

Potential role of activin A in follicular development during the second half of pregnancy in the golden hamster: utero-placental source of activin A

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Abstract

Numerous antral follicles develop during the second half of pregnancy in the golden hamster. However, mechanisms regulating follicular development during this period are unknown. Because inhibin and activin are related to follicular development, these hormones were studied to gain insight into any potential roles in follicular development. Plasma inhibin A and B suddenly increased from day 8 of pregnancy, reached peak levels on day 10 and gradually declined to term. Plasma activin A gradually increased from day 8 to day 15 of pregnancy, and this was followed by an abrupt decrease at day one of lactation. Ovariectomy on day 12 of pregnancy rapidly reduced plasma inhibin A and B, but not activin A levels.

Hysterectomy or placentectomy on day 12 of pregnancy caused an abrupt decrease in the levels of plasma activin A and FSH, but not inhibin A and B at 6 h after surgery. Hysterectomy also induced atresia of large antral follicles at 24 h after surgery. These results indicate that antral follicles are the main source of circulating inhibin A and B, whereas uteri and placentae are the main source of circulating activin A. These results suggest that increased levels of activin A may be involved in folliculogenesis in the ovary during the second half of pregnancy in the golden hamster.

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Introduction

Inhibins and activins are structurally related dimeric proteins first identified in ovarian follicular fluid (Robertson *et al.* 1985, Vale *et al.* 1986). Inhibins are heterodimers consisting of a common α subunit and either a β A or β B subunit, resulting in inhibin A or inhibin B respectively (De Jong 1988). Inhibins are important factors in regulating secretion of follicle stimulating hormone (FSH) in mammals. Inhibins, through the regulation of FSH secretion, have been reported to be key factors for determining species-specific ovulation rates (Taya *et al.* 1999, Taya 1993, Kishi *et al.* 1996, Taya *et al.* 1996). Activins are dimers of the β subunits, with three forms currently identified: activin A (β A- β A), activin B (β B- β B) and activin AB (β A- β B) (Ying 1988). Activins are important proteins that have several physiological functions in reproduction, such as stimulation of FSH secretion

(Vale 1986, Schwall *et al.* 1989) and increasing the number of FSH receptors (Hasegawa *et al.* 1988, Nakamura *et al.* 1993). Activin is also well known as an important factor that may act in the early development in the formation of the mesoderm (Albano *et al.* 1990, Smith *et al.* 1990).

During the second half of gestation in the golden hamster, numerous antral follicles develop (Greenwald 1964), but all large antral follicles degenerate at the end of pregnancy. Post partum ovulation, as seen in rats (Ying *et al.* 1973), does not occur in the golden hamster (Greenwald 1965) and large antral follicles do not reappear until the end of lactation. The proliferation of numerous antral follicles during the second half of pregnancy in the golden hamster is also markedly different compared with the pattern in the pregnant rat (Greenwald 1966, Taya & Sasamoto 1977). Mechanisms of follicular dynamics during the second half of pregnancy in the golden hamster are unclear.

In the present study, to investigate mechanisms of follicular dynamics in the second half of pregnancy in the golden hamster, secretory patterns of inhibin A, inhibin B and activin A, and the source of these hormones were determined.

Materials and Methods

Animals

Adult female golden hamsters (*Mesocricetus auratus*), maintained on a 14 h light:10 h darkness schedule (lights on from 0500 h to 1900 h) were mated after two consecutive 4-day oestrous cycles. Female hamsters were placed with males on the evening of pro-oestrus, and the presence of sperm in the vagina was designated as day 0 of pregnancy. In our colony, gestation lasts 15 days, with most hamsters delivering on day 15 of pregnancy. The day of parturition was designated as day 0 of lactation. Food and water were available *ab libitum*.

Experimental design

Plasma concentrations of inhibin A, inhibin B and activin A during the second half of pregnancy

Groups of animals ($n=5$) were decapitated daily at 1100 h from day 8 of pregnancy to day 1 of lactation, and trunk blood and ovaries were collected. Blood samples were centrifuged immediately at 1700 g for 15 min at 4 °C and plasma was separated and stored at -20 °C until assayed. Ovaries, uteri and placentae were removed immediately after death and weighed. Ovaries, uteri and placentae were stored at -20 °C in 1 ml 0.85% (w/v) NaCl solution. Ovaries, uteri and placentae were thawed on ice, homogenized (UR 200P; Tomy, Tokyo, Japan) and centrifuged at 20 000 g for 30 min at 4 °C. Supernatants were stored at -20 °C until assayed for activin A.

Ovarian follicular development during pregnancy

To determine the number of large healthy antral follicles in the ovary during the second half of pregnancy, 10 IU human chorionic gonadotrophin (hCG) dissolved in 0.2 ml 0.85% (w/v) NaCl solution were injected into the jugular vein under ether anaesthesia (2200 iu/mg; Sankyo Zoki Co., Tokyo, Japan) on days 2, 4, 6, 8, 10, 12 or 14 of pregnancy. Animals were decapitated at 18–20 h after hCG injection, and the oviducts were examined for oocytes.

Plasma concentrations of inhibin A, inhibin B and activin A after surgery

To examine the sources of inhibin A, inhibin B and activin A, pregnant hamsters underwent one of the following: ovariectomy, placentectomy (placentae with decidual tissues and fetuses were removed, and the uterus was not sutured), hyster-

ectomy (uterus, placentae with decidual tissues and fetuses were removed), ovariectomy or sham-operation under ether anaesthesia at 1100 h on day 12 of pregnancy. Just before, and 1, 3 and 6 h after surgery, groups of animals were decapitated and trunk blood was collected. Blood samples were centrifuged immediately at 1700 g for 15 min at 4 °C, and plasma was separated and stored at -20 °C until assayed.

The number of large healthy antral follicles after hysterectomy on day 12 of pregnancy

To examine the effect of hysterectomy on follicular activity, pregnant hamsters were hysterectomized at 1100 h on day 12 of pregnancy, then 10 IU hCG dissolved in 0.2 ml 0.85% (w/v) NaCl were injected into the jugular vein under ether anaesthesia at 1100 h on day 13 of pregnancy (24 h after the surgery). Animals were decapitated 18–20 h after hCG injection, and the oviducts were examined for oocytes.

Enzyme linked immunoabsorbent assay (ELISA) of inhibin A, inhibin B and activin A

Concentrations of inhibin A, inhibin B and activin A were measured by ELISA (Serotec, Oxford, Oxon, UK) as described previously (Knight *et al.* 1996, Ohshima *et al.* 1999). Amounts of inhibin A and inhibin B were expressed in terms of recombinant human inhibin A and inhibin B respectively. Amounts of activin A were expressed in terms of recombinant activin A.

RIA of luteinising hormone (LH), FSH, progesterone, and oestradiol-17 β

Concentrations of LH and FSH in plasma were measured using NIDDK RIA kits for rat LH and FSH as described previously (Bast & Greenwald 1974a). Iodinated preparations were rat LH-I-5 and FSH-I-5. The antisera used were anti-rat LH-S-9 and anti-rat FSH-S-11. The intra- and interassay coefficients of variation were 8.9% and 6.7% for LH and 4.4% and 14.6% for FSH respectively.

Concentrations of progesterone and oestradiol-17 β in plasma were measured by double antibody RIA systems using ¹²⁵I-labelled radioligands as described previously (Taya *et al.* 1985). Antisera against progesterone (GDN 337; Gibori *et al.* 1977) and oestradiol-17 β (GDN 244; Korenman *et al.* 1974) were kindly provided by Dr G D Niswender (Colorado State University, Fort Collins, CO, USA). The intra- and interassay coefficients of variation were 6.3 and 15.4% for progesterone and 3.7% and 6.2% for oestradiol-17 β respectively.

Statistics

All data were expressed as means \pm s.e.m. One-way ANOVA was performed, and the significance between two means was determined by Student's *t*-test or Cochran-Cox test, and the significance among more than

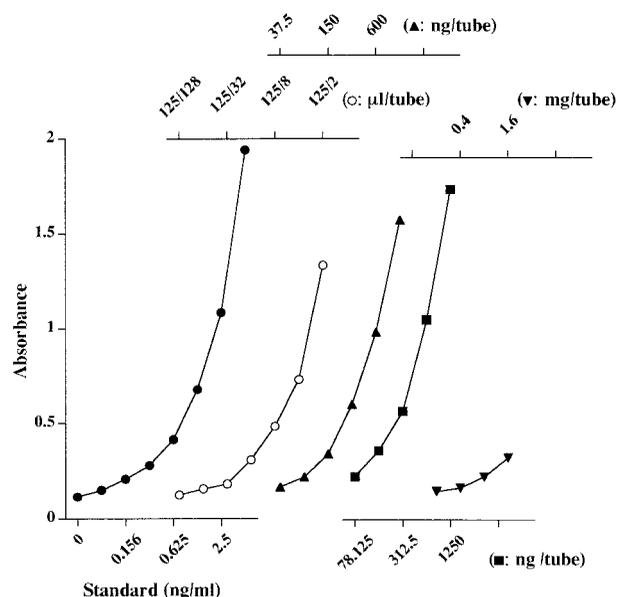


Figure 1 ELISAs for activin A. Dose–response curves of activin A in standard (●), plasma (○), placental homogenates (▲), uteri homogenates (■) and ovarian homogenates (▼) on day 12 of pregnancy in the female golden hamsters.

two means was determined by Duncan's Multiple Range test (Steel & Torrie 1960). A value of $P < 0.05$ was considered statistically significant.

Results

Characterization of activin A ELISA (Fig. 1)

Competition between labelled and unlabelled antigen with the ovarian homogenates, peripheral plasma, placental homogenates and uterine homogenates of pregnant hamsters produced excellent dose–response curves in activin A assays. Each curve was reliably parallel with the respective standard curve, indicating that it was possible to measure the concentration of activin A of the female golden hamster using this kit. These results also clearly demonstrated that placenta and uterus of pregnant golden hamsters contained large amounts of activin A.

Induction of ovulation during the second half of pregnancy (Table 1)

Although a constant number of follicles ovulating in response to hCG (16.0 ± 0.7 on day 2; 15.8 ± 1.5 on day 8) was maintained in the ovary from day 2 to day 8 of pregnancy, the number of follicles ovulating in response to hCG approximately doubled on day 10 of pregnancy (33.2 ± 3.2) and was maintained until day 14 of pregnancy.

Table 1 Induction of ovulation by 10 IU human chorionic gonadotrophin (hCG) administered on various days during pregnancy in the golden hamster. All hamsters (5 per group) ovulated at the expected time. Results are means \pm S.E.M.

Day of pregnancy	No. of oocytes/ovulating hamster
2	16.0 ± 0.7^a
4	15.4 ± 2.2^a
6	18.4 ± 1.4^a
8	15.8 ± 1.5^a
10	33.2 ± 3.2^b
12	34.6 ± 3.8^b
14	27.4 ± 4.4^b

Values with different superscripts are significantly different ($P < 0.05$, Duncan's test).

Plasma concentrations of inhibin A, inhibin B, activin A, FSH, LH, oestradiol-17 β and progesterone during the second half of pregnancy (Fig. 2)

Plasma concentrations of inhibin A and inhibin B abruptly increased from day 8 and reached maximum levels on day 10 of pregnancy. Thereafter, plasma concentrations of inhibin A and inhibin B gradually decreased until day 1 of lactation.

Plasma concentrations of activin A significantly increased from day 8 (0.55 ± 0.03 ng/ml) to day 9 (1.48 ± 0.31 ng/ml) of pregnancy. Thereafter, plasma concentrations of activin A increased gradually, and reached the maximum level on day 15 of pregnancy; this was followed by an abrupt decrease with delivery.

Plasma concentrations of FSH and LH remained at basal levels and the levels were unchanged during the second half of pregnancy.

Plasma concentrations of oestradiol-17 β increased from day 8 and reached plateau levels on day 9 of pregnancy. The plateau level was maintained until day 13, and was followed by an abrupt decline to below the limit of detection on day 15 of pregnancy.

Plasma concentrations of progesterone were maintained at plateau levels from day 8 to day 10 of pregnancy. Thereafter, plasma concentrations of progesterone further increased and reached the maximum level on day 13, followed by an abrupt decline on day 15 of pregnancy.

Plasma concentrations of inhibin A, inhibin B, activin A and FSH after ovariectomy, ovariectomy, placentectomy or hysterectomy (Fig. 3)

Plasma concentrations of inhibin A and inhibin B declined markedly after ovariectomy or ovariectomy,

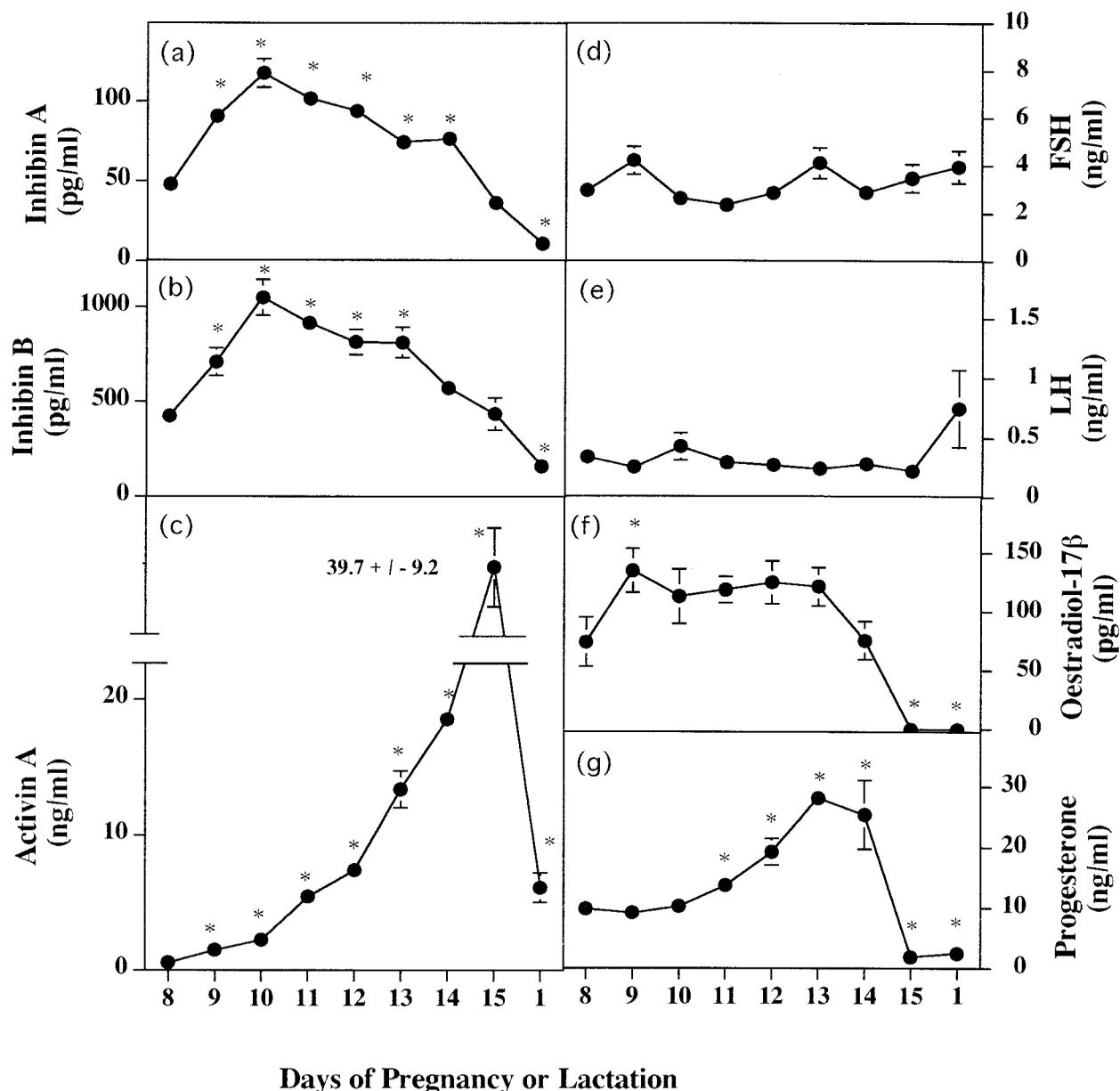


Figure 2 Changes in plasma concentrations of (a) inhibin A, (b) inhibin B, (c) activin A, (d) FSH, (e) LH, (f) oestradiol-17 β and (g) progesterone during the second half of pregnancy and in early lactation in the golden hamster. Each value represents the mean \pm S.E.M. of five animals. * $P < 0.05$ compared with the value at day 8 of pregnancy (Duncan's Multiple Range test for inhibin A, inhibin B, FSH, LH and oestradiol-17 β ; Student's *t*-test for activin A and progesterone).

whereas there were no differences between sham-operation and placentectomy or hysterectomy.

Plasma concentrations of activin A in the placentectomized, hysterectomized and ovariectomized groups significantly declined at 1 h after operation and the level gradually decreased to 34.6, 8.6 or 9.6% of sham-operated controls, respectively, at 6 h post surgery. On the other

hand, plasma concentrations of activin A in the sham-operated and ovariectomized groups did not show any significant changes throughout the 6 h following surgery.

Plasma concentrations of FSH increased markedly at 6 h after ovariectomy and ovariectomized groups, whereas plasma concentrations of FSH decreased at 6 h after placentectomy or hysterectomy.

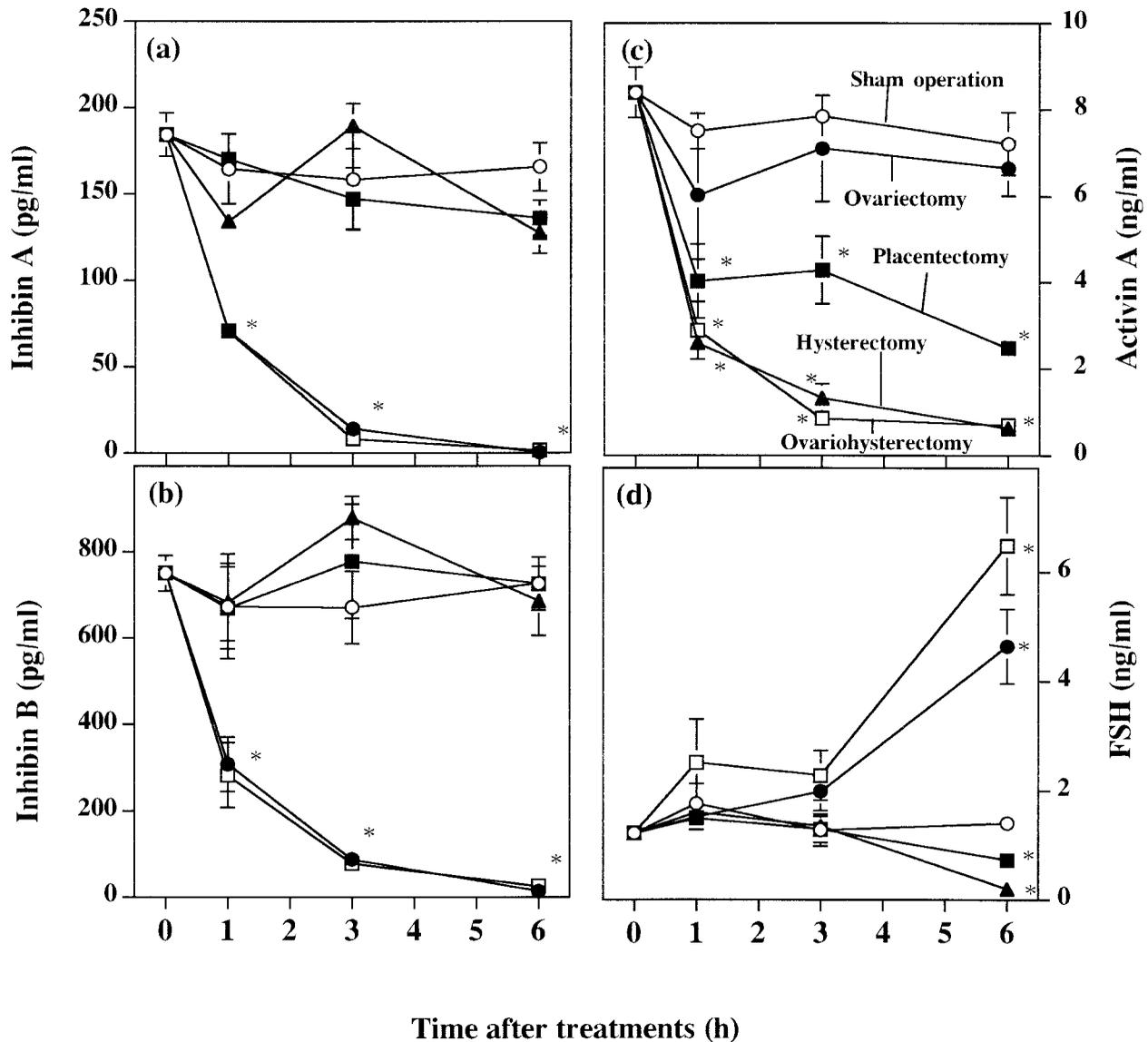


Figure 3 Plasma concentrations of (a) inhibin A, (b) inhibin B, (c) activin A and (d) FSH after ovariectomy (●), placentectomy (■), hysterectomy (▲), ovari hysterectomy (□) or sham operation (○) at 1100 h on day 12 of pregnancy in the golden hamster. Each value represents the mean \pm S.E.M. of five observations. * $P < 0.05$, significantly different from time-matched sham-operated group (Student's *t*-test for inhibin A and activin A; Duncan's New Multiple Range test for inhibin B and FSH).

Induction of ovulation after hysterectomy (Table 2)

Hysterectomy at 1100 h on day 12 of pregnancy significantly reduced the number of follicles ovulating in response to hCG as compared with the sham-operated group.

Discussion

This study clearly demonstrates that plasma concentrations of inhibin A, inhibin B, oestradiol-17 β and activin A

Table 2 Induction of ovulation in sham-operated or hysterectomized hamsters by 10 IU hCG on day 13 of pregnancy. Pregnant hamsters were hysterectomized or sham-operated on day 12 of pregnancy

Treatment group	Number of ova on day 14	Hamsters ovulating/hamsters examined
Sham-operated	25.4 \pm 1.4	5/5
Hysterectomized	1.5 (1, 2)	2/6

increased during the second half of pregnancy in the golden hamster. These changes in circulating inhibins correlated well with the number of healthy large antral follicles ovulating in response to hCG. Greenwald (1967) demonstrated that the number of follicles larger than 415 μm in diameter had a relationship with the number of ovulations induced by hCG throughout pregnancy in the hamster. Previous studies (Kishi *et al.* 1995, Ohshima *et al.* 1999) also demonstrated that the changing pattern of plasma concentrations of inhibin corresponded well with the number of healthy large antral follicles during the oestrous cycle. Similar correlations of inhibin and follicular development have also been observed in female rats in the various reproductive stages (Taya *et al.* 1989). In the present study, after ovariectomy but not after hysterectomy or placentectomy, plasma concentrations of inhibin A and inhibin B decreased abruptly. These observations clearly indicate that the ovarian antral follicles are responsible for the increase in plasma inhibins during the second half of pregnancy in the golden hamster. On the other hand, after placentectomy or hysterectomy, but not after ovariectomy, plasma concentrations of activin A markedly decreased 1 h after surgery, and placental and uterine homogenates contained a large amount of activin A. These results indicate that activin A is likely to be secreted from placenta and uteri during the second half of pregnancy in the golden hamster. Results in a previous paper demonstrated that activin βA mRNA was expressed in the rat placenta (Gu *et al.* 1995). Previous reports demonstrated that the treatment of granulosa cells with activin increased FSH receptor (Hasegawa *et al.* 1988, Nakamura *et al.* 1993). In addition, hysterectomy on day 12 of pregnancy, which lowered activin A secretion within 1 h, noticeably reduced the number of ova shed in response to hCG. The present study, in conjunction with those of Hasegawa *et al.* (1988) and Nakamura *et al.* (1993), suggests that elevated activin A concentrations secreted from uteri and placenta increase ovarian FSH receptors resulting in follicular recruitment in the presence of the low but unchanging levels of FSH during the second half of pregnancy in the golden hamster. In the present study, however, hysterectomy also decreased FSH secretion. Therefore, lowered FSH secretion may also be responsible for the decrease in ovulation rate after surgery.

It has been demonstrated that atresia destroys all large multilayered and vesicular follicles on the day after parturition (Greenwald 1964). Changes in plasma concentrations of oestradiol-17 β in the present study suggest that all large antral follicles have already become atretic on day 15 of pregnancy although plasma concentrations of activin A are still high. The cause of atresia at the end of pregnancy is still unknown. A number of studies have indicated that activin supports the development of follicles. However, a previous study has demonstrated that an injection of a high dose of activin A into the ovarian intrabursal spaces causes follicular atresia (Woodruff *et al.*

1990). Therefore, activin at high concentrations may induce follicular atresia. Because a very high level of plasma activin coincided with the decline of plasma oestradiol on day 15 of pregnancy, the elevated activin A may be a cause for follicular atresia before parturition.

In the present study, a marked increase in the number of large antral follicles was found during the second half of pregnancy in the golden hamster, although plasma concentrations of FSH and LH were unchanged during this time. The levels of circulating FSH and LH during the second half of pregnancy agree with a previous report (Bast & Greenwald 1974b). The present study also indicated that plasma concentrations of FSH decreased at 6 h after placentectomy and hysterectomy probably due to reduced circulating levels of activin A. Doi *et al.* (1992) demonstrated that exogenous activin A induced an increase in the secretion of FSH from the pituitary gland in the immature rat. Schwall *et al.* (1989) have also reported that daily subcutaneous injections of recombinant human activin A for 3 days to immature female rats caused a marked increase in serum FSH levels. These results suggest that activin A stimulates FSH secretion and maintains the basal levels of FSH in spite of high levels of inhibin A and inhibin B in the second half of pregnancy in the golden hamster.

In conclusion, the present study clearly demonstrates that during the second half of pregnancy in the golden hamster, a large amount of inhibin A and inhibin B are secreted mainly from the ovarian follicles, whereas activin A is secreted mainly from placenta and uteri. Furthermore, a large amount of activin A may be an important factor in the stimulation of follicular development during the second half of pregnancy in the golden hamster.

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References

- Albano RM, Godsavage SF, Huylebroeck D, Van Nimmen K, Isaacs HV, Slack JM & Smith JC 1990 A mesoderm-inducing factor produced by WEHI-3 murine myelomonocytic leukemia cells is activin. *Development* **110** 435–443.
- Bast JD & Greenwald GS 1974a Serum profiles of follicle-stimulating hormone, luteinizing hormone and prolactin during the estrus cycle of the hamster. *Endocrinology* **94** 1295–1299.

- Bast JD & Greenwald GS 1974b Daily concentrations of gonadotropins and prolactin in the serum of pregnant or lactating hamsters. *Journal of Endocrinology* **63** 527–532.
- De Jong FH 1988 Inhibin. *Physiological Reviews* **68** 555–607.
- Doi M, Igarashi M, Hasegawa Y, Eto Y, Shibata H, Miura T & Ibuki Y 1992 *In vivo* action of activin-A on pituitary–gonadal system. *Endocrinology* **130** 139–144.
- Gibori G, Antczak E & Rothchild I 1977 The role of estrogen in regulation of luteal progesterone secretion in the rat after day 12 of pregnancy. *Endocrinology* **100** 1483–1495.
- Greenwald SG 1964 Ovarian follicular development in the pregnant hamster. *Anatomical Record* **148** 605–609.
- Greenwald SG 1965 Histologic transformation of the ovary of the lactating hamster. *Endocrinology* **77** 641–650.
- Greenwald SG 1966 Ovarian follicular development and pituitary FSH and LH content in the pregnant rat. *Endocrinology* **79** 572–578.
- Greenwald SG 1967 Induction of ovulation in the pregnant hamster. *American Journal of Anatomy* **121** 249–258.
- Gu Y, Srivastava RK, Ou J, Krett NL, Mayo KE & Gibori G 1995 Cell-specific expression of activin and its two binding proteins in the rat decidua: role of $\alpha 2$ -macroglobulin and follistatin. *Endocrinology* **136** 3815–3822.
- Hasegawa Y, Miyamoto K, Abe Y, Nakamura T, Sugino H, Eto Y, Shibai H & Igarashi M 1988 Induction of follicle stimulating hormone receptor by erythroid differentiation factor on rat granulosa cell. *Biochemical and Biophysical Research Communications* **156** 668–674.
- Kishi H, Taya K, Watanabe G & Sasamoto S 1995 Follicular dynamics and secretion of inhibin and oestradiol- 17β during the oestrous cycle of the hamster. *Journal of Endocrinology* **146** 169–176.
- Kishi H, Okada T, Otsuka M, Watanabe G, Taya K & Sasamoto S 1996 Induction of superovulation by immunoneutralization of endogenous inhibin through the increase in the secretion of follicle-stimulating hormone in the cyclic golden hamster. *Journal of Endocrinology* **151** 65–75.
- Knight PG, Muttukrishna S & Groome NP 1996 Development and application of a two-site enzyme immunoassay for the determination of total activin-A concentrations in serum and follicular fluid. *Journal of Endocrinology* **148** 267–279.
- Korenman SG, Stevens RH, Carpenter LA, Robb M, Niswender GD & Sherman BM 1974 Estradiol radioimmunoassay without chromatography: procedure, validation and normal values. *Journal of Clinical Endocrinology and Metabolism* **38** 718–720.
- Nakamura M, Minegishi T, Hasegawa Y, Nakamura K, Igarashi S, Ito I, Shinozaki H, Miyamoto K, Eto Y & Ibuki Y 1993 Effect of an activin A on follicle-stimulating hormone (FSH) receptor messenger ribonucleic acid levels and FSH receptor expression in cultured rat granulosa cells. *Endocrinology* **133** 538–544.
- Ohshima K, Kishi H, Itoh M, Watanabe G, Arai K, Uehara K, Groome NP & Taya K 1999 Secretion of inhibin A, inhibin B and inhibin pro- αC during the oestrous cycle of the golden hamster (*Mesocricetus auratus*). *Journal of Endocrinology* **162** 451–456.
- Robertson DM, Foulds LM, Leversha L, Morgan FJ, Hearn FJ, Hearn MT, Burger HG, Wettenhall RE & de Kretser DM 1985 Isolation of inhibin from bovine follicular fluid. *Biochemical and Biophysical Research Communications* **126** 220–226.
- Schwall R, Schmelzer CH, Matsuyama E & Mason AJ 1989 Multiple actions of recombinant activin-A *in vivo*. *Endocrinology* **125** 1420–1423.
- Smith JC, Price BMJ, Van Nimmen K & Huylebroeck 1990 Identification of potent *Xenopus* mesoderm-inducing factor as a homologue of activin A. *Nature* **345** 729–731.
- Steel RGD & Torrie JH 1960 *Principles and Procedures of Statistics*, pp 107–109. New York: McGraw-Hill.
- Taya K 1993 Roles of inhibin in regulation of FSH secretion and folliculogenesis in mammals. *Current Trends in Experimental Endocrinology* **1** 97–116.
- Taya K & Sasamoto S 1977 Induction of ovulation by exogenous gonadotrophin during pseudopregnancy, pregnancy or lactation in the rat. *Journal of Reproduction and Fertility* **51** 467–468.
- Taya K & Watanabe G 1999 Inhibin as a key hormone in determining species-specific ovulation rates in mammals. In *Recent Progress in Molecular and Comparative Endocrinology*, pp 134–143. Eds HB Kwon, JMP Joss & S Ishii. Korea: Hormone Research Center.
- Taya K, Watanabe G & Sasamoto S 1985 Radioimmunoassay for progesterone, testosterone and estradiol- 17β using ^{125}I -iodohistamine radioligands. *Japanese Journal of Animal Reproduction* **31** 186–197.
- Taya K, Komura H, Watanabe G & Sasamoto S 1989 Peripheral blood levels of immunoreactive inhibin during pseudopregnancy, pregnancy and lactation in the rat. *Journal of Endocrinology* **121** 545–552.
- Taya K, Kaneko H, Taketomi T, Kishi H & Watanabe G 1996 Role of inhibin in the regulation of FSH secretion and folliculogenesis in cows. *Animal Reproduction Science* **42** 563–570.
- Vale W, Rivier J, Vaughan J, McClintock R, Corrigan A, Woo W, Karr D & Spiess J 1986 Purification and characterization of an FSH releasing protein from porcine ovarian follicular fluid. *Nature* **321** 776–779.
- Woodruff TK, Lyon RJ, Hansen SE, Rice GC & Mather JP 1990 Inhibin and activin locally regulate rat ovarian folliculogenesis. *Endocrinology* **127** 3196–3205.
- Ying SY 1988 Inhibins, activins, and follistatin: gonadal proteins modulating the secretion of follicle-stimulating hormone. *Endocrine Reviews* **9** 267–293.
- Ying SY, Gove S, Fang VS & Greep RO 1973 Ovulation in postpartum rats. *Endocrinology* **92** 108–116.

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