mammary gland is favored, and represents over 70% of available energy (ingested and endogenous). The metabolic context of the nutritional insufficiency of early lactation is, however, different from that associated with fasting or undernutrition. First, increased demand, not decreased provision of nutrients, is the primary cause of the undernourished state in early lactation. Second, because the undernutrition associated with the onset of milk synthesis confers no direct benefits to the mother, lactation must call on additional hormonal and cellular adaptations to harness maternal energy metabolism (Bauman & Currie 1980, Bell 1995, Vernon & Pond 1997, Bauman 2000).

In ruminants, key adaptations of early lactation that have been identified in recent years include increased secretion of growth hormone (GH) and decreased responsiveness of skeletal muscle and WAT to insulin (Bell & Bauman 1997, Vernon & Pond 1997, Etherton & Bauman 1998). Changes in the plasma concentration of leptin, a protein hormone secreted almost exclusively by adipocytes (Zhang et al. 1994, Friedman & Halaas 1998, Ji et al. 1998, Ahima & Flier 2000), could also be an important adaptation, particularly given the role of WAT in support of early lactation in dairy cattle. In ruminants and other animals, leptin is synthesized in proportion to the overall degree of adiposity (Friedman & Halaas 1998, Ahima & Flier 2000, Blache et al. 2000, Delavaud et al. 2000, Ehrhardt et al. 2000), and acts on the central nervous system (CNS) to reduce voluntary feed intake (Friedman & Halaas 1998, Henry et al. 1999, Ahima & Flier 2000, Schwartz et al. 2000). During fasting, the plasma concentration of leptin is
reduced rapidly, concomitant with changes in neuroendocrine signals that promote fasting metabolism and the attenuation of dispensable, energy-dependent functions such as reproduction and immune response (Ahima et al. 1996, 1999, Lord et al. 1998). Because leptin therapy partly reverses these adaptations in fasted animals (Ahima et al. 1996, Finn et al. 1998, Lord et al. 1998, Nagatani et al. 2000), an equally important role of leptin is to signal the energy status of the periphery to the CNS (Flier 1998, Ahima & Flier 2000).

So far, most research on leptin has focused on its contribution to the development of obesity and diseases in humans and rodent models (Friedman & Halaas 1998, Ahima & Flier 2000). The role of leptin in regulating complex and dynamic changes in energy balance (EB), such as those taking place during the transition from pregnancy to lactation, has received only limited attention (Kawai et al. 1997, Brogan et al. 1999, Butte et al. 1999, Woodside et al. 2000); no information is available in dairy cattle, a species which suffers from a more intense energy deficit after parturition than do humans or rodents (Barber et al. 1997, Bauman 2000). As a first step towards understanding the role of leptin in dairy cattle, we have measured changes in the concentration of plasma leptin during the periparturient period. Our results indicate that the energy deficit of early lactation reduces leptin synthesis in WAT, and suggest roles for insulin, GH and metabolic fuels in mediating this effect of EB.

Materials and Methods

Animals and design

All experimental procedures were conducted with the approval of the Cornell University Institutional Animal Care and Use Committee. Multiparous Holstein cows (age, 3-7±0.8 years; parity number, 1-6±0.7) were housed in individual stalls, except around parturition when they were moved to individual maternity pens. The facility was lit between 0600 and 2300 h.

Periparturient period study

To study the transition from pregnancy to lactation, eight cows were used in the period between 35 days before and 56 days after parturition (day −35 to +56 relative to parturition). Cows were fed a total mixed ration (TMR) ad libitum once daily at 1100 h. Nutrient composition of the TMR varied with the stage of pregnancy and lactation. The TMR contained 1·45 Mcal of net energy of lactation (NE\textsubscript{L}) and 125 g crude protein (CP) per kg of dry matter (DM) between day −35 and −21, 1·63 Mcal NE\textsubscript{L} and 143 g CP per kg DM between day −21 and parturition, and 1·76 Mcal NE\textsubscript{L} and 183 g CP per kg DM during lactation (National Research Council 1989). Cows were milked thrice daily at 0900, 1600 and 2300 h.

Blood samples were obtained thrice weekly between day −28 and −9, daily between day −9 and +7, and on alternate days thereafter; they were collected between 0900 and 1000 h by coccygeal venipuncture. In addition, hourly blood samples were obtained for 26 consecutive hours on day −33±5, +3 and +56 via an indwelling jugular catheter. Plasma was prepared immediately and frozen at −20 °C until analyzed. Biopsies of subcutaneous WAT were obtained from the tailhead region (Houseknecht & Bauman 1997) at day −32±5, −7±2, +3, +21 and +56. Variation in the time of hourly blood samples or WAT biopsy during pregnancy reflects differences between predicted and actual time of parturition. Biopsies were snap frozen in liquid nitrogen and stored at −80 °C.

Energy balance study

A second study was performed to study the effects of EB in early lactation. Starting at parturition, 14 multiparous cows were either milked thrice daily (lactating) or never milked (non-lactating). From parturition until day +7 (relative to parturition), lactating cows were offered ad libitum levels of a low energy TMR (1·52 Mcal NE\textsubscript{L} and 189 g CP per kg DM). Between day +7 and +33, intake of this TMR was restricted to the amount consumed on day +7 and averaged 13-6±1·2 kg/day. During this same time-period, non-lactating cows consumed ad libitum levels of a high energy TMR (1·70 Mcal NE\textsubscript{L} and 188 g CP per kg DM; average DM intake = 13·4±4·8 kg/day). Blood samples were collected on three different days between day +5 and +11 (week +1) and between day +26 and +33 (week +4). They were obtained by coccygeal venipuncture at 0800 h and processed immediately to plasma.

Whole body energetics

Dry matter intake, milk yield and aliquots of milk for composition analysis were obtained daily during the periparturient period study. During the EB study, these data were collected identically, except that milk composition was analyzed twice weekly. Energy content of milk and feeds was estimated from chemical composition (National Research Council 1989). Every week, body weights were recorded at a standard time on 2 consecutive days. These data were used to calculate individual estimates of EB (National Research Council 1989, Beam & Butler 1997). Four percent fat-corrected milk yields were calculated according to the National Research Council (1989). To estimate changes in body fatness, two independent individuals assigned a weekly body condition (BC) score (thin=1, fat=5) to each cow as described previously (Beam & Butler 1997).

Analysis of leptin mRNA

Total RNA was isolated from biopsied WAT by the acid guanidium thiocyanate–phenol–chloroform method, and
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quantified by absorbance at 260 nm (Boisclair et al. 2000). RNA quality was assessed on formaldehyde agarose gel by staining with Sybr Green II RNA stain (Molecular Probes, Eugene, OR, USA). A ribonuclease protection assay (RPA) was used to measure simultaneously leptin and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA. Briefly, DNA fragments corresponding to nt+64 to +316 (ATG, +1) of the bovine leptin cDNA (GenBank accession #U50365), and to nt+717 to +823 of the partial bovine GAPDH cDNA (numbering relative to first nt reported in GenBank accession #A000039) were cloned into the plasmid pCR II (Invitrogen, Carlsbad, CA, USA). After linearization of each plasmid by digestion with the appropriate restriction endonuclease, leptin and GAPDH antisense RNAs were synthesized in the presence of [32P]UTP and T7 RNA polymerase using a commercial kit (Maxiscript, Ambion, Inc., Austin, TX, USA). Two micrograms of total WAT RNA were analyzed using the RPA III kit (Ambion, Inc.). Protected bands (253 bp for leptin, 106 bp for GAPDH) were resolved on 6% polyacrylamide, 7 M urea gels. Signals were quantified by phosphorimaging using a Fujix-Bio-Imaging Analyzer BAS 1000 (Fuji Medical Systems Ltd, Stanford, CT, USA).

Analysis of metabolites and hormones

Plasma glucose was measured by the glucose oxidase method and non-esterified fatty acids (NEFA) by the Acyl-CoA synthetase/oxidase method (Boisclair et al. 1994). Plasma leptin was assayed by a recently developed, double-antibody bovine radioimmunoassay (RIA) (Ehrhardt et al. 2000). The assay is based on a primary rabbit antibody raised against recombinant bovine leptin, and a secondary goat antibody raised against rabbit γ-globulin. Recombinant bovine leptin was used for iodination and standards. This RIA has a sensitivity of 0·5 ng/ml and a range of 0·5 to 20 ng/ml. Plasma concentrations of insulin, GH and insulin-like growth factor (IGF-I) were measured by specific RIAs established in our laboratory (Boisclair et al. 1994, Beam & Butler 1997). Plasma cortisol was determined by a commercially available RIA (Diagnostic Systems Laboratories, Inc, Webster, TX, USA). Inter- and intra-assay coefficients of variation for all assays averaged less than 8% and 9% respectively.

Statistical analysis

Data were analyzed by repeated measure models using the SAS statistical package (SAS Institute, Cary, NC, USA). For the periparturient period study, the mixed model accounted for time as the fixed effect and animal as the random effect. Because the most dramatic metabolic adaptations occur immediately before and after parturition, statistical analysis was performed on data compiled for week −4 (day −28 to −22), week −1 (day −8 to −2), week +1 (day +2 to +8), week +3 (day +18 to +25) and week +8 (day +50 to +57). When the effect of time was significant (P<0·05), variation was partitioned between the following pre-planned orthogonal contrasts: (1) physiological state (STATE, week −5 and −1 vs week +1, +3 and +8); (2) stage of pregnancy (PREG, week −4 vs −1); (3) stage of lactation (LACT, week +1 and +3 vs week +8); (4) early lactation (E-LACT, week +1 vs +3).

For the EB study, the mixed model accounted for treatment (lactating, non-lactating), time (week +1, +4) and their interaction as fixed effects, and animal as the random effect. The term animal (treatment) was used to test the significance of treatment, and the residual error term was used to test the significance of time and time × treatment.

Results

Periparturient period

Profiles of energy-related variables are shown in Fig. 1, and the orthogonal contrasts are reported in Table 1. Voluntary intake of dry matter and energy did not differ during pregnancy, resulting in similarly positive estimates of EB at week −4 and −1 (PREG, P>0·05; Table 1). Estimates of EB became negative at parturition with the onset of milk secretion (Fig. 1), and were lower during lactation than during pregnancy (STATE, P<0·001). The calculated energy deficit was maximal between week +1 and +3 (Fig. 1), and was eliminated by week +8 due to increased feed intake (LACT, P<0·001). Consistent with these temporal changes in EB, BC scores did not differ during pregnancy, but were lower during lactation (STATE, P<0·001). Variation was partitioned between the following fixed effects, and animal as the random effect: (1) physiological state (STATE, week −5 and −1 vs week +1, +3 and +8); (2) stage of pregnancy (PREG, week −4 vs −1); (3) stage of lactation (LACT, week +1 and +3 vs week +8); (4) early lactation (E-LACT, week +1 vs +3).

Next we measured the plasma concentration of leptin during the periparturient period (Fig. 2 and Table 2). The plasma concentration of leptin did not differ during pregnancy (PREG, P>0·05), but was reduced by ∼50% during lactation (STATE, P<0·01). Inspection of the leptin profile suggests that this reduction was initiated a few days before parturition (Fig. 2). Despite the gradual improvement in EB that occurred after the third week of lactation, the depression in the concentration of plasma leptin persisted at week +8 (LACT, P>0·05; Fig. 2). Blood samples were also obtained at hourly intervals over a 26-h period at day −33, +3 and +56 (Fig. 2). Small fluctuations occurred throughout the day but they were

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not associated with time of feeding or milking, or the light:darkness cycle. Therefore, unlike rodents and humans (Schoeller et al. 1997, Ahima et al. 1998), dairy cattle do not have a higher concentration of leptin at night than during the day. This likely reflects the continuous nature of digestion and nutrient absorption in ruminants because this diurnal rhythm is entrained to the timing of meals in humans (Schoeller et al. 1997).

To determine whether changes in leptin expression during the periparturient period could account for changes in the plasma concentration of leptin, abundance of leptin mRNA was measured in subcutaneous WAT (Fig. 3). Leptin mRNA was easily detected during pregnancy and its abundance was similar at week −4 and −1 (Fig. 3). Abundance of leptin mRNA was reduced to ~42% of precalving values during lactation (STATE, P<0.001). This reduction was obvious during the first week of lactation and was sustained over the next 7 weeks (Fig. 3). Therefore, leptin synthesis was reduced immediately after parturition and did not increase with advancing lactation despite improving EB.

Finally, we examined whether changes in WAT mRNA and circulating leptin were related to changes in the concentration of plasma hormones and metabolites (Table 2). Consistent with the EB data, the plasma concentration of NEFA did not differ during pregnancy (PREG, P>0.05), but was elevated during lactation (STATE, P<0.001). The effect of lactation reflects primarily an increase of plasma NEFA in early lactation (E-LACT and LACT, P<0.001). The plasma concentration of insulin changed during lactation in a reciprocal fashion to the plasma concentration of NEFA (Table 2). Therefore, as shown for plasma leptin, the greatest rate of change in the concentrations of insulin and NEFA occurred around the time of parturition. However, in contrast to leptin, their concentrations returned towards pre-calving values as EB improved between weeks 4 and 8 of lactation (Fig. 1 and Table 2).

Plasma concentration of GH increased at parturition and remained elevated during lactation (Table 2; STATE, P<0.001). The plasma concentration of IGF-I was lower during lactation than pregnancy (STATE, P<0.001). This reduction was initiated at the end of pregnancy (PREG, P<0.001), and was sustained until the end of the study (E-LACT and LACT, P>0.05). Plasma concentrations of glucose and cortisol did not differ significantly at any time (Table 2).

**Effect of postpartum EB**

The previous study showed that the plasma concentration of leptin was reduced at parturition, coinciding with the sudden increase in the use of endogenous lipids as a source of energy, but preceding significant depletion of WAT. This suggests that, at parturition, the state of negative EB associated with the onset of milk synthesis may be partly responsible for the reduction of plasma leptin. To test this hypothesis, cows were either never milked after parturition (non-lactating) or milked thrice daily (lactating). When cows were studied at week +1 and +4, the lactating group produced 30.8±2.4 and 33.7±4.6 kg/day of 4% fat-corrected milk respectively. As expected, this experimental manipulation caused dramatic differences in EB after parturition, with non-lactating cows maintaining positive EB and BC, and lactating cows being in negative EB and losing BC (Table 3).
Table 1 Changes in energy-related variables during the transition period

<table>
<thead>
<tr>
<th>Variables</th>
<th>Weeks relative to parturition*</th>
<th>Significance of contrastb</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>−4</td>
<td>−1</td>
</tr>
<tr>
<td>Dry matter intake (kg/day)</td>
<td>13·8</td>
<td>12·1</td>
</tr>
<tr>
<td>Energy intake (Mcal/day)</td>
<td>20·6</td>
<td>19·8</td>
</tr>
<tr>
<td>Milk yield (kg/day)c</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Energy balance (Mcal/day)</td>
<td>6·9</td>
<td>5·9</td>
</tr>
<tr>
<td>Body condition score</td>
<td>3·6</td>
<td>3·7</td>
</tr>
</tbody>
</table>

*Multiparous cows (n=8) were studied during the period between 28 days before and 56 days after parturition. Week −4 corresponds to day −28 through −22, week −1 to day −8 through −2, week +1 to day +2 through +8, week +3 to day +18 through +25 and week +8 to day +50 through +57.

bLinear contrasts were: STATE, physiological state (week −4 and −1 vs week +1, +3 and +8); PREG, stage of pregnancy (week −4 vs −1); LACT, stage of lactation (week +1 and +3 vs week +8); E-LACT, early lactation (week +1 vs +3). Type I error probability where NS=non-significant, P<0.05.

Consistent with the EB data, lactating cows had lower plasma concentrations of insulin and glucose and higher plasma concentrations of GH and NEFA than non-lactating cows (Table 3; P<0.001). Lactating cows also had lower plasma concentration of leptin than non-lactating cows (P<0.001). In contrast, the plasma concentration of cortisol did not differ between treatments. Effects of treatment were constant across times for all variables measured (time and treatment × time, P>0.05). At week +1, the plasma concentration of leptin was positively correlated with plasma concentrations of glucose and insulin (R²=0.7 and 0·6 respectively, P<0.001; Fig. 4), and negatively correlated with the plasma concentrations of GH and NEFA (R²=0·5 for both, P<0.01). Similar correlations were observed at week +4 (data not shown). Overall, these data demonstrate that the onset of an energy deficit at parturition is partly responsible for the lower concentration of plasma leptin in early lactating dairy cows.

Discussion

Lactation is essential to the survival of mammals and represents a substantial transfer of energy from mother to offspring. This transfer is facilitated by numerous endocrine and cellular adaptations. For example, elevated concentration of plasma GH in early lactating ruminants induces metabolic adaptations in liver, skeletal muscle and adipose tissue that promote partitioning of glucose to the mammary gland (Bell 1995, Vernon & Pond 1997, Bauman 2000). Leptin, because of its role in the regulation of feed intake and energy disposition, could also participate in the co-ordination of metabolism during the transition from pregnancy to lactation. There is, however, no information on the dynamics of plasma leptin during the periparturient period in dairy cattle.

We now show that, as in rodents and humans (Kawai et al. 1997, Brogan et al. 1999, Butte et al. 1999, Woodside et al. 2000), the transition from pregnancy to lactation in dairy cows is associated with a reduction in the plasma concentration of leptin. This similarity exists despite the use of different strategies by these species to accommodate the energy needs of early lactation. For example, in this study, mobilization of endogenous lipids was estimated to
meet approximately 33% of the cow’s energy requirements between parturition and the third week of lactation. In contrast, rodents rely primarily on increased feed intake (Barber et al. 1997, Bauman 2000) whereas women, with relatively little change in energy demand from late pregnancy to early lactation (Butte et al. 1999), do not need major adjustments in either intake or lipid mobilization. The postpartum reduction in plasma leptin in all three species may relate to the contrasting priorities of pregnancy and lactation (i.e. storage of energy during pregnancy compared with export during lactation).

The reduction in the plasma concentration of leptin was temporally associated with a 57% decrease in the abundance of leptin mRNA. In other species, additional factors have been shown to contribute to changes in plasma leptin during the periparturient period. In primates, the placenta synthesizes leptin and contributes to circulating leptin during late pregnancy (Masuzaki et al. 1997, Henson et al. 1999). In the mouse, the placenta secretes massive amounts of the soluble form of the leptin receptor, extending the half-life of leptin and accounting for much of the 30-fold greater concentration of leptin in late pregnancy.

### Table 2

<table>
<thead>
<tr>
<th>Variables</th>
<th>Metabolite</th>
<th>Weeks relative to parturition*</th>
<th>Significance of contrastb</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>NEFA (μM)</td>
<td>−4</td>
<td>−1</td>
</tr>
<tr>
<td></td>
<td>Glucose (mg/dl)</td>
<td>107</td>
<td>121</td>
</tr>
<tr>
<td>Hormones</td>
<td>Leptin (ng/ml)</td>
<td>5·8</td>
<td>5·5</td>
</tr>
<tr>
<td></td>
<td>Insulin (ng/ml)</td>
<td>0·8</td>
<td>0·7</td>
</tr>
<tr>
<td></td>
<td>GH (ng/ml)</td>
<td>6·7</td>
<td>6·0</td>
</tr>
<tr>
<td></td>
<td>IGF-I (ng/ml)</td>
<td>124</td>
<td>77</td>
</tr>
<tr>
<td></td>
<td>Cortisol (ng/ml)</td>
<td>2·6</td>
<td>2·4</td>
</tr>
</tbody>
</table>

*a Multiparous cows (n=8) were studied during the period between 28 days before and 56 days after parturition. Week −4 corresponds to day −28 through −22, week −1 to day −8 through −2, week +1 to day +2 through +8, week +3 to day +18 through +25 and week +8 to day +50 through +57.

*b Linear contrasts were: STATE, physiological state (week −4 and −1 vs week +1, +3 and +8); PREG, stage of pregnancy (week −4 vs −1); LACT, stage of lactation (week +1 and +3 vs week +8); E-LACT, early lactation (week +1 vs +3). Type I error probability where NS = non-significant, P > 0·05.

![Figure 3](image-url) Expression of leptin mRNA in WAT during the periparturient period. Eight multiparous dairy cows were studied during the period between 28 days before and 56 days after parturition (day −28 to +56). (Left panel) Total RNA (2 μg) was obtained from WAT at the indicated times and analyzed simultaneously by a ribonuclease protection assay for the abundance of leptin and GAPDH mRNA. The sizes of the protected fragments are 253 bp for leptin and 106 bp for GAPDH. Data are from two representative animals. (Right panel) At each time, the leptin signal was normalized to the signal obtained with the GAPDH probe. Normalized means±S.E. (eight cows) obtained at various times during the periparturient period are represented. The leptin signal was higher during pregnancy than during lactation (STATE, P<0·001), but did not differ during pregnancy or during lactation (PREG, LACT and E-LACT, P>0·05).
pregnant than in non-pregnant animals (Gavrilova et al. 1997). However, leptin is not expressed in ovine or bovine placenta at any stage of pregnancy (R A Ehrhardt, unpublished data). Moreover, the nearly identical concentrations of plasma leptin in late pregnant and postparturient, non-lactating cows indicate that loss of the placenta has no impact on leptin dynamics. Therefore, reduced synthesis of leptin in WAT is largely responsible for the lower concentration of plasma leptin in early lactating dairy cows.

In all species, plasma concentration of leptin is increased by overall adiposity, representing the positive e
cfect of total mass of WAT and adipocyte hypertrophy (Friedman & Halaas 1998, Ahima & Flier 2000, Delavaud et al. 2000, Ehrhardt et al. 2000). In addition, undernutrition decreases and overfeeding increases circulating levels of leptin, well before significant changes in the mass of WAT occurs (MacDougald et al. 1995, Kolaczynski et al. 1996a, 1997, Blache et al. 2000). Our data suggest that the interplay between nutrition and adiposity accounts for most of the changes in plasma leptin in the periparturient dairy cow. Thus, around parturition, onset of an energy deficit causes a reduction in the concentration of circulating leptin. With advancing lactation, however, other factors become more important because the concentration of plasma leptin and leptin mRNA remained low despite improving EB. The most likely factor responsible for this uncoupling of plasma leptin concentration and EB is depletion of WAT as shown by lower BC scores with advancing lactation. It is also possible that failure to return to a threshold EB or other factors associated with lactation contribute to this uncoupling.

The significant correlations between the plasma concentrations of leptin, insulin, GH, glucose and NEFA could represent co-regulation by EB, and perhaps a role for these factors in mediating the effect of EB on leptin synthesis. In support of the latter, insulin upregulates leptin expression in vivo and in vitro in rodent and human WAT (Saladin et al. 1995, Kolaczynski et al. 1996b, Boden et al. 1997), and in bovine WAT explants (Houseknecht et al. 2000). These effects of insulin are dependent on adequate uptake of glucose, suggesting that cellular energy availability is the primary factor regulating leptin synthesis (Mueller et al. 1998, Wellhoener et al. 2000). The CNS, via sympathetic innervation of WAT, could also play an important role in reducing leptin synthesis in early lactation: β-adrenergic signals are potent inhibitors of leptin expression in adipocytes (Hardie et al. 1996, Carulli et al. 1999), and ruminant WAT is particularly sensitive to their metabolic effects in early lactation (Vernon & Pond 1997, Bauman 2000). Sustained periods of negative EB are also characterized by elevation in the plasma concentration of GH and by GH resistance in the liver, resulting in depressed plasma concentration of IGF-I (McGuire et al. 1995, Kobayashi et al. 1999). GH attenuates the ability of insulin and dexamethasone to stimulate leptin synthesis in bovine WAT explants (Houseknecht et al. 2000), but neither GH nor IGF-I has these effects in rat adipocytes (Hardie et al. 1996). Prolactin stimulates leptin synthesis in rats (Gualillo et al. 1999). The concentration of prolactin is increased around the time of parturition in dairy cows (Bell 1995) but the effects of prolactin on leptin synthesis have not been studied in ruminants. Additional studies are needed to ascertain the roles of these factors, and their interactions, in regulating leptin synthesis in the periparturient dairy cow.

The functional consequences of the periparturient reduction in plasma leptin in dairy cows remain to be

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**Table 3** Changes in energy-related variables and plasma metabolites and hormones after parturition

<table>
<thead>
<tr>
<th>Variables</th>
<th>Treatmenta</th>
<th>Non-lactating</th>
<th>Lactating</th>
<th>s.e.</th>
<th>Significanceb</th>
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<td>Whole body energetics</td>
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<td></td>
</tr>
<tr>
<td>Energy balance (Mcal/day)</td>
<td></td>
<td>12·1</td>
<td>−16·8</td>
<td>4·7</td>
<td>0·001</td>
</tr>
<tr>
<td>Δ Body condition scorec</td>
<td></td>
<td>0·1</td>
<td>−0·6</td>
<td>0·2</td>
<td>0·001</td>
</tr>
<tr>
<td>Plasma concentration</td>
<td></td>
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<tr>
<td>Glucose (mg/dl)</td>
<td></td>
<td>59</td>
<td>34</td>
<td>6</td>
<td>0·001</td>
</tr>
<tr>
<td>NEFA (µM)e</td>
<td></td>
<td>218</td>
<td>883</td>
<td>121</td>
<td>0·001</td>
</tr>
<tr>
<td>Leptin (ng/ml)</td>
<td></td>
<td>5·6</td>
<td>2·9</td>
<td>1·4</td>
<td>0·001</td>
</tr>
<tr>
<td>Insulin (ng/ml)</td>
<td></td>
<td>1·9</td>
<td>0·3</td>
<td>0·5</td>
<td>0·001</td>
</tr>
<tr>
<td>Cortisol (ng/ml)</td>
<td></td>
<td>1·5</td>
<td>1·2</td>
<td>1·0</td>
<td>NS</td>
</tr>
<tr>
<td>GH (ng/ml)</td>
<td></td>
<td>2·0</td>
<td>4·6</td>
<td>1·0</td>
<td>0·001</td>
</tr>
</tbody>
</table>

*After parturition, cows were either milked three times per day (lactating, n=7) or never milked (non-lactating, n=7). Cows were studied between day 5 and 11 (week +1) or between day 26 and 33 (week +4). Overall averages are presented because the effects of time and time x treatment were not significant.

*bType I error probability where NS=non-significant, P>0·05.

*cBody condition score=change in body condition score between week +1 and week +3.
characterized but, in the context of the energy deficit associated with early lactation, two possible benefits are immediately obvious. First, reduction in plasma leptin could promote a faster increase in voluntary feed intake. This effect, however, is likely to be minor because physical factors play an important role in constraining feed intake in early lactating ruminants (Ingvartsen & Andersen 2000). Moreover, a 50% decline in plasma leptin may not stimulate appetite significantly given the modest anorexic effect of low doses of exogenous leptin in normal animals (Pelleyounter et al. 1995, Harris et al. 1998). A more immediate benefit of low plasma leptin concentration may be the induction and the co-ordination of the neuroendocrine adaptations responsible for partitioning energy towards essential functions, as shown in fasted rats (Ahima et al. 1996, 1999). Energy conservation is achieved by suppressing functions that are dispensable in the short term, such as reproduction and immunity. Both functions are depressed in early lactating dairy cows, but must be restored rapidly to avoid prolonged infertility and infectious diseases such as mastitis (Mallard et al. 1998, Butler 2000). The possibility that the leptin deficit of early lactation contributes to the development of these problems, as suggested by recent studies in fasted animals (Ahima et al. 1996, Finn et al. 1998, Lord et al. 1998, Nagatani et al. 2000), requires further studies.

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References


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