Isoproterenol is a positive regulator of the suppressor of cytokine signaling-3 gene expression in 3T3-L1 adipocytes

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Abstract

SOCS (suppressor of cytokine signaling)-3 has recently been shown to be an insulin- and tumor necrosis factor (TNF)-α-induced negative regulator of insulin signaling. To further clarify a potential involvement of SOCS-3 in the development of insulin resistance, we measured differentiation-dependent SOCS-3 mRNA expression in 3T3-L1 adipocytes and studied its regulation by various hormones known to impair insulin signaling using quantitative real-time RT-PCR. There was a differentiation-dependent downregulation of SOCS-3 mRNA by 50% over the 9 day adipocyte differentiation course. Interestingly, besides insulin and TNF-α, chronic treatment of differentiated 3T3-L1 cells with 10 µM isoproterenol for 16 h stimulated SOCS-3 gene expression by about 3.5-fold. Furthermore, isoproterenol stimulated SOCS-3 mRNA expression in a dose-dependent manner with significant activation detectable at concentrations as low as 10 nM isoproterenol. Moreover, a strong 27- and 47-fold activation of SOCS-3 mRNA expression could be seen after 1 h of isoproterenol and GH treatment respectively. The stimulatory effect of isoproterenol could be almost completely reversed by pretreatment of 3T3-L1 cells with the β-adrenergic antagonist propranolol. Finally, isoproterenol’s action could be mimicked by stimulation of Gs-proteins with cholina toxin and of adenyl cyclase with forskolin and dibutyryl cAMP. Taken together, our results demonstrate a differentiation-dependent downregulation of SOCS-3 in adipocytes and suggest that SOCS-3 gene expression is stimulated by β-adrenergic agents via activation of a Gs-protein–adenyl cyclase-dependent pathway. As SOCS-3 is a novel inhibitor of insulin signaling, the data support a possible role of this protein as a selectively regulated mediator of catecholamine-induced insulin resistance. Journal of Endocrinology (2002) 175, 727–733

Introduction

Insulin resistance, a diminished biological response of tissues to insulin, is a major risk factor for the development of type 2 diabetes, hypertension and coronary artery disease (Matthaei et al. 2000). At the cellular level, defects of various insulin signaling molecules are frequently observed in insulin resistance and are regarded as a major cause of this pathological condition. Thus, various studies have shown impaired activation of insulin receptor (IR), IR substrates (IRS), phosphatidylinositol (PI)-3 kinase and protein kinase B in insulin resistance and type 2 diabetes mellitus, leading to impaired activation of insulin-induced metabolic end points such as glucose uptake (Kahn & Flier 2000). Over the last couple of years, various circulating molecules such as free fatty acids (Bluher et al. 2001), insulin (Bluher et al. 2001), tumor necrosis factor (TNF)-α (Hotamisligil 1999), catecholamines (Bluher et al. 2000), angiotensin (AT) 2 (Folli et al. 1997), growth hormone (GH) (Takano et al. 2001) and glucocorticoids (Andrews & Walker 1999) have been demonstrated to impair insulin signaling. However, molecules mediating these insulin resistance-inducing effects are only partly understood.

Suppressor of cytokine signaling (SOCS) proteins were originally identified as negative regulators of cytokine action. Several cytokines such as interleukin-2, -3 and -6, and interferon-γ induce SOCS expression in a tissue-specific manner (Endo et al. 1997, Naka et al. 1997, Starr et al. 1997). SOCS proteins modulate cytokine-activated Jak/Stat pathways by binding to either Jak or the cytoplasmic tail of cytokine receptors (Endo et al. 1997, Naka et al. 1997, Starr et al. 1997). Recently, it was demonstrated that insulin and TNF-α stimulate SOCS-3 gene expression in 3T3-L1 adipocytes and that SOCS-3 is increased in insulin resistance and obesity (Emanuelli et al. 2000, 2001). Most interestingly, it was shown that SOCS-3 inhibited insulin signaling (Emanuelli et al. 2000, 2001). Thus, the data accumulated so far suggest that SOCS-3 might be an important novel mediator of cellular events.
insulin resistance and a potential new target in the treatment of insulin resistance and obesity.

It appears possible that various hormones might decrease insulin sensitivity at least partly by upregulating SOCS-3. In the current study, we therefore examined the effect of differentiation and hormones such as isoproterenol, dexamethasone, AT2, GH and triiodothyronine (T3) on SOCS-3 gene expression in 3T3-L1 adipocytes in vitro. We demonstrate for the first time a di

an initial denaturation at 94 °C for 1 s, 61 °C for 7 s, and 72 °C for 11 s. The following primers were used: SOCS-3 (accession no. U88328) CCCTGC AACGCCCTCCTTTTCTAC (sense) and GCCCCCAC CCAGCCCCCATACCC (antisense); 36B4 (accession no. NM007475) AAGC GGCTCTGCGCAT TTGCT (sense) and CCGCAGGGGCCAGGTGGGT (antisense). SYBR Green I fluorescence emissions were monitored after each cycle and mRNA levels of SOCS-3 and 36B4 were quantified by using the second-derivative maximum method of the LightCycler Software (Roche). This method determines the crossing points of individual samples by an algorithm identifying the first turning point of the fluorescence curve. SOCS-3 synthesis was expressed relative to 36B4, which has been frequently used as an internal control due to its resistance to hormonal regulation (Lin et al. 2002). Linearity between total RNA used per reaction and amount of mRNA measured by the LightCycler software was obtained in the range between 5 and 200 ng total RNA for SOCS-3 and 36B4 respectively (data not shown).

To confirm amplification of specific transcripts, melting curve profiles (cooling the sample to 68 °C and heating slowly to 95 °C with continuous measurement of fluorescence) were produced at the end of each PCR. The specificity of the PCR was further verified by subjecting the amplification products to agarose gel electrophoresis.

Materials and Methods

Materials

Isoproterenol, TNF-α, insulin, dexamethasone, AT2, T3 and GH were purchased from Sigma Chemical Co. (St Louis, MO, USA). Cell culture reagents were obtained from Life Technologies, Inc. (Grand Island, NY, USA), oligonucleotides from MWG-Biotech (Ebersberg, Germany).

Culture and differentiation of 3T3-L1 cells

3T3-L1 adipocytes (American Type Culture Collection, Rockville, MD, USA) were cultured and differentiated as described (Fasshauer et al. 2001b,c). Briefly, confluent preadipocytes were cultured for 3 days in DMEM containing 25 mM glucose, 10% fetal bovine serum and antibiotics (culture medium) further supplemented with 1 µM insulin, 0.5 mM isobutylmethylxanthine and 0.1 µM dexamethasone, and for 3 days in culture medium with 1 µM insulin. After additional 3–6 days in culture medium more than 90% of the cells had accumulated fat droplets.

Analysis of SOCS-3 gene expression

SOCS-3 gene expression was measured by quantitative real-time RT-PCR in a fluorescent temperature cycler (LightCycler; Roche Molecular Biochemicals, Mannheim, Germany) as previously described (Fasshauer et al. 2001d). Briefly, total RNA was isolated from 3T3-L1 adipocytes using TRizol (Life Technologies, Inc., Grand Island, NY, USA) and 1 µg RNA was reverse transcribed using standard reagents (Life Technologies). Ten percent of each RT reaction was amplified in a 20 µl PCR containing 3 mM MgCl2, 0.5 µM each primer and 1 X LightCycler DNA Master SYBR Green I mix (Roche). Samples were incubated in the LightCycler for an initial denaturation at 94 °C for 30 s, followed by 40 PCR cycles. Each cycle consisted of 95 °C for 1 s, 61 °C for 7 s, and 72 °C for 11 s. The following primers were used: SOCS-3 (accession no. U88328) CCCTGC AACGCCCTCCTTTTCTAC (sense) and GCCCCCAC CCAGCCCCCATACCC (antisense); 36B4 (accession no. NM007475) AAGC GGCTCTGCGCAT TTGCT (sense) and CCGCAGGGGCCAGGTGGGT (antisense). SYBR Green I fluorescence emissions were monitored after each cycle and mRNA levels of SOCS-3 and 36B4 were quantified by using the second-derivative maximum method of the LightCycler Software (Roche). This method determines the crossing points of individual samples by an algorithm identifying the first turning point of the fluorescence curve. SOCS-3 synthesis was expressed relative to 36B4, which has been frequently used as an internal control due to its resistance to hormonal regulation (Lin et al. 2002). Linearity between total RNA used per reaction and amount of mRNA measured by the LightCycler software was obtained in the range between 5 and 200 ng total RNA for SOCS-3 and 36B4 respectively (data not shown).

Statistical analysis

Results are shown as means ± s.e. For analysis of differences between various treatments, unpaired Student’s t-tests were used. P<0.01 is considered highly significant.

Results

SOCS-3 mRNA expression is inhibited during differentiation

SOCS-3 mRNA expression during differentiation was analyzed. As compared with confluent 3T3-L1 preadipocytes on day 0, SOCS-3 mRNA synthesis was inhibited by about 50% between days 6 and 9 of differentiation (P<0.01) (Fig. 1A). In contrast, SOCS-3 gene expression was not inhibited but rather augmented if NIH-3T3 cells, which do not differentiate, were exposed to the adipocyte differentiation protocol (Fig. 1B).

Isoproterenol and TNF-α are potent stimulators of SOCS-3 gene expression

As SOCS-3 has recently been suggested as a potent negative regulator of insulin signaling, we tested whether various hormones which have been shown to induce insulin resistance might influence SOCS-3 gene expression in 3T3-L1 adipocytes in vitro. Interestingly, treatment
of 3T3-L1 cells with 10 µM isoproterenol and 10 ng/ml TNF-α for 16 h increased SOCS-3 mRNA expression by 3.5- and 2.7-fold respectively, as compared with untreated controls (P < 0.01) (Fig. 2). Furthermore, insulin dramatically and transiently increased SOCS-3 gene expression almost 7-fold by 1 h of stimulation (data not shown). In contrast, AT2 (10 µM), dexamethasone (Dexa, 100 nM), GH (500 ng/ml) and T3 (1 µM) did not significantly influence expression of SOCS-3 (Fig. 2).

Isoproterenol stimulated SOCS-3 gene expression in a dose-dependent fashion. Thus, a significant 1.7-fold increase of SOCS-3 mRNA was detectable at isoproterenol concentrations as low as 10 nM (P < 0.01) (Fig. 3). A maximal 6-fold increase was found at 1 µM of the β-adrenergic agonist (P < 0.01) (Fig. 3). Furthermore, TNF-α induced SOCS-3 dose-dependently with a maximal stimulation detectable at 100 ng/ml effector (data not shown).

Isoproterenol and GH acutely stimulate SOCS-3 gene expression

To determine acute regulation of SOCS-3 mRNA synthesis, 3T3-L1 adipocytes were treated for different periods of time with 10 µM isoproterenol or 500 ng/ml GH. As shown in Fig. 4, maximal 27- and 47-fold activation of SOCS-3 gene expression could be detected 1 h after addition of isoproterenol and GH respectively. Interestingly, SOCS-3 mRNA expression returned to
basal levels after 8 h GH treatment, whereas it remained upregulated in isoproterenol-treated cells (Fig. 4A and B).

**The stimulatory effect of isoproterenol on SOCS-3 mRNA expression is mediated via β-adrenergic receptors, G<sub>S</sub>-proteins and adenylyl cyclase**

To verify that stimulation of SOCS-3 gene expression by isoproterenol is, in fact, mediated via β-adrenergic receptors, fully differentiated 3T3-L1 cells were pretreated with selective antagonists of α- (phenolamine, 100 µM) and β- (propranolol, 100 µM) adrenergic receptors for 1 h before isoproterenol (10 µM) was added for 16 h. SOCS-3 mRNA expression significantly increased 3.8-fold after addition of isoproterenol as compared with untreated control cells ($P<0.01$) (Fig. 5). This increase was completely blocked after pretreatment of 3T3-L1 adipocytes with propranolol ($P<0.01$), whereas phentolamine did not have any effect (Fig. 5). Taken together, these results suggest that isoproterenol stimulates SOCS-3 gene expression via β-adrenergic receptors.

Typically, stimulation of β-adrenergic receptors leads to activation of G<sub>S</sub>-proteins which, in turn, activate adenylyl cyclase. To confirm that the stimulatory effect of isoproterenol on SOCS-3 gene expression is mediated via these signaling molecules, 3T3-L1 adipocytes were treated with cholera toxin (1000 ng/ml), an activator of G<sub>S</sub>-proteins, forskolin (200 µM), a direct stimulator of adenylyl cyclase, and dibutyryl cAMP (100 mM), a stable cAMP-analogue. As shown in Fig. 6, all three effectors significantly activated SOCS-3 gene expression between 2.4- and 3.9-fold ($P<0.01$). These effects were dose-dependent with significant stimulation detectable at concentrations as low as 10 ng/ml cholera toxin, 2 µM forskolin and 10 mM dibutyryl cAMP (data not shown).

**Discussion**

Insulin’s metabolic effects are mediated through the IR. Various substrate molecules including IRS proteins bind to the activated IR and are phosphorylated by its kinase activity (Saltiel & Kahn 2001). Several studies using knock-out mice and cells derived from transgenic animals have demonstrated that activation of IRS proteins is essential for insulin action (Araki et al. 1994, Tamemoto 1995).
Isoproterenol-induced SOCS-3 expression · M FASSHAUER and others

Figure 6 Stimulation of SOCS-3 gene expression by isoproterenol is mediated via Gs-proteins and adenyl cyclase. After 6 h serum-starvation, 3T3-L1 adipocytes were cultured for 16 h with isoproterenol (Iso, 10 μM), cholera toxin (Cholera, 1000 ng/ml), forskolin (For, 200 μM), and dibutyryl-cAMP (cAMP, 100 mM). Total RNA was extracted and quantitative real-time RT-PCR was performed as described in Materials and Methods. SOCS-3 mRNA levels normalized to 36B4 are expressed relative to non-treated control (Co) cells (n=100%). Results are the means ± SE of four independent experiments. **P<0.01 comparing non-treated with isoproterenol-, cholera toxin-, forskolin- and dibutyryl-cAMP-treated cells.

et al. 1994, Withers et al. 1998, Fashshauer et al. 2000, 2001a). Furthermore, it has been demonstrated that insulin resistance is accompanied by decreased activation of several insulin signaling molecules, although the exact mechanisms are not completely understood (Matthaei et al. 2000).

There is growing evidence that catecholamines impair insulin sensitivity and that increased activity of the sympathetic nervous system contributes to insulin resistance (Facchin et al. 1996, Reaven et al. 1996, Landsberg 1999, Hoieggen et al. 2000, Maison et al. 2000). We and others have previously demonstrated impaired insulin signaling and impairment of insulin action after pretreatment of adipocytes with β-adrenergic agents (Klein et al. 1999, 2000). Thus, insulin-induced activation of IR, IRS-1 and PI-3 kinase, as well as insulin-stimulated glucose uptake, were impaired after pretreatment of adipocytes with β3-adrenergic agonists (Klein et al. 1999). Similarly, activation of IR and IRS-1 and insulin-induced glucose uptake were blunted after pretreatment of adipocytes with β-adrenergic agents (Kirsch et al. 1983, Arsenis & Livingston 1986). In a clinical context, our group recently demonstrated that patients with pheochromocytoma are insulin resistant due to increased serum levels of catecholamines and that insulin resistance could be reduced by surgical removal of the tumors in most cases (Bluher et al. 2000). However, the proteins mediating inhibition of insulin signaling after β-adrenergic activation are still unclear. In the present study, we demonstrate for the first time that isoproterenol increases SOCS-3 gene expression in 3T3-L1 adipocytes. Recently, Emanuelli et al. (2000, 2001) demonstrated convincingly that SOCS-3 is a novel negative regulator of insulin signaling. The authors suggested competitive binding of SOCS-3 to phosphotyrosine 960 of the IR, which impairs insulin-induced activation of IRS-1 and Stat5b thereby inducing insulin resistance. Based on these observations, β-adrenergic up-regulation of SOCS-3 may be an important new mechanism by which isoproterenol impairs insulin sensitivity. Moreover, upregulation of SOCS-3 in obesity and insulin resistance might not only be mediated by insulin and TNF-α (Emanuelli et al. 2000, 2001) but also via β-adrenergic activation. Interestingly, Madiehe et al. (2001) recently demonstrated downregulation of SOCS-3 after adrenalectomy, which might at least partly be mediated via decreased levels of catecholamines. In the present study, we show evidence that isoproterenol stimulates SOCS-3 expression via activation of Gs-proteins and adenyl cyclase, in accord with the classical view of β-adrenergic receptors being Gs-protein-coupled (Collins & Survit 2001). GASPERINI et al. (2002) recently demonstrated convincingly that SOCS-3 gene expression was upregulated in leukocytes after cholina toxin, forskolin and dibutyryl-cAMP treatment. Interestingly, inhibition of protein kinase A (PKA) did not reverse dibutyryl-cAMP-induced SOCS-3 synthesis, indicating that increased cAMP levels might mediate their positive effect on SOCS-3 expression independently of PKA. Furthermore, we present evidence that GH acutely and transiently upregulates SOCS-3 mRNA expression in 3T3-L1 adipocytes, consistent with data obtained in 3T3-F442A cells (Adams et al. 1998). Thus, transient upregulation of SOCS-3 expression by GH might be one mechanism by which GH impairs insulin signaling (Takano et al. 2001).

Increased serum levels of AT2, glucocorticoids or thyroid hormones have also been shown to impair glucose tolerance profoundly. At the molecular level, AT2 causes decreased activity of insulin signaling proteins such as IRS molecules, which we and others have shown to be essential for insulin action (Folli et al. 1997, Fashshauer et al. 2000, 2001a). Dexamethasone induces insulin resistance at least partly via downregulation of IRS-1 (Turnbow et al. 1994). Thyroid hormone-induced insulin resistance is accompanied by decreased expression and translocation of the insulin-responsive glucose transporter-4 (Fickova et al. 1997). In our in vitro system, chronic incubation of 3T3-L1 adipocytes with pharmacological doses of dexamethasone, AT2 and T3 does not influence SOCS-3 gene expression. Therefore, our data do not support a major role for SOCS-3 in insulin resistance induced by these hormones; however, we cannot exclude the possibility that they might alter SOCS-3 mRNA expression after different incubation periods or in different cellular systems.

In summary, we demonstrate for the first time that isoproterenol significantly stimulates SOCS-3 gene expression in 3T3-L1 adipocytes. Furthermore, we show evidence that isoproterenol exerts its effects via...
β-adrenergic receptors, Gs-proteins and adenyl cyclase. Moreover, insulin, TNF-α and GH are also positive regulators of SOCS-3 synthesis. These data indicate that upregulation of SOCS-3 is a selectively regulated mechanism that might constitute an important element in the pathogenesis of insulin resistance and the insulin resistance syndrome.

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