Characteristics of 3,5,3′-triiodothyronine (T3)-uptake system of tadpole red blood cells: effect of endocrine-disrupting chemicals on cellular T3 response

N Shimada and K Yamauchi
Department of Biology, Faculty of Science, Shizuoka University, 836 Ohya, Shizuoka 422-8529, Japan
(Requests for offprints should be addressed to K Yamauchi; Email: sbkyama@ipc.shizuoka.ac.jp)

Abstract
We characterized the 3,5,3′-triiodothyronine (T3)-uptake system on the plasma membrane of Rana catesbeiana tadpole red blood cells (RBCs) in the presence of a variety of inhibitors and potentially competing amino acids. Saturable [125I]T3 uptake was inhibited by phlorizin, monodansylcadaverine, bromosulfophthalein, sodium taurocholate and tryptophan. Saturable uptake obeyed simple Michaelis–Menten kinetics with an apparent $K_m$ of 110 nM and a $V_{max}$ of 2.5 pmol/min per 10$^6$ cells at 23 °C. These results suggested that a large proportion of T3 transported into RBCs was mediated by the aromatic amino acid transporter (System T)-linked transporter. To investigate the effect of endocrine-disrupting chemicals (EDCs) on [125I]T3 uptake, RBCs were incubated with [125I]T3 in the presence of each chemical. Among the test chemicals, di-n-butyl phthalate, n-butylbenzyl phthalate and the miticide, dicofol, were the most powerful inhibitors of [125I]T3 uptake, with an IC$_{50}$ of 2.2 µM, which was one order of magnitude greater than that for T3 (IC$_{50}$, 0.14 µM), and diethylstilbestrol and ethinylestradiol were modest inhibitors. Tributyltin accelerated saturable initial [125I]T3 uptake by 2-fold at 3.2 µM. When RBCs were cultured with 10 nM T3 at 25 °C for 2 days in the presence of monodansylcadaverine, ethinylestradiol, ioxynil or dicofol at the defined concentrations, these compounds inhibited significantly the induction of the thyroid hormone receptor α gene by T3. However, not all chemicals competed with T3 binding to the receptor at the same concentrations. Our results raise the possibility that the T3-uptake system on the plasma membrane of the tadpole RBCs could be a candidate target site for some EDCs and can modulate cellular T3 response.


Introduction
Thyroid hormones (THs), like steroids, retinoids and vitamin D, are hydrophobic signaling molecules. Within the cell, their actions are mediated by nuclear receptors, which regulate the expression of specific target genes. The nuclear receptors for these molecules belong to a superfamily of hormone nuclear receptors, which function as ligand-activated transcription factors (Yen & Chin 1994). 3,5,3′-triodothyronine (T3) is the active form of TH that binds to TH nuclear receptors (TRs). Before reaching its intracellular targets, TH must cross the plasma membrane. T3 either enters the cell directly from plasma or is converted from the precursor form of TH, t-thyroxine (T4), by deiodinases in the target cells. Because of the lipophilic nature of TH, it was thought that TH traversed the plasma membrane by simple diffusion. However, in the past decade, increasing evidence from a variety of biological sources indicates that TH is actively imported and exported at the cell surface via membrane-bound transporters. TH transporters characterized so far include amino acid transporters and organic anion transporters (Hennemann et al. 2001). The amino acid transporters involved in TH uptake consist of the transporter of large zwitterionic amino acids (System L) and the transporter linked to aromatic amino acid transporters (System T; Blondeau et al. 1988), both of which are Na$^+$-independent systems. The organic anion transporters comprise a large family of Na$^+$-independent transporters and Na$^+$-dependent taurocholate transporters (Abe et al. 2002). Very recently, it was found that fatty acid translocase, which is a family distinct from the above transporters, can transport TH into cells (van der Putten et al. 2003). Experiments involving the overexpression and inactivation of System L demonstrated an important role of this system in the regulation of gene expression mediated by TRs (Ritchie et al. 2003). Therefore, TH transport via these transporters may be one of the critical processes controlling the concentration of intracellular TH and, subsequently, the regulation of gene expression mediated by TRs.
Our recent studies have focused on the effects of endocrine-disrupting chemicals (EDCs) on the amphibian and avian thyroid systems, especially on T₃ binding to the major plasma TH-binding protein, transthyretin (TTR), and to TR. These studies revealed that most EDCs tested at micromolar concentrations did not compete with T₃ binding to TR but did compete with T₃ binding to TTR (Yamauchi et al. 2000, 2002, 2003, Ishihara et al. 2003a, 2003b). This was in contrast to the steroid system where a number of chemicals are capable of competing with estrogen and progesterone binding to their respective receptors (Tran et al. 1996, Matthews et al. 2000). To understand the action of EDCs in the thyroid system, a research focus on processes other than the competitive binding to TR is required. Recent studies on the effects of agricultural chemicals on the thyroid system support their action on the processes other than the competitive inhibition of T₃ binding to TRs. The herbicide acetochlor, for which amphibian TRs and TTRs had no significant affinity (Yamauchi et al. 2002, Ishihara et al. 2003a), altered TH-dependent gene expression and amphibian metamorphosis (Cheek et al. 1999, Velthooven & Helbing 2001, Crump et al. 2002). Xenopus tadpoles previously exposed to methoprene did not respond to exogenously added T₄ (Fort et al. 2000), although Xenopus TR and TTR showed no significant affinity for this chemical (Yamauchi et al. 2002). However, there have been few reports that focus on processes other than competitive binding of EDCs to TTR and TR in the thyroid system.

To elucidate the role of a T₃-uptake system on the cellular T₃ response and identify what kind of EDCs target the T₃-uptake system and interfere with the thyroid system, we investigated the characteristics of T₃ uptake into Rana catesbeiana tadpole red blood cells (RBCs). We chose 35 chemicals: 11 chemicals that inhibit the activity of TH transporters (Blondeau et al. 1988, McLeese & Eales 1996a) and 24 chemicals known to affect plasma TH levels in mammals (Brucker-Davis 1998), and examined their effects on [¹²⁵I]T₃ uptake into the tadpole RBCs. In this study, it was found that T₃ was transported into tadpole RBCs by the System T-linked transporter. Several compounds, including EDCs, significantly inhibited [¹²⁵I]T₃ uptake in a dose-dependent manner. Furthermore, some of these inhibitors of the T₃-uptake system suppressed the induction of the early primary T₃-response gene trα by T₃. We report here that the T₃-uptake system on the plasma membrane of tadpole RBCs can modulate T₃ signaling by controlling the amount of intracellular T₃, and that it could be a candidate target site for some EDCs that affect the thyroid system.

Materials and Methods

Reagents

[¹²⁵I]T₃ (122 MBq/µg; carrier-free) was purchased from NEN Life Science Products (Boston, MA, USA). Un-labeled T₃, bisphenol A, 1-aminopyrene, benzo[a]pyrene, pentachlorophenol, 2-amino[2,2]heptane-2-carboxylic acid (BCH), monodansylcadaverine and ouabain were obtained from Sigma (St Louis, MO, USA). Ethynyl-estradiol, dicyclohexyl phthalate, tributyltin chloride, benzophenone, bis(2-ethylhexyl) adipate, p-ter-octylphenol, mirex (dodecachloropentacyclo [5·3·0·02,6.03,9.04,8]decane, analytical standard), malathion (diethyl(dimethoxyphosphoryl)thio)succinate, analytical standard), dicofol (Keltane; 2,2,2-trichloro-1,1-bis (4-chlorophenyl)ethanol, analytical standard), t-tryptophan, t-phenylalanine, t-leucine, N-ethylmaleimide, phloretin, sodium taurocholate and choline chloride were purchased from Wako Pure Chemicals (Osaka, Japan). Ioxynil (3,5-di-iodo-4-hydroxybenzonitril, analytical standard) and acetochlor (2-chloro-N-(ethoxymethyl)-N-(2-ethyl-6-methylphenyl) acetamide) were purchased from Riedel-de Haën (Seeize, Germany). Methoprene ((E,E)-1-methylethyl-11-methoxy-3,7,11-trimethyl-2,4-dodecadienoate (9 Cl)) was from Ehrenstorfer Quality (Augsburg, Germany). 4-Nonylphenol, di-n-butyl phthalate and n-butylbenzyl phthalate were obtained from Nacalai Tesque (Kyoto, Japan). 2,4-Dichlorophenoxyacetic acid, di-2-ethylhexyl phthalate, diethyl phthalate and 2,4-dinitrophenol were from Kanto Chemicals (Tokyo, Japan), and diethyl-stilbestrol (DES) was from Tokyo Chemical Industry (Tokyo, Japan). Bromosulfophthalein was purchased from ICN Biochemicals (Irvine, CA, USA). All other chemicals used in this study were of chromatography grade or the highest grade available and purchased from Wako Pure Chemicals or Nacalai Tesque.

All chemicals tested as EDCs, except for 1-aminopyrene, benzo[a]pyrene and mirex, which were dissolved in benzene to a concentration of 10 mM, were dissolved in ethanol to concentrations of 5–10 mM. Phloretin and T₃ were dissolved in dimethylsulfoxide to concentrations of 0·6–200 mM. Monodansylcadaverine was dissolved in methanol to a concentration of 50 mM. Tryptophan, leucine, phenylalanine, bromosulfophthalein, BCH, sodium taurocholate, ouabain, N-ethylmaleimide and choline chloride were dissolved in frog Ringer solution (111 mM NaCl, 3·4 mM KCl, 2 mM CaCl₂ and 2·4 mM NaHCO₃) or 70% Leibovitz’s L-15 medium (Sigma). These chemicals were diluted with the frog Ringer solution or 70% Leibovitz’s L-15 medium to give less than 0·5% (v/v) organic solvent. Control assays were performed in the presence of the corresponding solvent only and at the same concentrations. The organic solvents added did not affect the assays for T₃ uptake, binding or cellular response.

Preparations of RBCs

R. catesbeiana tadpoles at stages X–XV (Taylor and Kallros 1946) were collected from ponds in Saitama Prefecture and Shizuoka Prefecture, Japan. The tadpoles were prepared for use in the experiments as follows: the tadpoles were killed by Ringer’s solution containing 80 µM phloretin and subjected to a cold shock. The heart and blood were slowly drained into a 2.5 mL centrifuge tube, then centrifuged at 1350 × g for 10 minutes at 4°C. The RBCs were washed by centrifugation at 1350 × g for 10 minutes at 4°C. The RBCs were then washed twice more, then suspended in the frog Ringer solution containing 80 µM phloretin (1·6–2·4 × 10⁶ cells/µL) and kept at 4°C until use. These cells were used for the experiments within 24 hours of their preparation.
Kinetic parameters were determined by fitting the plot of ARC-2000, Aloka, Japan). γ scintillation counter (Auto Well Gamma System in the extracellular fraction were determined in a V γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ γ gamma scintillation counter (Auto Well Gamma System instruction manual A-2000, Aloka, Japan).

Kinetic studies were carried out at 23 °C for 2 min. Kinetic parameters were determined by fitting the plot of initial velocity (V_i) versus T_3 concentration (S) to the Michaelis–Menten equation: V_i = V_{max} / (1 + K_{m} / S), where V_{max} is the maximum uptake rate and K_{m} is the Michaelis constant.

RNA analysis by real-time PCR

Tadpole RBCs (5·0×10^6 cells/35 mm dish) were incubated in 70% of Leibovitz’s L-15 medium for 2 days at 25 °C with solvent only or each chemical in the presence or absence of 10 nM T_3. The medium did not change during the incubation. Total RNA was extracted from tadpole RBCs using a QIAamp RNA Blood Mini kit (Qiagen, Hilden, Germany) according to the manufacturer’s directions. The RNA (5 μg per lane) was electrophoresed in 1% agarose gel containing 2·6 M formaldehyde to check the integrity of the RNA samples. Amounts of specific RNA species were estimated by real-time PCR using SYBR Green Master Mix and an ABI Prism 7000 thermocycler (Applied Biosystems, Foster City, CA, USA) after the RNA samples were treated with reverse transcriptase (TaqMan Reverse Transcript Reagents; Applied Biosystems). Each PCR was run in triplicate to control for PCR variation. The endpoint used in real-time PCR quantification, Ct, is defined as the PCR cycle number that crosses an arbitrarily placed signal threshold and is a function of the amount of target DNA present in the starting material. Quantification was determined by applying the 2^(-Ct) formula and calculating the average of the three values obtained for each sample. To standardize each experiment, the amount of bullfrog TRα (bTRα) transcript was divided by the amount of 18 S rRNA in the same samples. Primer sequences used were as follows: bTRα transcript sense 5’-GGGACTCCGAAAGGAGGAGGATGG-3’ (nucleotides 229–247) and antisense 5’-TCCCATCTCTGCTTGCTTT-3’ (294–277; Schneider & Galton 1991); and 18 S rRNA sense 5’-TGGCAATGCTTTCGCTTT-3’ (970–952), which were designed by comparing the corresponding Xenopus laevis and human sequences (Maden 1986).

[^125]I/T_3-binding assay

The ligand-binding domain of bTRα fused with glutathione S-transferase (GST-bTRα LBD) was expressed in Escherichia coli and then purified from the cell extract as described previously (Ishihara et al. 2003a). Competitive[^125]I/T_3 binding was performed with solvent only or each chemical, which was diluted with a buffer (10 mM Tris/HCl, pH 7·5, 1·5 mM EDTA, 1 mM dithiothreitol and 10% (v/v) glycerol) containing [^125]I/T_3, at defined concentrations. [^125]I/T_3 binding was initiated by mixing 50 μl GST-bTRα LBD (150 ng/tube) in the buffer with 200 μl of the above buffer containing [^125]I/T_3 with or without each chemical at 4 °C, adjusting the final concentration of [^125]I/T_3 to 0·1 nM. After 1·5 h incubation at 4 °C, protein-bound[^125]I/T_3 was separated from free[^125]I/T_3 by the Dowex method (Lennon et al. 1980, Lennon 1992). Radioactivity was measured in a gamma scintillation counter. The amount of[^125]I/T_3 bound non-specifically was derived from the radioactivity of the samples incubated with 1 μM unlabeled T_3. The non-specific binding value was subtracted from the amount of total bound T_3 to give the value of specifically bound[^125]I/T_3.

Statistical analysis

The data are presented as means ± s.e.m. Differences between groups were analyzed with either Student’s t test

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or Cochran–Cox test to evaluate the significance of the differences. \( P < 0.05 \) was considered statistically significant. The number of observations \( (n) \) is shown in parentheses.

### Results

**Characteristics of \( T_3 \)-uptake system on the plasma membrane of tadpole RBCs**

\( \left[ ^{125}I \right] T_3 \) uptake into tadpole RBCs reached equilibrium by 10 min at 23°C. As the initial uptake was approximately linear for at least the first 3-min period, the \( V_i \) of uptake was routinely measured after 2 min of incubation. Saturability of \( T_3 \) uptake was demonstrated by measuring \( \left[ ^{125}I \right] T_3 \) uptake in the presence of 5 \( \mu M \) unlabeled \( T_3 \). Approximately 85% of the \( V_i \) of \( \left[ ^{125}I \right] T_3 \) uptake was inhibited by 5 \( \mu M \) unlabeled \( T_3 \) (data not shown). These experiments suggest that a saturable uptake system is responsible for transporting most of the circulating \( T_3 \) into RBCs.

Saturable uptake conformed to Michaelis–Menten kinetics. RBCs were incubated with 0.1 nM \( \left[ ^{125}I \right] T_3 \) and various concentrations of unlabeled \( T_3 (S) \) ranging from \( 10^{-8} \) to \( 10^{-6} M \). The ratio of \( S/V_i \) was calculated and plotted as a function of \( S \) according to the Hanes representation (Fig. 1). The plots are linear and repeated experiments gave a \( K_m \) of 110 \( \pm 10 nM \) and a \( V_{max} \) of 2.5 \( \pm 0.3 \) pmol/min per 10\(^6\) cells in the frog Ringer solution \( (n=3) \) and a \( K_m \) of 170 \( \pm 20 nM \) and a \( V_{max} \) of 2.3 \( \pm 0.2 \) pmol/min per 10\(^6\) cells in 70% Leibovitz’s L-15 medium \( (n=5) \). The \( K_m \) value was 1.5 times greater when 70% Leibovitz’s L-15 medium was used instead of the frog Ringer solution. It is believed that this culture medium may have contained some competitors that interfered with the saturable \( T_3 \)-uptake system of tadpole RBCs.

To characterize the saturable \( T_3 \)-uptake system in tadpole RBCs, \( \left[ ^{125}I \right] T_3 \) uptake was investigated in the presence of chemicals known to inhibit the activity of amino acid transporters (Table 1). Of the three amino acids tested, tryptophan was the most powerful inhibitor while phenylalanine was a weak inhibitor and leucine had no effect, at 1 mM. The effect of BCH, a prototypical substrate of the amino acid transporter System L, was weak even at 5 mM. Organic anions, bromosulfophthalein and taurocolate were strong inhibitors. Bromosulfophthalein, at the concentration of 0.3 mM, inhibited \( \left[ ^{125}I \right] T_3 \) uptake by more than 90%. Choline chloride (111 mM), substituted

### Table 1

Effect of compounds known to inhibit the activity of amino acid transporters on \( \left[ ^{125}I \right] T_3 \) uptake into tadpole RBCs. Each value shown is the mean ± S.E.M.

<table>
<thead>
<tr>
<th>Inhibitor</th>
<th>Concentration (mM)</th>
<th>( \left[ ^{125}I \right] T_3 ) uptake (% of control)</th>
<th>( n )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (frog Ringer solution)</td>
<td>100 ± 3</td>
<td>100 ± 6</td>
<td>4</td>
</tr>
<tr>
<td>Tryptophan</td>
<td>1</td>
<td>24 ± 3*</td>
<td>7</td>
</tr>
<tr>
<td>Leucine</td>
<td>1</td>
<td>94 ± 3</td>
<td>7</td>
</tr>
<tr>
<td>Phenylalanine</td>
<td>1</td>
<td>76 ± 3*</td>
<td>7</td>
</tr>
<tr>
<td>BCH</td>
<td>5</td>
<td>97 ± 4*</td>
<td>4</td>
</tr>
<tr>
<td>Bromosulfophthalein</td>
<td>0.3</td>
<td>45 ± 4*</td>
<td>4</td>
</tr>
<tr>
<td>Sodium taurocholate</td>
<td>0.5</td>
<td>79 ± 5*</td>
<td>4</td>
</tr>
<tr>
<td>Choline chloride</td>
<td>111</td>
<td>91 ± 2</td>
<td>4</td>
</tr>
<tr>
<td>N-Ethylmaleimide</td>
<td>1</td>
<td>108 ± 1</td>
<td>3</td>
</tr>
<tr>
<td>Ouabain</td>
<td>1</td>
<td>79 ± 5*</td>
<td>4</td>
</tr>
</tbody>
</table>

\* \( P < 0.001 \), significant difference from the control values.
for sodium chloride in the frog Ringer solution, showed no effect. N-Ethylmaleimide (1 mM), a thiol reagent, inhibited only 20% of the saturable T₃ uptake. Ouabain (1 mM) did not inhibit, but phloretin (0·1 mM) and monodansylcadaverine (0·5 mM) inhibited the saturable T₃ uptake by 80%.

Figure 2 shows dose-dependent curves of eight selected compounds, including unlabeled T₃, on the saturable initial uptake of [¹²⁵I]T₃ into tadpole RBCs in the frog Ringer solution. The most effective compound was unlabeled T₃. The rank order affinity was T₃ (the concentration of T₃ required for 50% inhibition of saturable [¹²⁵I]T₃ (IC₅₀)=0·14 µM)>phloretin (52 µM)>monodansylcadaverine (59 µM)>bromosulfophthalein (75 µM)>taurocholate (270 µM)>tryptophan (470 µM)>>leucine=BCH (Table 2). Leucine and BCH had no effect at all concentrations tested.

Effect of EDCs on T₃ uptake into tadpole RBCs
Saturable initial uptake of [¹²⁵I]T₃ into the tadpole RBCs was examined in the presence of various chemicals (8 µM) at 23 °C (Fig. 3). As unlabeled T₃ inhibited completely the saturable initial uptake of [¹²⁵I]T₃ at 5–8 µM, a concentration of 8 µM for the chemicals was chosen to determine their relative potencies. T₄ inhibited the saturable initial uptake of [¹²⁵I]T₃ by 60%, suggesting that this uptake system can recognize both T₃ and T₄ but is more specific for T₃ than for T₄. Of the five xenoestrogens, ethinylestradiol, DES, p-t-octylphenol, 4-nonylphenol and bisphenol A, ethinylestradiol and DES were the most potent inhibitors: approximately 50% of the saturable initial uptake of [¹²⁵I]T₃ was inhibited. Of the two pyrene compounds, benzo[a]pyrene activated the uptake up to 150%. Among the five phthalates, di-n-butyl phthalate and n-butylbenzyl phthalate were the most potent.

Table 2 Inhibition of [¹²⁵I]T₃ uptake into tadpole RBCs by amino acid transport inhibitors.

<table>
<thead>
<tr>
<th>Chemical</th>
<th>IC₅₀ (µM)</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>Unlabeled T₃ (in frog Ringer solution)</td>
<td>0·14±0·01</td>
<td>3</td>
</tr>
<tr>
<td>Phloretin</td>
<td>52±3</td>
<td>3</td>
</tr>
<tr>
<td>Monodansylcadaverine</td>
<td>59±2</td>
<td>3</td>
</tr>
<tr>
<td>BSP</td>
<td>75±16</td>
<td>3</td>
</tr>
<tr>
<td>Sodium taurocholate</td>
<td>270±30</td>
<td>3</td>
</tr>
<tr>
<td>Tryptophan</td>
<td>470±40</td>
<td>5</td>
</tr>
<tr>
<td>Leucine</td>
<td>&gt;&gt;1000</td>
<td>3</td>
</tr>
<tr>
<td>BCH</td>
<td>&gt;&gt;1000</td>
<td>3</td>
</tr>
<tr>
<td>Unlabeled T₃ (in Leibovitz’s L-15 medium)</td>
<td>0·24±0·02</td>
<td>4</td>
</tr>
<tr>
<td>Di-n-butyl phthalate</td>
<td>2·2±0·2</td>
<td>3</td>
</tr>
<tr>
<td>n-Butylbenzyl phthalate</td>
<td>2·2±0·1</td>
<td>3</td>
</tr>
<tr>
<td>Dicofol</td>
<td>2·2±0·2</td>
<td>5</td>
</tr>
<tr>
<td>Tributyltin chloride</td>
<td>3·2±0·5*</td>
<td>3</td>
</tr>
</tbody>
</table>

*This value is the ED₂⁰₀, because only tributyltin chloride accelerated saturable initial uptake of [¹²⁵I]T₃.
Inhibitors: approximately 70–80% of the saturable initial uptake of $[125\text{I}]T_3$ was inhibited. Of the other chemicals, including agricultural chemicals, dicofol was the most powerful inhibitor: more than 80% of the saturable initial uptake of $[125\text{I}]T_3$ was inhibited. Mirex and tributyltin accelerated the saturable initial uptake of $[125\text{I}]T_3$ up to 150 and 290%, respectively.

Figure 4 shows dose-dependent curves of the five selected chemicals, including unlabeled $T_3$, on the saturable initial uptake of $[125\text{I}]T_3$ into tadpole RBCs in 70% Leibovitz’s L-15 medium. The IC$_{50}$ values of di-$n$-butyl phthalate, $n$-butylbenzyl phthalate and dicofol were all 2.2 µM (Table 2). The concentration of tributyltin required to accelerate the saturable initial uptake of $[125\text{I}]T_3$ up to 200% (ED$_{200}$) was 3.2 µM (Table 2). These effective concentrations were one order of magnitude greater than the IC$_{50}$ of $T_3$ (0.24 µM), but may be less than the IC$_{50}$ of $T_4$ (approximately 8 µM; see Fig. 3) although we did not determine the accurate IC$_{50}$ value of $T_4$.

Effect of the inhibitors of the saturable $T_3$-uptake system on the induction of TR$\alpha$ transcript by $T_3$ in tadpole RBCs

To evaluate the biological effect of the inhibitors of the saturable $T_3$-uptake system on $T_3$ signaling in tadpole...
RBCs, we examined their interference with the induction of the early primary T₃-response gene, trα, by 10 nM T₃. The addition of T₃ increased the amount of the TRα transcript by 6–7 fold. Further addition of monodansylcadaverine (final concentration of 60 µM), phloretin (50 µM), bromosulfophthalein (1 µM), ethinylestradiol (8 µM) and dicofol (0·4 µM), depressed significantly the T₃ response (Fig. 5). Di-n-butyl phthalate, but not n-butylbenzyl phthalate, had a significant synergic effect on the induction of the TRα transcript by T₃ at 8 µM, the concentration at which the saturable initial uptake of [¹²⁵I]T₃ was strongly inhibited by these phthalates (Fig. 3). This effect was abolished in the absence of T₃, Bisphenol A (8 µM) and ioxynil (0·2 µM) inhibited the induction of the TRα transcript by T₃ although they did not inhibit the saturable initial uptake of [¹²⁵I]T₃ at the same or higher concentrations (Fig. 3). We could not evaluate the effect of tributyltin on the induction of the TRα transcript by T₃ because of its severe cytotoxicity at 10⁻⁷-10⁻⁵ M.

To examine whether the chemicals that affected the induction of the TRα transcript by T₃ interact directly with TRα to compete with T₃ binding, we next examined their effect (at the same concentrations) on [¹²⁵I]T₃ binding to purified GST-bTRα LBD (Fig. 6). Phloretin, bromosulfophthalein and DES inhibited [¹²⁵I]T₃ binding to GST-bTRα LBD; however, monodansylcadaverine, ethinylestradiol, bisphenol A, dibutyl phthalate, ioxynil and dicofol showed no effect on [¹²⁵I]T₃ binding to the GST-bTRα LBD at concentrations that affected the induction of the TRα transcript by T₃. The T₃-binding activity of GST was negligible when purified GST, which was expressed from pGEX-6P-3 in E. coli, was substituted for the GST-bTRα LBD fusion protein in the assay.

Discussion

Here we have described the inhibitory mode of the T₃-uptake system of bullfrog tadpole RBCs: sensitivity to monodansylcadaverine, phloretin and tryptophan but insensitivity to leucine, BCH and ouabain (Table 1). These were consistent with the inhibitory mode of the
T₃-uptake system of rat and rainbow trout RBCs (Zhou et al. 1990, McLeese & Eales 1996a) and rat liver (Blondeau et al. 1988), some of which were characterized as System T-linked T₃-uptake system (Osty et al. 1988, Sanson et al. 1992). The fact that the T₃-uptake system in tadpole RBCs was sensitive to taurocholate and bromosulfophthalein but was insensitive to choline suggests the possibility that some of Na⁺-independent organic anion transporters (Abe et al. 1998, 1999, Fujiwara et al. 2001) are involved in the tadpole T₃-uptake system. The Kₘ values obtained from our studies were within the same range of those values obtained for the System T-linked T₃-uptake system (Blondeau et al. 1988, Osty et al. 1988, Zhou et al. 1990, McLeese & Eales 1996b), but one order of magnitude lower than those values obtained for System L (Blondeau et al. 1993, Ritchie et al. 1999, Ritchie & Taylor 2001, Friesema et al. 2001) and the organic anion transporters (rat oatp2 and oatp3, human LST-1 and OATP-E; Abe et al. 1998, 1999, Fujiwara et al. 2001). Therefore, the T₃-uptake system that is responsible for a major part of saturable T₃ uptake into tadpole RBCs may be the System T-linked uptake system in tadpole RBCs. This is the first report demonstrating the existence of the System T-linked T₃-uptake system in frogs. The expression of this T₃-uptake system in RBCs has probably been conserved during vertebrate evolution. Tadpole RBCs (larval type) are specifically removed from the bloodstream during metamorphic climax stages by apoptotic cell death under the influence of THs (Hasebe et al. 1998). Therefore, the system T-linked T₃-uptake system on the plasma membrane would play an important role in T₃-induced apoptosis in tadpole RBCs. The Kₘ value determined in 70% Leibovitz’s L-15 medium was 1.6 times greater than that determined in the frog Ringer solution. As Leibovitz’s L-15 medium contains 1.5 mM phenylalanine and 0.1 mM tryptophan, it is likely that these amino acids compete with saturable T₃ uptake into tadpole RBCs.

The effect of N-ethylmaleimide on System T-linked T₃-uptake systems was contradictory. N-ethylmaleimide, at a concentration of 1 mM, hardly inhibited saturable [¹²⁵I]T₃ uptake into tadpole RBCs (Table 1). Our result was consistent with the report for rat hepatocytes (Blondeau et al. 1988). Like us, Blondeau et al. examined [¹²⁵I]T₃ uptake in the presence of N-ethylmaleimide without preincubation with it. The saturable [¹²⁵I]T₃ uptake into rat and rainbow trout RBCs was strongly inhibited by N-ethylmaleimide when the cells were preincubated with it at 0.5–1.0 mM for 10–15 min (Zhou et al. 1990, McLeese & Eales 1996a). However, Kemp & Taylor (1997) found that approximately 40% of the saturable [¹²⁵I]T₃ uptake into sinusoidal membrane vesicles obtained from rat liver homogenate, corresponding to the System T-linked uptake system, was...
Table 3 Summary of the possible target sites of chemicals affected on the thyroid system and their effective concentrations. Data were collected from Figs 2–6 and summarized

<table>
<thead>
<tr>
<th>Chemical</th>
<th>Concentration (µM)</th>
<th>Uptake system</th>
<th>T3 binding to TRα</th>
<th>Induction of TRα transcript by T3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phloretin</td>
<td>50</td>
<td>↓</td>
<td>↓</td>
<td>↓</td>
</tr>
<tr>
<td>DES</td>
<td>8</td>
<td>↓</td>
<td>↓</td>
<td>↓</td>
</tr>
<tr>
<td>Monodansylcadaverine</td>
<td>60</td>
<td>↓</td>
<td>—</td>
<td>↓</td>
</tr>
<tr>
<td>Ethinylestradiol</td>
<td>8</td>
<td>↓</td>
<td>—</td>
<td>↓</td>
</tr>
<tr>
<td>Bromosulfophthalein</td>
<td>1</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Ioxynil</td>
<td>0·2</td>
<td>—**</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Dicofol</td>
<td>0·4</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Bisphenol A</td>
<td>8</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Di-n-butyl phthalate</td>
<td>8</td>
<td>↓</td>
<td>—</td>
<td>↑*</td>
</tr>
<tr>
<td>n-Butylbenzyl phthalate</td>
<td>8</td>
<td>↓</td>
<td>—</td>
<td>—***</td>
</tr>
</tbody>
</table>

—, No effect; ↓, inhibition; ↑, activation (*synergic effect with T3 but no effect without T3); ***, no effect at even 8 μM; ***, no significant effect although this mean value was higher than the mean value of the control.

To facilitate our understanding of the effect of the chemicals used in this study on TH signaling in tadpole RBCs, their possible target sites are summarized in Table 3, where the chemicals are divided into five groups according to their mode of action. The first group includes the chemicals that inhibited both the T3-uptake system and T3 binding to TR: phloretin and DES. The second group consists of the chemicals that inhibited the T3-uptake system but not T3 binding to TR: monodansylcadaverine and ethinylestradiol. Conversely, the chemical belonging to the third group, bromosulfophthalein, did not inhibit the T3-uptake system but did inhibit T3 binding to TR. The fourth group includes the chemicals that inhibited neither the T3-uptake system nor T3 binding to TR: ioxynil, dicofol and bisphenol A. All chemicals belonging to the first four groups suppressed the induction of the TRα transcript by T3 at the indicated concentrations. Therefore, the chemicals in the fourth group would target some process other than the two processes we investigated here. The fifth group contains those chemicals that inhibited T3 uptake but not T3 binding to TR and that did not suppress the induction of TRα transcript by T3. Di-n-butyl phthalate, a chemical belonging to the fifth group, activated synergically the induction of the TRα transcript with T3 by an unknown mechanism. These results indicate that thyroid-disrupting chemicals target several processes within the thyroid system in a chemical-dependent manner, although at present we cannot elucidate which process when inhibited by the chemicals exerts the most profound effect on the thyroid system. The present study raises the possibility that the T3-uptake system could be a candidate target for these chemicals belonging to the first and second groups.
From the present study, we cannot evaluate the metabolic effect of these chemicals that might occur in organisms. Further studies are necessary for understanding the effects of the chemicals on cells or organisms for an extended period of time. We examined the $V_{r}$ of saturable $[^{125}I]T_{3}$ uptake for the first 2-min period and the induction of the $\alpha$ gene in 2-day cultured RBCs, in the presence of EDCs. The difference in the cellular responses to EDCs between the 2-min and 2-day treatments needs to be considered.

Acknowledgments

We thank Ms Yumiko Kudo for the preparation of recombinant GST-TRα LBD. We are also grateful to Ms Julie Monk for a thorough and critical reading of the manuscript. This work was supported by a Grant-in-Aid for Scientific Research on Priority Area (2) (no. 14042223) from the Ministry of Education, Culture, Sports, Science and Technology of Japan and a Grant-in-Aid for Scientific Research (B) (nos 13559001 and 16310038) from the Japan Society for the Promotion of Science. The authors declare that there is no conflict of interest that would prejudice the impartiality of this scientific work.

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