Testosterone as a protective factor against atherosclerosis – immunomodulation and influence upon plaque development and stability

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Abstract

Inflammation plays a central pathogenic role in the initiation and progression of coronary atheroma and its clinical consequences. Cytokines are the mediators of cellular inflammation and promote local inflammation in the arterial wall, which may lead to vascular smooth muscle apoptosis, degradation of the fibrin cap and plaque rupture. Platelet adhesion and thrombus formation then occur, resulting clinically in unstable angina or myocardial infarction. Recent studies have suggested that cytokines are pathogenic, contributing directly to the disease process. 'Anti-cytokine' therapy may, therefore, be of benefit in preventing or slowing the progression of cardiovascular disease. Both oestrogens and testosterone have been shown to have immune-modulating effects; testosterone in particular appears to suppress activation of pro-inflammatory cytokines. Men with low testosterone levels are at increased risk of coronary artery disease. An anti-inflammatory effect of normal physiological levels of sex hormones may, therefore, be important in athero-protection. In this article, we discuss some of the mechanisms involved in atherosclerotic coronary artery disease and the putative link between testosterone deficiency and atheroma formation. We present the hypothesis that the immune-modulating properties of testosterone may be important in inhibiting atheroma formation and progression to acute coronary syndrome.

Cytokines in coronary artery disease

It has become clear in recent years that the clinical events of the acute coronary syndromes in patients with coronary artery disease (CAD) result from a change from quiescent atheroma to ruptured plaque in which inflammation within the vessel wall is intimately associated (Libby et al. 2002). Patients with CAD have elevated circulating levels of cytokines and C-reactive protein (CRP) (Ikonomidis et al. 1999), with higher levels present in those with unstable angina or acute myocardial infarction (Manten et al. 1998). The CRP level, a marker of cytokine activation, is not only a strong independent predictor of mortality and subsequent events in patients admitted to hospital with an acute infarct (Tommasi et al. 1999) but is the best predictor of clinically significant atherosclerotic disease in unselected ‘healthy’ populations (Ridker et al. 1997). It was previously thought that the degree of coronary artery narrowing was the main predictor of subsequent events, but angiographic studies have shown that most infarcts occur in vessels with stenoses of less than 70% (Falk et al. 1995). Thus, the clinical manifestations of coronary atheroma are not directly related to the local burden of atheroma but to the amount of plaque inflammation. Moreover, statins reduce plasma lipid levels and coronary events and alter plaque constituents but have only a minimal effect on the degree of stenosis (Jukema et al. 1995). It would appear, therefore, that it is the quality and not the quantity of plaque which is most important.

The initial process of atheroma formation involves accumulation of cholesterol in the arterial wall and expression of adhesion molecules and chemokines by endothelial cells, in particular monocyte chemoattractant protein-1 (MCP-1) (Reckless et al. 1999). As a consequence, monocytes and macrophages infiltrate the vessel wall and produce inflammatory cytokines including tumour
In vivo synthesis of connective tissue and inhibits smooth muscle are closely regulated: TNF and interleukin (IL)-1 are produced by inflammatory cells within the lesion and secrete proteinases (MMPs) such as collagenases and gelatinases. These enzymes digest and break down plaque matrix and produce collagen and elastin to bolster the lumen by a fibrin cap. Smooth muscle cells migrate into the plaque and produce collagen and elastin to bolster the lumen by a fibrin cap. Smooth muscle cells migrate into the plaque and produce collagen and elastin to bolster the lumen by a fibrin cap. Smooth muscle cells migrate into the plaque and produce collagen and elastin to bolster the lumen by a fibrin cap.

Activated macrophages produce matrix metalloproteinase (MMP) TIMP, tissue inhibitor of matrix metalloproteinase (MMP), monocyte chemoattractant protein, a central mediator of disease (see Table 1). These factors, along with oxidised low-density-lipoprotein (LDL) cholesterol promote further MCP-1 expression and amplify the production of cytokines by macrophages and endothelial cells (Terkeltaub et al. 1998, Krishnaswamy et al. 1999) (Fig. 1).

Within the plaque, a lipid core is separated from the lumen by a fibrin cap. Smooth muscle cells migrate into the plaque and produce collagen and elastin to bolster the cap and extracellular matrix. The stability of the fibrin cap is affected by continuing inflammation; T lymphocytes within the lesion secrete γ-interferon, which reduces the density of connective tissue and inhibits smooth muscle cell proliferation, serving to weaken the cap (Warner et al. 1989). Activated macrophages produce matrix metalloproteinases (MMPs) such as collagenases and gelatinases. These enzymes digest and break down plaque matrix and are closely regulated: TNF and interleukin (IL)-1β promote their release while anti-inflammatory cytokines such as IL-4 and IL-10 reduce MMP activity (Saren et al. 1996, George 1998). The balance of stimulatory and inhibitory cytokines is crucial to the stability of the plaque. Degradation of the fibrin cap may lead to erosion with consequent breach of endothelium and exposure of collagen or to the catastrophic breakdown of the plaque, ‘plaque rupture’, with release of the lipid core and tissue factor. Plaque erosion and rupture are responsible for most acute coronary syndromes. Myocardial infarction is caused by thrombosis in the coronary artery as a complication of plaque breakdown in 90% of occasions. Despite recent advances in treatment, the case fatality rate for myocardial infarction is still high with about 50% of patients dying (Tunstall-Pedoe et al. 1999). Long-term survivors in the absence of cardiac failure have a better prognosis but still have an average 3% annual risk of a vascular event, in most cases because of complications from atherosclerotic plaque (Haq et al. 1995).

The stability of the fibrous cap may be threatened when the rate of extracellular matrix breakdown exceeds the rate of synthesis. This may occur as a consequence of excess cytokine-induced production of MMPs or due to increased rate of vascular smooth muscle cell apoptosis. Rupture of the fibrous cap most often occurs where the layer of vascular smooth muscle cells is thinnest, highlighting their importance (Davies & Thomas 1985). The relative rates of vascular smooth muscle cell proliferation and apoptosis are critical in governing plaque stability (Fig. 2). Vascular smooth muscle cells taken from plaques show higher rates of apoptosis (Geng & Libby 1995), have senescent morphology (Orekhov et al. 1984) and are more sensitive to apoptotic stimuli than smooth muscle cells from normal vascular tissue (Bennett et al. 1995). The common areas of plaque rupture - the shoulders of the plaque - have the highest density of apoptotic vascular smooth muscle cells. Once again, inflammatory cells and their cytokines are the key. Macrophages can directly induce vascular smooth muscle cell apoptosis (Boyle et al. 2001), as can TNF, IL-1β and interferon-γ (Geng et al. 1996).

### Table 1 Actions of tumour necrosis factor

<table>
<thead>
<tr>
<th>In vitro</th>
<th>In vivo</th>
</tr>
</thead>
<tbody>
<tr>
<td>Impaired endothelial nitric oxide synthase production</td>
<td>Impaired left ventricular function</td>
</tr>
<tr>
<td>Increased inducible nitric oxide synthase production</td>
<td>Heart failure</td>
</tr>
<tr>
<td>Impaired vasodilation</td>
<td>Impaired peripheral vasodilation</td>
</tr>
<tr>
<td>Reduced myocyte contractility</td>
<td>Protein breakdown</td>
</tr>
<tr>
<td>Increased metalloproteinase activity</td>
<td>Impaired collagen synthesis</td>
</tr>
</tbody>
</table>

TIMP, tissue inhibitor of matrix metalloproteinase; IL, interleukin; MCP, monocyte chemoattractant protein.

**Testosterone and coronary artery disease**

Men are more than twice as likely as women to die from coronary heart disease, and this ratio is consistent in all populations and is not related to differences in risk factors (British Heart Foundation Statistics Database 1998). Sex hormones decline with age in both sexes but the relationship of sex hormones to cardiovascular risk is complex. Pre-menopausal women have a lower incidence of CAD, but this rises after menopause so that the risk rapidly approaches that of males. One explanation for this phenomenon is that sex hormones influence the development and progression of coronary artery disease. In males the increasing incidence of CAD with age is associated with a decline in testosterone levels. A similar, more pronounced epidemiological pattern occurs in women after the menopause, so that there is a catch-up time of around 10 years. The presumed protective effects of oestrogens in women have been widely investigated with a series of large observational studies tending to support this notion (Gordon et al. 1978, Stampfer et al. 1991). However, randomised control trials designed to assess the effect of female hormone replacement on cardiovascular events have reported either no benefit (Hulley et al. 1998) or harm (Rossouw et al. 2002). The explanation for these seemingly opposing results is unclear. The positive observational data have been attributed to ‘compliance bias’ whereby women on hormone replacement therapy (HRT) tend to be healthier, from a higher socio-economic class and exhibit fewer cardiovascular risk factors (Petitti 1998). The negative trial data of HRT in women with established vascular disease (HERS) (Hulley et al. 1998)
has been attributed to the higher use of statins in the control group (Pan & Boal 1999) and to late randomisation of subjects (with consequent failure to prove a late trend to cardio-protection) (Ong et al. 1999). The Women’s Health Initiative study (Rossouw et al. 2002) in unselected ‘healthy’ women also found no benefit and indeed small absolute increases in the risk of vascular disease and cancer. This trial has also been criticised for the high age of the study population, with 50% over 65 years old and thus high prevalence of sub-clinical vascular disease (Radford & Church 2002). Both trials have been criticised for the relatively high doses of hormones used and particularly the synthetic progestin, medoxyprogesterone, since this hormone can inhibit the beneficial effects of oestrogen (Pearson Murphy 2002). These recent randomised control trials are disappointing and surprising, but it should be noted that hormone replacement therapy does not replicate the same physiology as seen in the pre-menopausal woman. Although female HRT cannot currently be recommended to reduce CAD risk it may well be that different hormone preparations and delivery in a younger population will have some efficacy.

In comparison, the role of androgens in CAD has been relatively neglected, although a detrimental effect is usually presumed. However, we have found that low rather than high testosterone levels are associated with CAD (English et al. 2000a) and that low serum testosterone is associated with increased aortic atheroma (Hak et al. 2002). Furthermore, low testosterone levels are associated with several risk factors for the development of CAD,
including systolic and diastolic hypertension, adverse lipid profile and high levels of fibrinogen, insulin and pro-coagulable factors (reviewed in English et al. 1997).

Despite this, it has yet to be established that low testosterone level is an independent risk factor for the development of CAD in man; it may simply be an epiphenomenon. There is, however, experimental evidence that androgens have a disease-modifying role in atheroma formation. Several male animal studies have shown that androgens reduce diet- and injury-induced atherosclerosis. These studies have their limitations, as most use the cholesterol-fed rabbit with or without induced endothelial injury. This is a well-established model but has some differences compared with native human atheroma. The cholesterol diet creates a hyperlipidaemia characterised by high very low-density-lipoprotein (VLDL) rather than a high LDL more commonly seen in humans and the lipid deposition seen is in the form of cholesterol ester-enriched fatty streaks rather than mature plaque (Arad et al. 1989). However fatty streaks are known precursors of mature plaque and the morphology and behaviour of cells in the vessels of animal model fatty streaks mimics that seen in human atheroma (Gordon et al. 1988). The results are tabulated in Table 2. All but one study has shown a beneficial or neutral effect of androgen therapy on the formation and progression of atherosclerosis. The only negative study, by Toda et al. (1984), used a dose of testosterone at 150 mg per day,

Figure 2 (A) Stable plaque characterised by small necrotic core (NC), abundant vascular smooth muscle cells, and a dense thick plaque cap with extensive collagen/elastin fibres. (B) Unstable plaque characterised by a larger necrotic core, thin plaque cap with sparse smooth muscle cells and collagen fibres, inflammatory cells producing cytokines and MMPs and apoptic/senescent smooth muscle cells.
which is a very high dose and caused increases in fatty accumulation in the aorta (as well as delayed maturation of external phenotype) whereas a lower 30 mg dose showed no increase in fatty accumulation. The other studies use doses of hormones that create levels either within the physiological range or modestly supra-physiological. The study of Alexandersen et al. (1999) is the most elegant model. Here, castration of male animals without hormone replacement induced a 100% increase in aortic atheroma compared with sham-operated animals. Testosterone replacement to a high physiological range with intramuscular injections or tablets inhibited this response. The final amount of aortic atheroma was less in the intramuscular testosterone group than in the sham-operated group. The reason for this is not certain but the final fasting testosterone level in the intramuscular testosterone group than in the sham-operated group. A higher physiological range with intramuscular injections or tablets inhibited this response. The final amount of aortic atheroma was less in the intramuscular testosterone group than in the sham-operated group. The reason for this is not certain but the final fasting testosterone level in the intramuscular testosterone group could be significantly higher than in the sham-operated group, suggesting a dose–response effect. Although these animal models have clear limitations, the overwhelming impression is that the androgens dehydroepiandrosterone and testosterone in high physiological doses inhibit experimental atheroma.

### Testosterone and cytokines – a possible mechanism of protection

The means by which testosterone may confer benefit in CAD is not clear. We have previously demonstrated that 3 months of physiological testosterone treatment leads to symptomatic improvement in men with angina (English et al. 2000b). The acute and chronic anti-ischaemic properties of testosterone have been reported by other groups (Jaffe 1977, Rosano et al. 1999) and this effect is most likely mediated by coronary artery vasodilatation (Webb et al. 1999), which appears to involve calcium channel antagonism in a gender-specific fashion (English et al. 2001, 2002). However, we propose that testosterone may also have a role in limiting the vascular inflammation and cytokine activity underpinning the pathophysiology of atherosclerosis. Macrophages, lymphocytes and vascular smooth muscle cells (VSMC) all possess androgen receptors (Fujimoto et al. 1994, Benten et al. 1999a,b). In addition, there is evidence of a biofeedback loop since cytokines appear to impair synthesis and release of testosterone (Mealy et al. 1990). There have been no studies directly examining the action of androgens on cytokine production in patients with CAD. However, there is evidence from several studies that androgens possess immune-modulating properties. In the majority of these reports, androgens, including testosterone, have been shown to suppress the activity of pro-inflammatory cytokines while enhancing that of anti-inflammatory factors (see Table 3). In a recent study, hypogonadal men were found to have a greater degree of inflammatory activation compared with healthy controls, including higher serum cytokine levels (Yesilova et al. 2000). Androgen therapy in these patients led to a reduction in circulating cytokines. Clearly these studies only provide circumstantial evidence of an anti-cytokine effect of testosterone and further studies are needed. It is of interest that testosterone has been used with some success to treat males with autoimmune rheumatic conditions (Bizzarro et al. 1987, Cutolo et al. 1991). Although the studies do not report levels of serum cytokines, these diseases are essentially cytokine mediated and clinical improvement would be expected to be associated with reduced expression of pro-inflammatory cytokines.

In addition to direct anti-inflammatory actions, testosterone also appears to have an important effect on rates of vascular smooth muscle cell proliferation and apoptosis, an important factor in maintaining plaque integrity (Newby & Zaltsman 1999). The anabolic actions of testosterone in increasing protein synthesis and skeletal muscle cell size via

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**Table 2 Effects of androgen on atherosclerosis in males**

<table>
<thead>
<tr>
<th>Reference</th>
<th>Model</th>
<th>Intervention</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Toda et al. (1984)</td>
<td>7-day-old chicks</td>
<td>T 150 mg daily</td>
<td>↑ lipid accumulation in aorta</td>
</tr>
<tr>
<td>Gordon et al. (1988)</td>
<td>Cholesterol-fed rabbits, aortic balloon-injury</td>
<td>T 30 mg daily</td>
<td>&quot;&quot; no changes</td>
</tr>
<tr>
<td>Arad et al. (1989)</td>
<td>Cholesterol-fed rabbits</td>
<td>DHEA</td>
<td>↓ plaque size</td>
</tr>
<tr>
<td>Larsen et al. (1993)</td>
<td>Cholesterol-fed rabbits, castrated</td>
<td>T 25 mg twice weekly</td>
<td>↓ fatty streak formation</td>
</tr>
<tr>
<td>Eich et al. (1993)</td>
<td>Cholesterol-fed rabbits, heterotopic heart transplanted</td>
<td>DHEA</td>
<td>Trend to lower cholesterol content of aorta</td>
</tr>
<tr>
<td>Bruck et al. (1997)</td>
<td>Cholesterol-fed rabbits, castrated</td>
<td>T 25 mg/kg/week</td>
<td>↓ plaque size</td>
</tr>
<tr>
<td>Alexandersen et al. (1999)</td>
<td>Cholesterol-fed rabbits or sham-operated</td>
<td>Castration</td>
<td>↑ aortic atherosclerosis</td>
</tr>
<tr>
<td>Hanke et al. (2001)</td>
<td>Endothelium-denuded rabbit aorta</td>
<td>Incubation with T</td>
<td>↓ aortic atherosclerosis</td>
</tr>
</tbody>
</table>

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<td>Cholesterol-fed rabbits</td>
<td>DHEA</td>
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</tr>
<tr>
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<td>T 25 mg/kg/week</td>
<td>↓ plaque size</td>
</tr>
<tr>
<td>Cholesterol-fed rabbits or sham-operated</td>
<td>Castration</td>
<td>↑ aortic atherosclerosis</td>
</tr>
<tr>
<td>Endothelium-denuded rabbit aorta</td>
<td>Incubation with T</td>
<td>↓ aortic atherosclerosis</td>
</tr>
</tbody>
</table>

T, testosterone; DHEA, dehydroepiandrosterone.
nuclear transcription are well documented. Testosterone has been shown to reverse castration-induced skeletal muscle apoptosis in rats (Boissonneault 2001) and inhibit human neuronal apoptosis (Hammond et al. 2001). Importantly, it has also been shown to enhance proliferation of human vascular smooth muscle cells (Williams et al. 2002). Testosterone could, therefore, potentially be involved in maintaining the fibrous cap of the atherosclerotic plaque by promoting smooth muscle cell stability.

Conclusion

Cytokine activation and vascular smooth muscle cell apoptosis play an integral part in the development of CAD and in the pathophysiology of acute coronary syndromes. Experimental studies show that androgens protect against the development of atheroma. Separately, androgens have been shown to suppress pro-inflammatory cytokine activity, inhibit apoptosis and enhance vascular smooth muscle cell proliferation. These actions may be responsible for their athero-protective effects. To test this hypothesis, further studies are needed, including clinical trials, to examine this mechanism and the role of androgens in cardiovascular disease.

References


Table 3 Effects of androgens on cytokines

<table>
<thead>
<tr>
<th>Reference</th>
<th>Androgen</th>
<th>Model</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chao et al. (1995)</td>
<td>T</td>
<td>Rat macrophage</td>
<td>Trend to ↓ TNF production</td>
</tr>
<tr>
<td>D'Agostino et al. (1999)</td>
<td>T</td>
<td>Mouse macrophage</td>
<td>↓ LPS-induced TNF</td>
</tr>
<tr>
<td>Kanda et al. (1996)</td>
<td>T</td>
<td>Human monocytes</td>
<td>↑ LPS-induced IL-10</td>
</tr>
<tr>
<td>Kanda et al. (1997)</td>
<td>T</td>
<td>Human monocytes (patients with systemic lupus erythematosus)</td>
<td>↓ IL-6 production</td>
</tr>
<tr>
<td>Li et al. (1993)</td>
<td>T</td>
<td>Human monocytes (patients with rheumatoid arthritis and healthy subjects)</td>
<td>↓ IL-1 production</td>
</tr>
<tr>
<td>Gornstein et al. (1999)</td>
<td>T</td>
<td>Human gingival fibroblasts</td>
<td>↓ IL-6 production</td>
</tr>
<tr>
<td>Hofbauer et al. (1999)</td>
<td>T</td>
<td>Human osteoblasts</td>
<td>↓ IL-6 production</td>
</tr>
<tr>
<td>Hatakeyama et al. (2002)</td>
<td>DHT</td>
<td>Human aortic endothelium</td>
<td>↓ TNF induced VCAM-1 and NFκB</td>
</tr>
<tr>
<td>Araneo et al. (1991)</td>
<td>DHT</td>
<td>Mouse cells</td>
<td>↓ γ-interferon, IL-4</td>
</tr>
<tr>
<td>Dalal et al. (1997)</td>
<td>DHT</td>
<td>Mice with auto-immune disease*</td>
<td>↓ γ-interferon, ↑ IL-10</td>
</tr>
<tr>
<td>Kimura et al. (1998)</td>
<td>DHEA</td>
<td>Obese rats*</td>
<td>↓ TNF</td>
</tr>
<tr>
<td>Ben-Nathan et al. (1999)</td>
<td>DHEA</td>
<td>Mice*</td>
<td>↓ IL-1, ↓ LPS-induced TNF</td>
</tr>
<tr>
<td>Padgett &amp; Loria (1998)</td>
<td>DHEA</td>
<td>Mice macrophages</td>
<td>↓ LPS-induced TNF, IL-1, IL-6</td>
</tr>
<tr>
<td>Straub et al. (1998)</td>
<td>DHEA</td>
<td>Human monocytes</td>
<td>↓ IL-6 production</td>
</tr>
<tr>
<td>Spindel et al. (1992)</td>
<td>Castration</td>
<td>Mice*</td>
<td>↑ LPS-induced TNF</td>
</tr>
</tbody>
</table>

* in vivo. LPS, lipopolysaccharide; T, testosterone; DHT, dihydrotestosterone; DHEA, dehydroepiandrosterone; VCAM-1, vascular cell adhesion molecule 1; IL, interleukin; TNF, tumour necrosis factor; NFκB, nuclear factor-kappaB.


Larsen BA, Nordestgaard BG, Stender S & Kjeldsen K 1993 Effect of testosterone on atherogenesis in cholesterol-fed rabbits with similar plasma cholesterol levels. *Arteriosclerosis* 99: 79–86.


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