Pharmacological profile of a new orally active growth hormone secretagogue, SM-130686

J Nagamine, R Nagata, H Seki, N Nomura-Akimaru, Y Ueki, K Kumagai, M Taiji and H Noguchi
Sumitomo Pharmaceuticals Co. Ltd, Discovery Research Laboratories II, 1–98 Kasugade Naka 3-chome, Konohana-ku, Osaka 554–0022, Japan
(Requests for offprints should be addressed to M Taiji, Sumitomo Pharmaceuticals Co. Ltd, Discovery Research Laboratories II, 3–1–98 Kasugadenaka, Konohana-ku, Osaka 554–0022, Japan; Email: taiji@sumitomopharm.co.jp)

Abstract
SM-130686, an oxindole derivative, is a novel orally active GH secretagogue (GHS) which is structurally distinct from previously reported GHSs such as MK-677, NN703 and hexarelin. SM-130686 stimulates GH release from cultured rat pituitary cells in a dose-dependent manner. Half-maximum stimulation was observed at a concentration of 6.3 ± 3.4 nM. SM-130686-induced GH release was inhibited by a GHS antagonist, but not by a GH-releasing hormone antagonist. SM-130686 dose-dependently inhibited the binding of radiolabeled ligand, 35S-MK-677, to human GHS receptor 1a (IC50=1.2 nM). This indicates that SM-130686 stimulates GH release through the GHS receptor. The effect of a single oral administration of SM-130686 on GH release in pentobarbital-anesthetized rats was studied. After treatment with 10 mg/kg SM-130686, plasma GH concentrations measured by radioimmunoassay significantly increased, reaching a peak at 20–45 min, and remained above baseline during the experimental period (60 min). The anabolic effect of repetitive SM-130686 administration was studied in rats. Rats received 10 mg/kg SM-130686 orally twice a day and were weighed every day for 9 days. At day 9 there was a significant increase in both the body weight and the fat free mass (19.5 ± 2.1 and 18.1 ± 7.5 g respectively). Serum IGF-I concentration was also significantly elevated 6 h after the last dose of SM-130686. An endogenous GHS ligand for the GHS receptor has recently been identified from stomach extract and designated as ghrelin. The GH-releasing activity in vitro relative to ghrelin (100%) was about 52% for SM-130686. It is likely that SM-130686 is a partial agonist for the GHS receptor.

In summary, we describe here an orally active GHS, SM-130686, which acts through the GHS receptor. Repetitive administration of SM-130686 to rats, similar to repetitive administration of GH, significantly increased the fat free mass by an amount almost equal to the gain in body weight.

Introduction
The secretion of growth hormone (GH) is regulated by the neuroendocrine control system like other anterior pituitary hormones. Two main hypothalamic factors, GH-releasing hormone (GHRH) and somatostatin, are known to be involved as positive and negative regulators, respectively, in GH release (Frohman et al. 1992). After GH-releasing peptide (GHRP)-6 and its analogues were found to enhance GH secretion as well as GHRH, a number of new chemical entities with similar characteristics were successfully synthesized. The peptidic factors with GH-releasing activity include GHRP-1, GHRP-2, GHRP-6 and hexarelin; other non-peptidic compounds include L-692,429 and MK-677 (Camanni et al. 1998). Among these GH secretagogues (GHSs), MK-677 has been reported to show good oral bioavailability. In addition, chemically synthesized GHSs including GHRP-6 and MK-677 have been reported to show highly potent GH-releasing activity in several species including humans (Walker et al. 1990, Bowers et al. 1992, Hartman et al. 1992, Chapman et al. 1996).

Using the affinity of MK-677 as a clue, a novel GHS receptor was cloned by Howard et al. (1996). This receptor is expressed in various kinds of tissues: pituitary, hypothalamus and other areas of the central nervous system (CNS) (Guan et al. 1997). Recently, an endogenous GHS ligand for the GHS receptor has been identified from stomach extract and designated as ghrelin, an acylated peptide of 28 amino acids (Kojima et al. 1999). Ghrelin mRNA is present in the hypothalamus arcuate nucleus and stomach. Intravenous injection of ghrelin stimulated GH release, suggesting that GH secretion in the pituitary may be regulated not only by ghrelin expressed in the hypothalamus, but also by ghrelin released from the stomach. Since ghrelin mRNA was reported most abundantly in the
482 | NAGAMINE and others · SM-130686: a new orally active GH secretagogue

stomach, it is very likely that ghrelin has some other physiological functions (Dieguez & Casanueva 2000).

We screened our chemical library for a new GHS using a rat pituitary cell culture system and found an oxindole derivative that showed GH-releasing activity. Modification of this oxindole derivative led to an entirely new class of non-peptidic GHSs represented by SM-130686 (Fig. 1) (Tokunaga et al. 2001), which are structurally different from GHRP-6, hexarelin, MK-677 and NN703 (Hansen et al. 1999). Here we report the pharmacological profile of a new type of GHS, SM-130686.

Materials and Methods

Animals

Adult male Wistar/ST rats and female F344/N rats were purchased from SLC Japan Inc., Sizuoka, Japan. All rats were kept in a temperature (23 ± 2 °C)- and humidity (55 ± 10%)-controlled room with lights on between 0800 and 2000 h. Water and standard laboratory chow (CE-2; Cler Japan, Inc., Tokyo, Japan) were freely available. All animal experiments were conducted according to the guidelines of the Sumitomo Pharmaceuticals Committee on Animal Research.

Materials

Horse and fetal calf sera, non-essential amino acids (100-fold concentrated), penicillin–streptomycin solution and Hank’s balanced salt solution (HBSS) were purchased from Gibco, New York, NY, USA. Dulbecco’s modified Eagle’s medium (DMEM) was from Iwaki, Tokyo, Japan. Collagenase was from Worthington Biochemical, Lakewood, NJ, USA. DNase type I was from Sigma, St Louis, MO, USA. EDTA, HEPES, NaCl, KCl, Na2HPO4, CaCl2 and dimethyl sulfoxide were from Nacalai Tesque, Kyoto, Japan. Ghrelin was synthesized using the method previously reported by Tschöp et al. (2000).

Rat pituitary cell assay

Anterior pituitaries were obtained from 7-week-old male Wistar/ST rats. The tissue was washed three times with HBSS, cut into small pieces and transferred to the isolation buffer (100 µl/lobe) containing 0.8% collagenase and 0.2% DNase type I. The mixture was incubated for less than 25 min at 37 °C. The cells were collected and washed three times with the culture medium; DMEM was supplemented with 5% horse serum, 2.5% fetal calf serum, 1% non-essential amino acids, 1% penicillin and 1% streptomycin. Cells were resuspended in the above medium and brought to a density of 1.0 × 10^5 cells/ml. The cells (200 µl/well) were seeded onto a 96-well plate (Nunc, Roskilde, Denmark) and cultured for 5 days at 37 °C under 5% CO2.

Following the culture period the cells were washed once with the stimulation buffer (culture medium containing 25 mM HEPES, pH 7.3) and then incubated for 90 min at 37 °C under 5% CO2. This buffer was then replaced with a fresh stimulation buffer containing SM-130686 followed by incubation for 15 min at 37 °C under 5% CO2. The medium was collected and assayed for GH concentration. To study the effect of antagonists, the cells were incubated with an antagonist (final additive concentration 10 µM) before the SM-130686 solution was added.

GHS-binding activity

Crude membranes were isolated from CHO cells stably expressing the human GHS receptor 1α. These were suspended in binding buffer (50 mM Tris–HCl, 10 mM MgCl2, 2.5 mM EDTA, pH 7.4) to a concentration of 1 mg protein/ml. These membranes (25 µg protein/well) were mixed with ^35^S-MK-677 (100 000 d.p.m./well), with or without different concentrations (0.1–100 nM) of SM-130686 and binding buffer up to a total volume of 250 µl. Non-specific binding was obtained by adding 500 nM cold MK-677. The membranes were incubated at 25 °C for 60 min, and bound radioligand was separated from free radioligand by washing with binding buffer through a GF/B filter (Perkin Elmer, Boston, MA, USA). The radioactivity on the filters was counted in a scintillation counter (Perkin Elmer).

GH response of SM-130686 in anesthetized rats

Adult male rats (8 weeks old, 250–300 g) were anesthetized by intraperitoneal injection of pentobarbital; a catheter was inserted in the right jugular vein for blood
GH response of SM-130686 in conscious rats

Twenty-seven adult male rats (16 weeks old, 350–400 g) were prepared 7 days before the experiment with indwelling right jugular catheters under pentobarbital anesthesia. After surgery, water was administered orally to the animals to minimize treatment stress. Catheters were kept patent by flushing with 200 µl heparinized saline (50 U/ml) every day. At 0830 h on the day of the experiment, all lines were fitted with extensions and collected into a tube containing 5 U (50 µl) heparin. Plasma (50 µl) was separated by centrifugation and mixed with 200 µl radioimmunoassay (RIA) buffer. The samples were stored at −40 °C until being assayed for rat GH.

Calculations

Using the EC50 value of SM-130686, the potency was calculated as the concentration inducing half-maximal stimulation with the following scheme (GH content = Emax × concentration of SM-130686/(EC50+ concentration of SM-130686) + basal GH content). Area under the concentration (AUC) in the in vivo study was calculated using the trapezoidal rule. All data are reported as means ± s.d. except for the EC50 and Emax values of GH release in the in vitro study (means ± s.e.m.) and GH release in the in vivo study. All statistical comparisons were performed using SAS (version 6.12; Cray, NC, USA).

Results

GH-releasing activity of SM-130686 through the GHS receptor

The GH-releasing activity of SM-130686 was studied using cultured rat pituitary cells. GH concentration in the culture medium was determined by RIA 15 min after treatment with SM-130686. SM-130686 enhanced GH release dose-dependently with EC50 values of 6.3 ± 3.4 nM (means ± s.e.m.) in five different experiments with rat pituitary primary culture (Fig. 2A). To evaluate the pharmacological properties of SM-130686, we investigated the effects of concomitant treatment of GHS antagonist ([D-Arg1,D-Phe5,D-Trp7,9,Leu11]-substance P) or GHRH antagonist ([N-acetyl-Try1,D-Arg2]–hGHRH(1–29)NH2) with SM-130686 on GH-releasing activity (Cheng et al. 1997). As shown in Fig. 2B, GHS antagonist inhibited the GH-releasing activity of SM-130686, whereas the GHRH antagonist had neither an additive nor a synergistic effect. These results suggested that SM-130686 increases GH release by acting on the GHS receptor.

We therefore decided to evaluate the binding affinity of SM-130686 for the GHS receptor which was originally identified as MK-677–binding protein and recently found to be the receptor for an endogenous ligand called ghrelin. As shown in Fig. 3, SM-130686 dose–dependently inhibited the binding of radioisotope–labeled ligand, 35S-MK-677, to recombinant human GHS receptor 1a that was stably expressed in CHO cells. The I50 value of...
SM-130686 was calculated to be $1.2 \pm 0.2$ nM in this assay. In order to investigate whether SM-130686 was specifically bound to the GHS receptor or not, in vitro binding activities of SM-130686 were extensively studied on more than 50 other receptors, including opiate, sigma, bradykinin, muscarine, neurokinin, benzodiazepine, serotonin, dopamine, gamma aminobutyric acid, GHRH and somatostatin receptors. The IC$_{50}$ values of SM-130686 for these receptors were more than 1 µM (data not shown) indicating that SM-130686 has specific and high affinity for the GHS receptor.

**Effect of SM-130686 administration on GH release in pentobarbital-anesthetized rats**

To study GH-releasing activity in vivo, SM-130686 was intravenously injected into pentobarbital-anesthetized rats through a catheter inserted into the right jugular vein. After intravenous injections of 1 and 3 mg/kg SM-130686, the plasma GH concentrations of each group of rats given SM-130686 reached a peak at 4 min and returned to baseline by 16 min (Fig. 4A). The peak plasma GH concentration increased in a dose-dependent manner.

We investigated the pharmacokinetic profile of SM-130686 in rats after oral administration. Bioavailability was found to be 27.6%, indicating that SM-130686 is an orally active GHS. It was therefore decided to investigate the effect of oral administration of SM-130686 on GH release in pentobarbital-anesthetized rats. After oral administration of 1, 3 and 10 mg/kg SM-130686, the peak plasma GH concentrations increased in a dose-dependent manner and remained above baseline during the experimental period of 60 min (Fig. 4B).

**GH response to oral administration of SM-130686 in conscious male rats**

Following the studies using anesthetized rats, the efficacy of SM-130686 oral administration was further assessed by monitoring GH-releasing activity in conscious rats. Rats were implanted with cannulae for blood sampling into the right jugular veins under pentobarbital anesthesia 7 days before the SM-130686 administration experiment. After a single oral administration of SM-130686, plasma GH concentration peaked within 3 h in all four rats (Fig. 5 lower panel). AUC and Cmax values for the plasma concentrations were significantly different from baseline.
GH concentration during the first 3 h after 10 mg/kg SM-130686 administration increased compared with vehicle-treated rats (Table 1).

Anabolic effects of SM-130686 in normal female rats

The results on GH-releasing activity of SM-130686 in vitro and in vivo encouraged us to investigate further the efficacy of repetitive administration. SM-130686 was orally administered to normal female rats every day for 9 days (starting at day 0). The rats were weighed every morning just before administration of the drug, and the fat free mass which was mainly composed of muscle, bone and water, was measured by the bioelectrical impedance method at days 1 and 9. Treatment with 6 mg/kg SM-130686 per day did not significantly affect the body weight gain in rats (Fig. 6). However, rats given 20 mg/kg SM-130686 per day showed a significantly higher body weight gain at day 1 compared with vehicle-treated rats. This enhanced body weight gain in the group given 20 mg/kg per day continued with no apparent sign of attenuation during the experimental period of giving repetitive daily administration. At the end of the treatment (day 9), the gain in fat free mass significantly increased in the SM-130686-treated (20 mg/kg per day) rats (Table 2). Administration of 6 and 20 mg/kg SM-130686 per day increased the plasma IGF-I levels in a dose-dependent manner 6 h after the last administration (day 9) (Table 2).

**Discussion**

Among the peptidic and non-peptidic chemical compounds with GH-releasing activity, MK-677 has been reported to show good oral bioavailability (Patchett et al. 1995). In this study we have demonstrated a new class of orally active GHSs represented by SM-130686. The EC_{50} of SM-130686 in enhancing GH release was 6.3 ± 3.4 nM, nearly as potent as MK-677 with an EC_{50} value of 2.7 ± 0.9 nM as measured in our experiment. The result of a binding assay using ^{35}S-MK-677 as a radioligand also showed that the affinity of SM-130686 with the human GHS receptor 1a (IC_{50}=1.2 nM) was similar to MK-677 (IC_{50}=0.5 nM). The biological activity of SM-130686 in enhancing GH release was blocked by the GHS antagonist, but not by the GHRH antagonist in rat pituitary cells. These pharmacological properties indicate that our SM-130686 enhances GH release by acting on the GHS receptor.

Physiologically, GH is secreted in a pulsatile fashion (Frohman et al. 1992, Müller et al. 1999). Since the dosing of exogenous GH (once daily or less) does not mimic the normal GH secretion profile, a more natural treatment needs to be developed for patients with GH deficiency. Furthermore, it is known that plasma concentrations of GH and IGF-I wane with aging in the human, and mean...
plasma GH concentration in people over 60 years old is approximately one third to one half that of young adults (Rudman 1985, Zadik et al. 1985, Ho et al. 1987, Iranmanesh et al. 1991). Notably, daily oral administration of MK-677 increases plasma GH concentration in healthy elderly subjects and also increases their plasma IGF-I concentration to the normal range of young adults (Chapman et al. 1996). Further, the enhancement of pulsatile GH release by MK-677 has been reported to be due to an increase of pulse height (=mass of GH secreted per pulse) rather than the number of pulses in plasma GH secretion profiles. Similarly, the present study

Table 1 GH response after oral administration of SM-130686 in conscious male rats. Values are for individual rats* or means ± S.D.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Tmax (min)*</th>
<th>Cmax (ng/ml) 0–360 min</th>
<th>Cmax (ng/ml) 180–360 min</th>
<th>AUC (ng/ml · h) 0–360 min</th>
<th>AUC (ng/ml · h) 180–360 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vehicle (n=3)</td>
<td>180, 200, 300</td>
<td>50 ± 26</td>
<td>50 ± 26</td>
<td>42 ± 23</td>
<td>24 ± 10</td>
</tr>
<tr>
<td>SM-130686 (20 mg/kg per day) (n=4)</td>
<td>80, 160, 160, 160</td>
<td>124 ± 64</td>
<td>56 ± 47</td>
<td>112 ± 53</td>
<td>33 ± 17</td>
</tr>
</tbody>
</table>
demonstrated that SM-130686 enhanced pulsatile GH secretion in conscious male rats. These characteristics indicate that GHSs might lead to a more desirable treatment not only for GH deficiency but also for healthy elderly subjects.

GH plays a critical role in a variety of physiological processes, including longitudinal bone growth and body metabolism. With regards to body metabolism, it has been shown previously that GH possesses anabolic, lipolytic and anti-natriuretic actions, and thus affects body composition (Møller et al. 1995). The effects of GH on body composition in mature female rats have been investigated using a two-compartment model of fat mass and fat free mass (mainly composed of muscle, bone and water). Results showed that body weight gain after GH treatment was largely due to fat free mass accretion (Azain et al. 1995). It has also been reported that peptidic GHSs such as GHRP-6 (Bowers et al. 1984), G7039 (McDowell et al. 1995) and NN703 (Hansen et al. 1999) significantly stimulate body weight gain in female rats after daily administration for more than 7 days. Peptidic GHSs have been known to stimulate food intake, possibly through direct action on the CNS (Locke et al. 1995, Okada et al. 1996), and thus the increase in body weight gain is partly due to fat mass gain.

Recently, an endogenous GHS ligand for the GHS receptor has been identified and designated as ghrelin (Kojima et al. 1999). Ghrelin increases body weight gain by stimulating fat mass gain without any change in fat free mass in mice (Tschöp et al. 2000). Thus, it may be that ghrelin itself enhances glucose catabolism and reduces fat utilization in mice and rats. Similar data were observed after administration of KP-102, a peptidic GHS, indicating a GHS/ghrelin receptor-mediated mechanism (Tschöp et al. 2000). In the present study, repetitive administration of SM-130686 significantly increased the fat free mass by almost the same amount as the gain in body weight. Further, we preliminarily confirmed that defatted dry mass (composed of body protein and minerals etc.) of rats given

### Table 2

<table>
<thead>
<tr>
<th>Parameter</th>
<th>SM-130686 (mg/kg per day)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weight gain (g)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>13.5 ± 2.3</td>
</tr>
<tr>
<td>6</td>
<td>15.1 ± 1.9</td>
</tr>
<tr>
<td>20</td>
<td>19.5 ± 2.1**</td>
</tr>
<tr>
<td>Fat free mass gain (g)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>7.2 ± 6.3</td>
</tr>
<tr>
<td>6</td>
<td>8.9 ± 8.0</td>
</tr>
<tr>
<td>20</td>
<td>18.1 ± 7.5*</td>
</tr>
<tr>
<td>% body fat</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>18.0 ± 4.9</td>
</tr>
<tr>
<td>6</td>
<td>17.2 ± 4.2</td>
</tr>
<tr>
<td>20</td>
<td>14.5 ± 3.9</td>
</tr>
<tr>
<td>Plasma IGF-I (ng/ml)</td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>729 ± 75</td>
</tr>
<tr>
<td>6</td>
<td>777 ± 39</td>
</tr>
<tr>
<td>20</td>
<td>842 ± 50**</td>
</tr>
</tbody>
</table>

*P<0.025, **P<0.005, using Williams test.
SM-130686: a new orally active GH secretagogue

regenerative administration of vehicle, and 6 mg/kg and 20 mg/kg SM-130686 per day were 32.3 g, 32.3 g and 34.1 g respectively. These data suggest that regenerative administration of SM-130686 also increases body protein. The food intake of rats given 20 mg/kg SM-130686 per day (7.6 g/100 g body weight per day) was not significantly different from rats given vehicle (6.9 g/100 g body weight per day). Compared with ghrelin and other GHSs, the action of SM-130686 may be closer to GH itself and more advantageous for the treatment of GH deficiency patients without deteriorating body composition or inducing obesity.

The reason why SM-130686 acts differently from ghrelin still remains unknown. We have demonstrated that the GH-releasing activity of SM-130686 was less potent (52 ± 12%) than ghrelin in rat pituitary cells, whereas SM-130686 has almost the same binding affinity as ghrelin (0–25 nM in Bednarek et al. 2000). These data indicate that SM-130686 might be a partial agonist for the GHS receptor, which may be one of the reasons for the difference between SM-130686 and ghrelin. Further studies will also be needed to unveil the mechanism of the action of SM-130686 in the endocrine system.

In summary, SM-130686 enhanced GH release from rat pituitary cells, possibly through a GHS receptor, and stimulated GH secretion both in anesthetized and conscious rats. Repetitive administration of SM-130686 significantly increased fat mass gain in addition to body weight gain in normal female rats. Taken together, this indicates that repetitive SM-130686 treatment has an anabolic effect similar to repetitive GH treatment.

Acknowledgements

We wish to thank Mr T Tokunaga, Mr E W Hume and Mr K Okazaki for their contribution and Ms N Okahara-Mitsuhata for her technical assistance.

References


Guan XM, Yu H, Palyha OC, McKee KK, Feighner SD, Sirinathsinhji DJS, Smith RG, Van der Ploeg LHT & Howard AD 1997 Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. Molecular Brain Research 48 23–29.


Received 3 May 2001
Accepted 28 August 2001