Both estrogen and raloxifene cause G1 arrest of vascular smooth muscle cells

K Takahashi1,2, M Ohmichi1,2, M Yoshida1, K Hisamoto2, S Mabuchi2, E Arimoto-Ishida2, A Mori1, S Tsutsumi1, K Tasaka2, Y Murata2 and H Kurachi1

1Department of Obstetrics and Gynecology, Yamagata University School of Medicine, 2-2-2 Iidanishi, Yamagata, Yamagata 990-9585, Japan
2Department of Obstetrics and Gynecology, Osaka University Medical School, 2-2 Yamadaoka, Suita, Osaka 565-0871, Japan

Abstract

The proliferation of vascular smooth muscle cells (VSMC) is a crucial pathophysiological process in the development of atherosclerosis. Although estrogen is known to inhibit the proliferation of VSMC, the mechanism responsible for this effect remains to be elucidated. In addition, the effect of raloxifene on VSMC remains unknown. We have shown here that 17β-estradiol (E2) and raloxifene significantly inhibited the platelet-derived growth factor (PDGF)-stimulated proliferation of cultured human VSMC. Flow cytometry demonstrated that PDGF-stimulated S-phase progression of the cell cycle in VSMC was also suppressed by E2 or raloxifene. We found that PDGF-induced phosphorylation of retinoblastoma protein (pRb), whose hyperphosphorylation is a hallmark of cell cycle G1–S transition, was significantly inhibited by E2 or raloxifene. Since VSMC express both ERα and ERβ, we examined which estrogen receptor (ER) is necessary for these effects of E2 and raloxifene. We obtained in VSMC. Moreover, E2 and raloxifene significantly inhibited the PDGF-induced cyclin D1 promoter activity in A10 cells transfected with cDNA for ERα but not in the parental cells. These results suggested that E2 and raloxifene exert an antiproliferative effect in VSMC treated with PDGF, at least in part through inhibition of pRb phosphorylation, and that the inhibitory effects of E2 and raloxifene may be mainly mediated by ERα.

Introduction

The proliferation of vascular smooth muscle cells (VSMC) is a crucial pathophysiological process in the development of atherosclerosis and of restenosis after coronary angioplasty (Ross 1995, Schwartz et al. 1995, Hanke 1996, Braun-Dullaeus et al. 1998). VSMC are normally quiescent and display contractile properties. This contractility contributes to vasoconstriction and vasodilatation in the regulation of blood pressure and flow volume. Mechanical and/or chemical stresses alter the VSMC phenotype from the contractile to the synthetic state, and synthetic VSMC then migrate into the intima (Ross 1995).

Many epidemiological and basic studies demonstrated that estrogen affords cardioprotection in postmenopausal women (Stampfer et al. 1991, Mendelsohn & Karas 1999). Although the results of the Women’s Health Initiative’s (WHI) large prospective randomized controlled study seem to raise questions about the cardioprotective effect of estrogen (Writing Group for the Women’s Health Initiative Investigators 2002), interim reports on women on estrogens alone did not show adverse effects on cardiovascular disease or breast cancer, and this part of the trial continues. Thus, the cardioprotective effect of estrogen itself was not necessarily disproven by the results of the WHI study. Moreover, there is a need for an ideal estrogen that does not increase the risk of breast cancer (Cummings et al. 1999). Raloxifene is a non-steroidal benzothiophene that has been classified as a selective estrogen receptor modulator based on the fact that it produces both estrogen-agonistic effects on bone and lipid metabolism and estrogen-antagonistic effects on uterine endometrium and breast tissue. Although we found that raloxifene rapidly induces nitric oxide synthase activation in vascular
endothelial cells (Hisamoto et al. 2001a), its ability to protect against cardiovascular disease has yet to be demonstrated and is now being tested in the Raloxifene Use for the Heart trial.

Several researchers have recently focused on the direct effects of estrogen on the endothelium and VSMC (Sullivan et al. 1995, Hisamoto et al. 2001b). We previously reported that estrogen rapidly induces nitric oxide synthase activation in vascular endothelial cells (Hisamoto et al. 2001b). Nitric oxide is secreted from normal endothelium and has important roles in maintaining normal vasculature, such as relaxation of vascular smooth muscle and inhibition of platelet activation (Moncada & Higgs 1993). Platelet-derived growth factor (PDGF) is released from platelets, macrophages and endothelial cells, and is important in triggering VSMC proliferation and migration (Ross 1986, Schwartz et al. 1995). Some investigators have reported that estrogen inhibits PDGF-induced proliferation and migration of VSMC in vitro (Dai-Do et al. 1996, Kolodgie et al. 1996, Suzuki et al. 1996, Somjen et al. 1998). However, the mechanism responsible for this effect is not yet fully understood.

Recently, VSMC proliferation has been reported to be inhibited by a reduction of retinoblastoma protein (pRb) phosphorylation and by modulation of cell cycle regulators (Kintscher et al. 2000, Marra et al. 2001). The status of pRb phosphorylation is critical for cell proliferation. In quiescent cells, pRb is present in a hypophosphorylated state in which it is able to bind to and sequester members of the E2F family of transcription factors (Weinberg 1995). Phosphorylation of pRb at multiple sites induces a conformational change that releases E2F, which in turn activates the transcription of genes required for further cell cycle progression (Nevins 1992, La Thangue 1996). Therefore, we hypothesized that the antiproliferative effect of estrogen in VSMC might be due to inhibition of pRb phosphorylation.

We report here that 17β-estradiol (E2) exerts an anti-proliferative effect through inhibition of pRb phosphorylation in PDGF-induced VSMC, and that the inhibitory effect of estrogen is mainly mediated by estrogen receptor α (ERα).

Materials and Methods

Materials

Raloxifene analogue LY117018 was a kind gift from Eli Lilly Research Laboratories (Indianapolis, IN, USA). E2 and propidium iodide were purchased from Sigma Chemical Co. (St Louis, MO, USA). ICI 182,780 was obtained from Toyo Inc. (Tokyo, Japan). Rabbit polyclonal anti-phospho pRb Ser807/811 was obtained from PharMingen (San Diego, CA, USA). Mouse anti-pRb antibody was obtained from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). The PicaGene dual sea pansy system was purchased from Toyo Inc. (Tokyo, Japan).

Constructs

The human ERα expression vector, pIE, was a kind gift from Dr D J Shapiro (University of Illinois, Urbana, IL, USA). The cyclin D1 (D1 pro-1749) promoter plasmid construct, which contains the cyclin D1 regulatory region (−1749 to +135) fused to a luciferase reporter gene, was kindly provided by Dr T Shiozawa (Shinshu University School of Medicine, Matsumoto, Japan).

Cell culture

Primary cultures of human aortic smooth muscle cells (VSMC) (Clonetics, Walkersville, MD, USA) and primary rat aortic smooth muscle cells (A10) (American Type Culture Collection, Rockville, MD, USA) were grown at 37 °C in Dulbecco’s modified Eagle’s medium (DMEM) (Life Technologies, Rockville, MD, USA) supplemented with 10% fetal bovine serum (FBS), 100 units/ml penicillin and 100 µg/ml streptomycin. Cultures were maintained at 37 °C in a water-saturated atmosphere of 95% air and 5% CO2, VSMC cultured for at most ten passages were used for the following studies.

Cell proliferation assay

Five thousand cells were seeded per well in a 96-well plate, incubated for 24 h in DMEM with 10% FBS, and then starved in phenol red-free DMEM with 0.4% charcoal–dextran–treated FBS for 48 h. E2 (10−6 to 10−8 M) or vehicle was added 1 h before VSMC or A10 were challenged with PDGF for 48 h. Cell proliferation was assessed by measuring the absorbance at 490 nm of the dissolved formazan product formed after addition of 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium inner salt (MTS) and incubation for 1 h as described by the manufacturer (Promega). All experiments were carried out in quadruplicate and proliferation was expressed as the ratio of the absorbance of treated cells to that of untreated quiescent cells cultured in 0.4% FBS. For cell number experiments, VSMC were plated at a density of 5 × 103 cells per well in a 24-well plate and allowed to attach overnight. The cells were starved as described above for 48 h, and were then
treated with fresh preparation of medium containing E2 or raloxifene every 48 h for 6 days, and counted each time the medium was exchanged. A Neubauer chamber was used to count the cell number and the trypan-blue exclusion test was carried out to determine the cell viability.

**Western blotting analysis**

VSMC were starved as described above for 48 h and then treated with the various agents indicated in the text. They were then washed twice with phosphate-buffered saline (PBS) and lysed in ice-cold lysis buffer (20 mM Tris, pH 7.4, 150 mM NaCl, 1% Triton X-100, 1 mM EDTA, 1 mM EGTA, 2.5 mM sodium pyrophosphate, 1 mM β-glycerol-phosphate, 1 mM sodium orthovanadate, 1 µg/ml leupeptin and 1 mM phenylmethylsulfonyl fluoride). The extracts were centrifuged to remove cellular debris, and the protein content of the supernatants was determined using the Bio–Rad protein assay reagent. Lysate samples containing 40 µg protein were boiled and the proteins were separated by electrophoresis on a 10% glycine–SDS–polyacrylamide gel and electroblotted onto a polyvinylidene difluoride membrane (Hybond-P; Amersham Pharmacia Biotech, Amersham, Bucks, UK). The membrane was incubated with blocking buffer for 1 h at room temperature, probed with primary antibody with gentle rocking at 4°C overnight, and incubated with horseradish peroxidase-conjugated anti-mouse or anti-rabbit secondary antibody (at 1:2000 dilution) for 1 h at room temperature. The signal was detected by chemiluminescence with ECL Western blotting detection reagents (Amersham Pharmacia Biotech). All Western blot experiments were repeated at least three times with identical results.

**Cell cycle distribution**

Quiescent VSMC were pretreated for 1 h with 10⁻⁸ M E₂ or vehicle (ethanol), and then treated with 5 ng/ml PDGF. After 24 h, cells were trypsinized, centrifuged at 1200 r.p.m. for 5 min, washed with PBS, and then treated with RNase A (20 µg/ml). DNA was stained with propidium iodide (100 µg/ml) for 30 min at 4°C in the dark. The DNA content of the cells was analyzed using a Becton Dickinson FACSCalibur flow cytometer (Becton Dickinson, San Jose, CA, USA). DNA histogram analysis was performed using ModFitLT software (Becton Dickinson).

**RT-PCR experiments**

Total RNA was extracted from VSMC and A10 cells with Tri-Reagent (Molecular Research Center, Inc., Cincinnati, OH) according to the manufacturer’s instructions. Total RNA (5 µg) was used as a template for
Figure 2. Effects of E₂ and raloxifene on the phosphorylation of pRb. Quiescent VSMC were pretreated with 10⁻⁸ M E₂ (A) or 10⁻⁸ M raloxifene (Ral) (B) for 1 h before treatment with PDGF (5 ng/ml) for 24 h. Control VSMC were cultured in growth medium (DMEM with 10% FBS). To analyze pRb phosphorylation, extracted proteins were resolved by 8% SDS-PAGE and subjected to Western blotting with anti-pRb antibody (A, upper panel) or antiphospho pRb ser807/811 antibody (B, upper panel). (C) VSMC were pretreated with 1 μM ICI 182,780 (ICI) or vehicle for 15 min, and then treated with 10⁻⁸ M E₂ or vehicle for 1 h. The cells were then treated with 5 ng/ml PDGF for 24 h. Extracted proteins were subjected to 8% SDS-PAGE followed by Western blotting with antiphospho pRb ser807/811 antibody. The positions of the hypophosphorylated (pRb) and hyperphosphorylated (ppRb) forms of pRb protein are indicated on the right. β-Actin analyzed similarly by immunodetection with anti-β-actin antibody served as a loading control (A–C, lower panel). Relative densitometric units of the ppRb bands are shown in the right panels, with the density of the control bands set arbitrarily at 1·0 (A–C). Data are shown as the means ± s.e.m. from at least three separate experiments. *P<0·05.
first-strand cDNA synthesis, cDNA was amplified using
an RNA PCR kit obtained from TaKaRa (Tokyo, Japan)
according to the manufacturer’s instructions. Primers
designed based on the published gene sequences were
obtained from Amersham Pharmacia Biotech Japan
(Tokyo, Japan) and Hitachi Instruments Service Co. Ltd
(Tokyo, Japan). The sequences of the PCR primers were
as follows: rat ERα: 5′-AATTCTGACAATCGACGC
CAG-3′ (sense), 5′-GTGCTTCAACATTCTCCTCC
TC-3′ (antisense); rat ERβ: 5′-TTCCGCAGCAC
CAGTAACC-3′ (sense), 5′-TCCGCCCTTTGCGTCTG
GACTA-3′ (antisense); human ERα: 5′-AATTCAGAT
AATCGACGCCAG-3′ (sense), 5′-GTGTTTCAACAT
TCTCCCTTCTCC-3′ (antisense); human ERβ: 5′-GTTC
CGCCGACCCCTGTATTAC-3′ (sense), 5′-CTCGTCG
GCACCTTTCTGTCTC-3′ (antisense). To check the
cDNA integrity, glyceraldehyde-3-phosphate dehydro-
genase (GAPDH) cDNA was amplified in parallel. All
transcripts were analyzed using a GeneAmp PCR System
9700 (Applied Biosystems, Foster City, CA, USA). The
amplification profile consisted of 35 cycles of denaturation
at 94 °C for 1 min, primer annealing at 59 °C for 1 min,
and extension at 72 °C for 1 min. PCR products were
electrophoresed on a 1.5% agarose gel and visualized by
ethidium bromide staining under UV illumination.

Cell transfections and reporter gene assays
Approximately 2 × 10^5 A10 cells were plated per well of a
six-well tissue culture plate 24 h before transfection. For
analysis of the effects of estrogen on reporter gene activity,
transfection complexes containing a total of 1.55 µg plas-
mids (1 µg D1 pro–1749, 0.5 µg pHE and 0.05 µg pRL–
CMV (RoYo Ink, Tokyo, Japan) as an internal control) were
added to each well of A10 cells as recommended by the
manufacturer (Invitrogen). After 3 h, the transfection
cocktail was replaced with growth medium, and the cells
were cultured for 24 h. The growth medium was replaced
with DMEM containing charcoal–dextran-stripped 0.4%
FBS to induce the A10 cells to become quiescent and the
cells were cultured in this medium for 24 h. The quiescent
A10 cells were pretreated for 1 h with 10^{-7} M E_2 or
vehicle (ethanol), and then treated with 5 ng/ml PDGF.
After 24 h, the cells were harvested and luciferase assays
were performed with the PicaGene dual sea pansy system.
Firefly-luciferase activity and sea pansy-luciferase activity
were measured as relative light units with a lumino-
imeter (Lumat LB9507; EG&G, Berthold, Bad Wildbad,
Germany). The firefly-luciferase activity was then normal-
ized relative to the sea pansy-luciferase activity to deter-
mine the transfection efficiency.

Statistics
Statistical analysis was performed by Student’s t-test, and
P<0.05 was considered significant. Data are expressed as
the means ± S.E.M.

E_f and raloxifene reduce pRb phosphorylation in VSMC
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Results
Effects of E_2 and raloxifene on VSMC proliferation
The effects of raloxifene and E_2 on the proliferation of
VSMC were evaluated (Fig. 1A). Both 10^{-8} M E_2 and
10^{-8} M raloxifene significantly inhibited the growth of
VSMC. Pathological proliferation of VSMC plays a major
role in the development of atherosclerotic lesions (Braun-
Dullaeus et al. 1998). PDGF released from platelets,
macrophages and endothelium is a major trigger of VSMC
proliferation. Therefore, we evaluated the effect of E_2 or
raloxifene on the proliferation of PDGF-stimulated
VSMC. When quiescent VSMC were treated with 5 ng/ml
PDGF in 0.4% FBS for 48 h, the proliferation of the
VSMC was stimulated by about 50% compared with
that of the quiescent VSMC. PDGF-stimulated VSMC
proliferation was significantly inhibited by E_2 in a
concentration-dependent manner in the range of 10^{-8} to
10^{-6} M (Fig. 1B) and also by raloxifene (data not shown).
We also determined the effect of E_2 or raloxifene on
PDGF-induced cell cycle progression by flow cytometry.
Subconfluent VSMC were synchronized by culturing
with 0.4% FBS for 48 h, pretreated with E_2 or vehicle,
and then treated with 5 ng/ml PDGF. Both 10^{-8} M E_2 (Fig.
1C) and 10^{-8} M raloxifene (data not shown) reduced the
proportion of VSMC that progressed to S phase.

Effects of E_2 and raloxifene on hyperphosphorylation of pRb
Because cell cycle progression is regulated by hyperphos-
horylation of pRb, we examined the effects of E_2 and
raloxifene on pRb hyperphosphorylation. In quiescent
cells, most pRb is hypophosphorylated (Weinberg 1995).
Hyperphosphorylation of pRb is required for S-phase
progression in most mammalian cells (Weinberg 1995).
Anti-pRb antibody, which recognizes both hyper- and
hypophosphorylated Rb protein, was used to assess the
phosphorylation status at Ser807/811 of pRb, we examined the
effects of E_2 and raloxifene on pRb hyperphosphorylation.
In quiescent cells, most pRb is hypophosphorylated (Weinberg 1995).
Hyperphosphorylation of pRb is required for S-phase
progression in most mammalian cells (Weinberg 1995).
Anti-pRb antibody, which recognizes both hyper- and
hypophosphorylated Rb protein, was used to assess the
phosphorylation status of pRb. Hyperphosphorylated pRb
(ppRb) was increased in cells cultured with 5 ng/ml
PDGF (Fig. 2A, lane 3) as well as in cells cultured with
10% FBS (Fig. 2A, lane 1) for 24 h. E_2 clearly attenuated
the PDGF-induced increase of ppRb (Fig. 2A, lane 4). To
investigate whether raloxifene also decreases the PDGF-
induced ppRb, phospho-specific antibody was used to
assess the phosphorylation status at Ser807/811 of pRb,
which is important for cdk-dependent function. ppRb was
increased in cells cultured with 5 ng/ml PDGF (Fig. 2B,
lane 3) and raloxifene clearly decreased the PDGF-
induced increase of ppRb (Fig. 2B, lane 3). To investigate
whether the inhibitory effect of E_2 or raloxifene on pRb
phosphorylation is ER-mediated, the effect of ICI
182,780, a potent ER antagonist, was examined.
Antiphospho-specific pRb Western blotting analysis
showed that 1 µM ICI 182,780 clearly attenuated the
inhibitory effects of E_2 (Fig. 2C, lane 4) and raloxifene

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(data not shown) on the PDGF-induced hyperphosphorylation of pRb. This result suggested that ER is involved in the antiproliferative effects of E2 and raloxifene in PDGF-stimulated VSMC.

Effects of E2 and raloxifene on the protein expression of G1 regulators

The phosphorylation of pRb is catalyzed, at least in part, by the D-type cyclins and their associated catalytic partners, cdk4 and cdk6, as well as by cyclin E and its catalytic partner, cdk2. We therefore examined the effects of E2 and raloxifene on the expression of cyclin D1 (Fig. 3). Treatment of cells with 5 ng/ml PDGF induced the expression of cyclin D1 (Fig. 3A and B, lane 2). E2 at $10^{-8}$ M (Fig. 3A, lane 4) and raloxifene at $10^{-8}$ M (Fig. 3B, lane 4) significantly decreased the PDGF-induced expression of cyclin D1. On the other hand, neither E2 nor raloxifene affected the PDGF-induced expression of cdk4 (data not shown). Induction of cdk inhibitors has been shown to accompany cell cycle arrest in response to various antiproliferative stimuli. Therefore, the effect of E2 on the level of p27kip1 was examined. Neither E2 nor raloxifene had any effect on the expression of p27kip1 (data not shown).

Antiproliferative effects of E2 and raloxifene are mediated by ERα

Next, we examined which ER is necessary for these effects of E2. RT-PCR analysis showed that VSMC expressed both ERα and ERβ mRNAs (Fig. 4). Although we confirmed that rat uterus expressed ERα (data not shown), A10 cells, which are derived from the thoracic aorta of embryonic rats and are a commonly used model of VSMC (Weiss et al. 2000), expressed ERβ but not ERα (Fig. 4). We therefore used A10 cells to examine whether or not ER is involved in the antiproliferative effect of E2. A10 cells that had been induced to enter a quiescent state by incubation in low-serum medium proliferated in response to PDGF stimulation in a dose-dependent manner (Fig. 5A). In contrast to the effect of E2 (Fig. 1B) and raloxifene in VSMC, E2 (Fig 5B) and raloxifene (data not shown) did not significantly inhibit the PDGF-induced proliferation of A10 cells, suggesting that the growth inhibition by estrogen and raloxifene may not be mediated by ERβ.

We next sought to determine whether E2 and raloxifene inhibit the PDGF-induced cyclin D1 promoter activity. A cyclin D1 promoter (~1749 to +135 bp)–luciferase reporter construct was transiently transfected into A10 cells. As shown in Fig. 6, addition of 5 ng/ml PDGF for 24 h enhanced the luciferase activity. To examine whether the inhibition of the PDGF-induced cyclin D1 promoter activity by E2 or raloxifene is mediated by ERα, A10 cells were transiently transfected with A10 cells.
with an ERα expression vector in addition to the cyclin D1 reporter construct. Although E2 and raloxifene had no effect on the PDGF-induced cyclin D1 promoter activity in A10 cells, transfection of the ERα expression vector rendered the PDGF-induced cyclin D1 promoter activity sensitive to inhibition by E2 and raloxifene (Fig. 6). These results indicated that ERα is required for the inhibition of the PDGF-induced cyclin D1 promoter activity by E2 and raloxifene.

**Discussion**

We have demonstrated here that E2 and raloxifene inhibit PDGF-induced VSMC proliferation, at least in part by reducing the PDGF-induced pRb phosphorylation. Protective effects of estrogen against cardiovascular disease in postmenopausal women are well documented (Forrester et al. 1991, Belchetz 1994). Putative mechanisms of these effects involve effects on endothelium-dependent and -independent vasodilation (Williams et al. 1990, 1992, Jiang et al. 1991, Rosano et al. 1993, Gilligan et al. 1994) and improvement in the lipid profile (Belchetz 1994). Estrogen stimulates endothelial nitric oxide synthase (eNOS) activity in vivo (Weiner et al. 1994) and in vitro (Hisamoto et al. 2001b), and inhibits smooth muscle cell proliferation by increasing the endothelium-derived nitric oxide (Garg & Hassid 1989). Proliferation and migration of VSMC are believed to significantly contribute to intimal thickening in atherosclerosis and venous bypass graft disease (Calcagno et al. 1992). PDGF is known to stimulate both proliferation and migration of VSMC, and the importance of this growth factor in the pathophysiology of intimal hyperplasia has been established. The mechanism of VSMC proliferation and migration is related to the stimulation of mitogen-activated protein kinase (MAP kinase) (Claesson-Welsh 1994, Graves et al. 1996), and the resultant increase of the transcription of many immediate early genes (Chen et al. 1992, Seth et al. 1992), including cyclin D1, which is required for transition from the G0 to the G1 phase in the cell cycle (Abrieu et al. 1996, Lavoie et al. 1996). Although phytoestrogen and estrogen inhibit VSMC proliferation by reducing MAP kinase activity (Dubey et al. 1999, 2000, Hwang et al. 2002), the mechanism of this inhibition by estrogen is not fully understood.

Mitogen-induced VSMC proliferation is inhibited by numerous agents such as sodium salicylate (Marra et al. 2000), doxazosin (Kintscher et al. 2000), retinoids (Wakino et al. 2001), 3-hydroxy-3-methylglutaryl co-enzyme A (HMG-CoA) reductase inhibitor (Yang et al. 2000), parathyroid hormone-related protein (Stuart et al. 2000) and dexamethasone (Reil et al. 2000). Studies of the inhibitory effects of these agents have mainly focused on the inhibition of pRb phosphorylation. pRb is hypophosphorylated in quiescent cells and binds to transcriptional factor E2F. E2F activates the transcription of genes required for S-phase DNA synthesis, including the genes for thymidine kinase dihydrofolate reductase, cell division cycle 2 and cyclin A (Nevins 1992, La Thangue 1996). Thus, pRb phosphorylation is among the most crucial steps regulating progression of the cell cycle. In this study, E2 and raloxifene significantly inhibited the PDGF-stimulated pRb phosphorylation (Fig. 2). The inhibitory effect of E2 on pRb phosphorylation was significant and similar to the effects reported for various other agents that inhibit the proliferation of VSMC (Kintscher et al. 2000, Marra et al. 2000, Yang et al. 2000, Wakino et al. 2001). In addition, some reports have shown that E2 inhibited [3H] thymidine incorporation into PDGF-stimulated VSMC and decreased [3H]thymidine uptake rates to approximately 60–70% of the control rates (Suzuki et al. 1996, Dubey et al. 2000). Collectively, the findings of the present study together with the published data indicate
that the inhibitory effect of E2 on S-phase transition is partial rather than complete in VSMC.

We investigated whether or not E2-induced G1 arrest was dependent upon ER. Pretreatment with ICI 182,780, a specific ER antagonist, restored the status of pRb phosphorylation from the hypophosphorylated to the hyperphosphorylated state (Fig. 2C). This result indicated that the inhibitory effect of E2 on pRb phosphorylation was mediated by ER. The expression of two ER subtypes, ERα and ERβ, is observed in VSMC; however, which subtype is responsible for the atheroprotective effect of E2 has not hitherto been determined. A10 cells, a primary rat embryonic thoracic aorta cell line, have commonly been used as a model of VSMC. We found that A10 cells expressed ERβ, but not ERα (Fig. 4). Although the proliferation of A10 cells was increased by PDGF similarly to that of VSMC, E2 could not inhibit the PDGF-induced proliferation of A10 cells (Fig. 5). Moreover, although E2 did not inhibit the PDGF-induced cyclin D1 promoter activity in A10 cells, E2 significantly inhibited the PDGF-induced cyclin D1 promoter activity in A10 cells transfected with ERα (Fig. 6). These results suggested that the inhibitory effect of E2 on the PDGF-induced proliferation of VSMC was mediated by ERα. However, the fact that ERβ is the predominantly expressed form of ER in VSMC suggests that the protective effects of estrogens in the cardiovascular system may be due to the genomic effects of ERβ in vascular tissue (Hodges et al. 2000). In addition, it was reported that estrogen inhibited VSMC proliferation in response to vascular injury in knockout mice lacking ERα (Iafrati et al. 1997). However, ERα may be involved in the protective effects because this knockout mouse expressed a variant ERα (Kos et al. 2002). Our data are in agreement with the recent finding that ERα mediates the protective effects of estrogen against vascular injury (Pare et al. 2002).

Estrogen is now believed to exert rapid membrane effects independently of the classical gene activation pathway of steroid action. Recently we (Hisamoto et al. 2001a,b) and other groups (Haynes et al. 2000, Simoncini et al. 2000) have shown that estrogen induces the activation of eNOS in a non-genomic manner in vascular endothelial cells. In VSMC, inhibition of Ca2+ influx via L-type Ca2+ channels by estrogen is one of estrogen’s acute, non-genomic vasoconstrictor actions (Ruehlmann et al. 1998), estrogen inhibits lysophosphatidylcholine-induced proliferation via a non-genomic antioxidant mechanism.

Figure 5 E2 does not inhibit PDGF-stimulated proliferation of A10 cells. A10 cells were stimulated with various concentrations of PDGF (A) or pretreated with various concentrations of E2 for 1 h followed by PDGF (5 ng/ml) stimulation (B). Cell proliferation is expressed as a percentage of that of A10 cells without PDGF stimulation. Data are shown as the means ± S.E.M. from at least three separate experiments. *P<0.05 vs A10 cells without PDGF stimulation.

Figure 6 Cyclin D1 promoter activity is suppressed by E2 or raloxifene (Ral) via ERα. A10 cells were transfected with a plasmid containing a cyclin D1 promoter–luciferase gene fusion construct (D1 pro-1749) together with an ERα expression vector (pIE) (+) or vector alone (−) and pRL-CMV internal control plasmid, as described in Materials and Methods. Transfected cells were pretreated with 10−8 M E2 or 10−8 M raloxifene for 1 h and then stimulated with 5 ng/ml PDGF for 24 h. Firefly-luciferase activity, normalized by sea pansy-luciferase activity, was expressed as a percentage of the activity obtained in the absence of E2 or raloxifene in cells transfected with pIE or vector alone. Data are shown as the means ± S.E.M. from at least three separate experiments. *P<0.05.


**References**


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(Yoon et al. 2001), and estrogen inhibits angiogenins II-induced proliferation via the activation and induction of phoshatases through non-genomic as well as genomic signaling (Takeda-Matsubara et al. 2002). Moreover, it was reported that in MCF-7 cells estrogen rapidly stimulates a p85-regulated phoshatylinositol kinase and Akt, and increases the expression and promoter activity of cyclin D1 as well as the entry of cells into S phase (Castoria et al. 2001). We recently found that E2- and raloxifene-induced inhibition of VSMC growth is in part due to induction of apoptosis through the p38 cascade with a non-genomic mechanism (authors’ unpublished data). However, it remains unclear whether the inhibitory effects of estrogen on the PDGF-induced cyclin D1 promoter activity, pRb phosphorylation and S-phase entry observed in the present study are mediated in a genomic or non-genomic manner. These inhibitory effects of E2 and raloxifene were not as rapid as the effects of estrogen and raloxifene on eNOS activation (Hisamotto et al. 2001a,b). Moreover, actinomycin D, an inhibitor of gene transcription, was tested to rule out the influence of non-genomic events, and it inhibited the effects of E2 and raloxifene observed in the present study (data not shown). We therefore consider it likely that the effects of E2 and raloxifene which we observed here mainly depend on the genomic effects of estrogen mediated by ERα.

Although estrogen enhances the proliferation of breast or uterine cells and has anti-apoptotic effects in vascular endothelial cells (Razandi et al. 2000), estrogen inhibits the proliferation of VSMC. In addition, it has been reported that estrogen induces apoptosis in an erythroid cell line (Blobel & Orkin 1996) and induces apoptosis and G1 cell cycle arrest of human multiple myeloma cells (Treon et al. 1998, Wang et al. 2001). Thus, E2 modulates diverse cell functions in a cell- and tissue-specific manner. Although the mechanism of this heterogeneity of estrogen’s actions remains unknown, it is possible that it involves ‘cross-talk’ between membrane-mediated events and nuclear receptor activation.

VSMC proliferation is regulated by direct cell cycle-specific effects as well as indirect effects via the regulation of VSMC mitogen production by endothelial cells (Kourembanas et al. 1998). Thus, it is possible that the inhibitory effects of E2 and raloxifene on VSMC proliferation are mediated by direct cell cycle-specific effects. It was reported recently that VSMC express telomerase activity when stimulated to proliferate (Minamino & Kourembanas 2001). We are therefore now investigating the effects of E2 and raloxifene on telomerase activity.


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