Characterisation of the prereceptor regulation of glucocorticoids in the anterior segment of the rabbit eye

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Abstract

The prereceptor regulation of glucocorticoids (GCs) by 11β-hydroxysteroid dehydrogenase type-1 (11β-HSD1), a bidirectional isozyme that interconverts active (cortisol) and inactive (cortisone) GCs, is an established determinant of GC function in tissues such as liver, adipose and bone. Although the therapeutic use of GCs is abundant in ophthalmic practice, where GC interactions with nuclear receptors modulate gene transcription, the prereceptor regulation of endogenous cortisol is not well described in ocular tissues. Recent descriptive studies have localised 11β-HSD1 to the human corneal epithelium and non-pigmented epithelium (NPE) of the ciliary body, indicating a link to corneal epithelial physiology and aqueous humour production. In this study, we characterise the functional aspects of the autocrine regulation of GCs in the anterior segment of the rabbit eye. Using our in-house generated primary antibody to human 11β-HSD1, immunohistochemical analyses were performed on paraffin-embedded sections of whole New Zealand white albino rabbits, (NZWAR) eyes. As in human studies, 11β-HSD1 was localised to the corneal epithelium and the NPE. No staining was seen in the albino ‘pigmented’ ciliary epithelium. Specific enzyme assays for oxo-reductase (cortisone → cortisol) and dehydrogenase (cortisol → cortisone) activity indicated predominant 11β-HSD1 oxo-reductase activity from both the intact ciliary body tissue (n = 12, median 2.1 pmol/mg per h and range 1.25–2.8 pmol/mg per h; P = 0.006) and primary cultures of corneal epithelial cells (n = 12, median 3.0 pmol/mg per h and range 1.0–7.4 pmol/mg per h, P = 0.008) compared with dehydrogenase activity (median 1.0 pmol/mg per h and range 0.5–2.0 pmol/mg per h; median 0.5 pmol/mg per h and range 0.25–1.9 pmol/mg per h respectively). These findings were supported by expression of 11β-HSD1 protein as visualised by Western blotting of ciliary body tissue and immunocytochemistry of corneal epithelial cells. Reduction of corneal epithelial cell proliferation was seen after primary cultures were co-incubated with cortisol and cortisone. 11β-HSD1 activity was not demonstrated in naïve conjunctival fibroblasts or corneal stromal keratocytes. Our results indicate that the distribution of 11β-HSD1 in the rabbit resembles that of the human eye and activates cortisone to cortisol in both corneal and uveal tissues. The NZWAR provides a suitable in vivo model for the further evaluation of 11β-HSD1 activity in the eye, especially its role in corneal epithelial and ciliary body physiology.

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Introduction

The cellular actions of glucocorticoids (GCs) are largely mediated through interactions with the glucocorticoid receptor (GR), a nuclear receptor that is found in its inactive form within the cytoplasm. Ligand–receptor binding induces conformational changes within the GR unmasking nuclear localisation signals. Translocation of the ligand-activated GR to hormone response elements in the nuclear chromatin modulates transcription of specific GC-responsive genes (Tsi & O’Malley 1994, Akner et al. 1995, Czar et al. 1996). The prereceptor regulation of GCs has emerged as a critical determinant of GC function in tissues such as the liver, adipose and bone. Regulation is dependent principally on the expression of a bidirectional isozyme, 11β-hydroxysteroid dehydrogenase (11β-HSD1), that interconverts active cortisol and inactive cortisone (Stewart & Krozowski 1999, Tomlinson et al. 2004). In vivo oxo-reductase activity (cortisone → cortisol) predominates through the provision of cofactor (NADPH) by hexose-6-phosphate dehydrogenase, thereby mediating classical GC responses (Draper et al. 2003, Bujalska et al. 2005). Interest in the isozyme has escalated primarily because of its putative role in diseases such as human obesity, insulin resistance and osteoporosis (Kotelevtsev et al. 1997, 1998).

Corticosteroids (cortisol and aldosterone) are critical determinants of normal ocular function—mediating processes such as ion transport, glucoconogenesis and immune regulation that are vital for sight. Not only MR and GR, but also 11β-HSD1, are expressed throughout the human eye (MR: corneal epithelium, endothelium, iris, non-pigmented epithelium (NPE), pigmented epithelium (PE), ciliary body and retinal pigmented epithelium (RPE); and GR: trabecular meshwork (TM), corneal epithelium, corneal endothelium, ciliary body stroma and retina) (Lin et al. 1984, Weinreb et al. 1985, Mirshahi et al. 1996, 1997). Recent descriptive studies have localised this prereceptor regulator of GC function in the human corneal epithelium and ciliary body epithelium (Stokes et al. 2000, Rauz et al. 2001, Suzuki et al. 2001).

The cornea is the transparent window of the eye formed both protective and refractive properties essential for sight. The corneal epithelium is the most superficial layer formed from highly specialised non-keratinised, stratified squamous cells that rapidly proliferate from a peripheral (limbal) stem cell population, replenishing the ocular surface (Dua & Azuara-Blanco 2000, Daniels et al. 2001, Kinoshita et al. 2001). Recent data have demonstrated the expression of both 11β-HSD1 and serum and GC-regulated kinase isoform 1 (SGK1, a downstream marker of GC function) to the basal cells of the corneal epithelium (Rauz et al. 2001, 2003a). SGK1 is also localised to the limbal epithelium and ciliary body epithelium (Rauz et al. 2003a). As GCs are known to delay wound healing and SGK1 is closely linked to the cell cycle (Buse et al. 1999, Hayashi et al. 2001), 11β-HSD1 may have a role in corneal epithelial cell renewal and consequently in the management of patients with ocular surface disease.

One of the principal sodium transporting tissues in the human eye is the ocular ciliary epithelium. This is a complex bilayer of pigmented and non-pigmented-polarised, neuro-epithelial cells producing aqueous humour (AH) fundamental to the maintenance of intraocular pressure (IOP) and nutrition to the avascular and transparent structures of the eye, such as the cornea, lens and trabecular meshwork. The expression of 11β-HSD1 to the NPE of the human ciliary body (Stokes et al. 2000, Rauz et al. 2001, 2003b) is an unexpected finding as 11β-HSD2 is the principal isozyme involved in regulating sodium and water homeostasis in classical sodium transporting epithelia such as the kidney. Nevertheless, studies evaluating the post-receptor pathways in the NPE have identified corticosteroid-mediated expression of SGK1 and epithelial sodium channel subunits (Rauz et al. 2003a). Furthermore, high ratios of AH cortisol:cortisone reflect a functional intraocular 11β-HSD1 (cortisone→cortisol) (Rauz et al. 2001, 2003b) and a significant reduction (15–20%) in IOP following systemic inhibition of 11β-HSD1 with the non-selective inhibitor, carbenoxolone, has been established in both ‘normal’ volunteers (Rauz et al. 2001) and patients with low to moderate risk ocular hypertension (patients with raised IOP without optic neuropathy) (Rauz et al. 2003b). In addition, patients with primary open-angle glaucoma (a prevalent, sight-threatening disease associated with uncontrolled IOP and optic nerve damage) exhibit increased peripheral vascular sensitivity to GCs and the ratio of cortisol to cortisone metabolites has been found to be elevated in these patients versus normal controls (Stokes et al. 2003). These data support the role of 11β-HSDs in IOP homeostasis and the pathogenesis of primary open-angle glaucoma, thereby providing a possible therapeutic target for the treatment of patients with uncontrolled IOP.

To explore these concepts further, animal models are essential for both endorsing the mechanism underlying our hypotheses linking 11β-HSD1 to corneal epithelial physiology and AH production, and for evaluating novel topical selective 11β-HSD1 inhibitors. Although 11β-HSD oxoreductase and dehydrogenase activities have been demonstrated in homogenised rat eyes (Stokes et al. 2000), no studies have described tissue- or cell-specific 11β-HSD activity in the mammalian eye. A possible animal model for these experiments appears to be the New Zealand white albino rabbit (NZWAR). This is an established model for the investigation of corneal disease (Nakamura et al. 2003, Tungsiripat et al. 2003, Iversen et al. 2004, Mimura et al. 2005) and AH dynamics (Tsukahara et al. 1986, Green et al. 1989, Munden & Schmidt 1992, Lim et al. 2005). The corticosteroid hormone profile is similar to humans (i.e. cortisol rather than corticosterone in rodents). Our study was designed to evaluate the expression of 11β-HSD1 in the NZWAR ocular tissues and to confirm its suitability as an in vivo model for evaluating the effects of local 11β-HSD1 expression, primarily in the corneal epithelium and NPE.

Materials and Methods

Animals and tissues

Ocular tissues were harvested from 16 NZWAR in a manner consistent with the FRAME Guidelines on research involving the use of laboratory animals, the UFAW Handbook on the care and management of laboratory animals and the ARVO Statement for the use of animals in ophthalmic and vision research. Conjunctiva were surgically dissected and fixed in formalin or stored in a sterile container for primary culture. One globe of each rabbit was fixed in formalin. The remaining globes were bisected in the coronal plane separating the anterior and posterior segments of the eye. The ciliary body
and corneal scleral rim were carefully dissected from the anterior segment with the aid of a dissecting binocular microscope.

**Primary cultures**

**Conjunctival fibroblasts** Conjunctival specimens were irrigated in sterile PBS in a class II laminar flow cabinet. Each explant was maintained in a single well of a six-well culture plate, in Dulbecco’s modified Eagle’s medium (DMEM) with glutamax, 1000 mg/ml glucose and sodium pyruvate (Invitrogen) supplemented with 10% (v/v) foetal calf serum (FCS) incubated in a humidified atmosphere set at 37°C with 5% CO₂ until outgrowth of fibroblasts was observed. Confluent fibroblast cultures were established on day 14 and were subsequently subcultured using trypsin in the conventional manner. All the experiments were carried out on passages 1 and 2.

**Corneal epithelial cells** Primary rabbit corneal epithelial cell cultures were derived by incubating corneal–scleral discs for 2 h at 37°C with 1·2 IU/ml neutral protease (Dispase II; Roche). The epithelium was stripped off with gentle scraping from the limbus to the centre into PBS, which was centrifugated at 500 g for 5 min. The cells were resuspended in keratinocyte medium (Gibco) supplemented with penicillin 100 IU/ml, streptomycin 100 mg/ml, epidermal growth factor 5 ng/ml, bovine pituitary extract 2·5 mg/ml (Gibco) and FCS 5%. Cells were cultured in 24-well plates at 37°C with 5% CO₂ in 95% humidified air until 80% confluent on 7–14 days. No experiments were performed on subcultured cells.

**Keratocytes** Primary rabbit keratocytes cultures were established by incubating central corneal buttons (trephined from the remnant corneal–scleral disc used for deriving primary corneal epithelial cells) in serum-free conditions consisting of DMEM with glutamax, 1000 mg/ml glucose and sodium pyruvate (Invitrogen) in a humidified atmosphere set at 37°C with 5% CO₂. Confluent keratocyte cultures were established on day 14 and were subsequently subcultured using conventional techniques. All the experiments were carried out on passages 1 and 2.

**Immunohistochemistry**

Immunohistochemical analyses were performed on 5 μm formalin-fixed, paraffin-embedded sections of 16 NZWAR eyes. Immunoperoxidase studies were performed using antisera raised in sheep against human 11β-HSD1 (amino acids 18–33) and 11β-HSD2 (amino acids 137–160 and 334–358) as previously reported (Shimojo et al. 1997, Ricketts et al. 1998). Antibody dilutions were 1:100 for 11β-HSD1 and 1:200 for 11β-HSD2. The control sections included the omission of primary antibody and use of antibody pretreated with the immunising peptides. Secondary antibodies comprised donkey anti-sheep peroxidase conjugate (1:100; Binding Site). The sections were visualised with the peroxidase substrate 3,3′-diaminobenzidine. NZWAR liver and kidney sections were used as positive control tissues for 11β-HSD1 and -2 respectively.

**11β-HSD1 enzyme assays**

Whole tissue ciliary body explants from each eye were divided into quarters and three pieces were placed in three wells of a 24-well plate. The fourth piece of tissue was formalin-fixed and paraffin embedded for confirmatory histology with haematoxylin and eosin. Primary cultures of human conjunctival fibroblasts, keratocytes and corneal epithelial cells were initiated as described earlier, grown to partial confluence in 12-well (conjunctival fibroblasts) and 24-well (keratocytes, corneal epithelial cells) plates. The medium was replaced with serum-free medium 2 h before incubation with various concentrations of corticosteroids. Dehydrogenase activity (cortisol → cortisone conversion) was assessed using 100 nM (3·625 × 10⁻⁵ mg/ml) unlabelled cortisol (Sigma) diluted in serum-free medium and trace amounts (1·5 nM) of [³H] cortisol (specific activity 78·4 Ci/mmole, NEN, Boston, MA, USA) at 37°C for 24 h. Conversion of cortisone to cortisol (oxo-reductase) was analysed by incubating cells with 100 nM (3·604 × 10⁻⁵ mg/ml) unlabelled cortisone and trace amounts of [³H]cortisone (50 000 c.p.m.) synthesised in-house as described previously (Bujalska et al. 1997). Whole tissue and primary cultures were incubated with GC.
substrates as described earlier and a 100-fold excess (4·707×10⁻³ mg/ml) of glycyrhretinic acid (GE; Sigma), an inhibitor of 11β-HSDs (Monder et al. 1989). After 24 h incubation, steroids were extracted from the medium with 10 volumes of dichloromethane separated by thin-layer chromatography with chloroform:ethanol (92:8) as a mobile phase and the fractional conversion of cortisol to cortisone or cortisone to cortisol was calculated after scanning analysis using a Bioscan 2000 radioimaging detector (Bioscan, Washington, DC, USA). Following removal of culture medium for enzyme assay, cell monolayers were lysed in 1 ml water for subsequent protein assays. Total protein in each well was determined using a standard protein assay reagent (Bio-Rad) and enzyme activities were expressed as pmol/h per mg protein. All assays were carried out in triplicate and data are presented as medians with full ranges analysed by non-parametric statistical methods (Mann–Whitney U and Wilcoxon) using the software packages SSPS for Windows Version 11.5.1 (SPSS, Inc., Chicago, IL, USA, 2002) and Prism for Windows Version 4.03c (GraphPad Software, Inc., San Diego, CA, USA).

Corneal epithelial cell proliferation assays

Corneal epithelial proliferation was assessed using a commercially available colorimetric assay for determining the number of viable cells (CellTiter 96 AQueousOne; Promega) according to the manufacturer's guidelines with appropriate controls (no cells). Cells were cocultured with 100 nM cortisone, cortisone and vehicle plus assay reagents for 1 h at 37°C (cells). Cells were coincubated with 100 nM cortisone, to the manufacturer's guidelines with appropriate controls (no

Results

Primary cultures

NZWAR conjunctival fibroblasts were established in culture for 14 days and demonstrated characteristic, elongated morphology (Fig. 1A). A confluent monolayer of corneal epithelial cells with distinctive hexahedral architecture was seen between 7 and 14 days of culture (Fig. 1B). Corneal epithelial cells phenotype was confirmed by cytokeratin-3 immunofluorescence staining (Fig. 1C). These cells also expressed 11β-HSD1 (Fig. 1D) that was absent when 11β-HSD1 antibody was preincubated with immunising peptide (Fig. 1E). Following 14 days in culture, outgrowth of stellate-shaped keratocytes could be observed from the central corneal explants (Fig. 1F).

Immunohistochemistry

Sections of rabbit liver tissue probed with 11β-HSD1 antibody revealed intense immunoreactivity within the cells surrounding the central vein (Fig. 2A) consistent with the pattern of staining observed in human liver (Ricketts et al. 1998). This confirmed good cross-species reactivity of the human 11β-HSD1 antibody with rabbit tissues, as seen for the immuno-cytchemistry analyses of the primary corneal epithelial cells (Fig. 1D). Minimal staining was detected in the liver using the preadsorbed antibody (Fig. 2B). 11β-HSD1 staining was seen in both corneal and conjunctival epithelia and also corneal endothelium (Fig. 2C–E respectively). In the NZWAR ciliary body, 11β-HSD1 immunoreactivity was restricted to the NPE layer (Fig. 2F) and could be specifically removed with immunising peptide (Fig. 2G). No staining was observed in the ciliary body using anti-human 11β-HSD2 antibody (Fig. 2H) despite good cross-reactivity with rabbit tissues confirmed by intense staining of rabbit kidney cortical collecting ducts (Fig. 2I). The conjunctival and corneal tissues did not express 11β-HSD2 (data not shown).

11β-HSD assays

Thin-layer radio–chromatograms (Fig. 3A–C) confirmed predominant oxo-reductase activity in dissected ciliary body tissues (NPE, PE and ciliary body stroma) indicated by a significantly higher substrate conversion of cortisone...
to cortisol (median 2·1 pmol/mg per h and range 1·25–2·8 pmol/mg per h) versus cortisol to cortisone conversion (median 0·87 pmol/mg per h and range 0·5–2 pmol/mg per h), *P*=0·006 (Fig. 3D). Coincubation with GE, a 11β-HSD1 inhibitor, confirmed a significant reduction of 11β-HSD1 oxo-reductase activity (median 0·4 pmol/mg per h and range 0·2–1·2 pmol/mg per h, *P*=0·025).

**Primary cultures** Following 24-h incubation, no 11β-HSD1 activity could be detected from either conjunctival fibroblast or keratocytes primary cultures. Primary corneal epithelial cells demonstrated oxo-reductase activity (median 3·0 pmol/mg per h and range 1·0–7·4 pmol/mg per h) that was significantly higher than dehydrogenase activity (median 0·5 pmol/mg per h and range 0·4–2·9 pmol/mg per h) after similar 24-h incubation, *P*=0·008 (Fig. 4).

**Corneal epithelial cell proliferation assays**

Isolated rabbit corneal epithelial cells proliferated rapidly (tenfold increase from baseline) in untreated culture medium...
Figure 2  (A) Immunohistochemistry using human anti-11β-HSD1 confirms characteristic distribution of 11β-HSD1 in the rabbit liver surrounding the central vein. (B) No staining was detected using the preadsorbed antibody. 11β-HSD1 staining was seen in both (C) corneal and (D) conjunctival epithelia and also (E) corneal endothelium. (F) In the ocular ciliary body, 11β-HSD1 immunoreactivity was restricted to the non-pigmented epithelium (NPE) with the absence of staining in the albino rabbit ‘pigmented’ ciliary epithelium. (G) NPE staining was specifically reduced with preadsorbed immunising peptide. (H) There was no expression of 11β-HSD2 in the ciliary body despite good cross-reactivity with rabbit tissues, confirmed by (I) intense staining of New Zealand white albino rabbit (NZWAR) kidney cortical collecting ducts (n=4 rabbits). CV, central vein; NPE, non-pigmented epithelial cells; PE, pigmented epithelial cells; KT, kidney tubule.
However, there was a 50% reduction of cell divisions observed over a 96-h period of culture when cells were co-incubated with either cortisone (the inactive GC substrate for 11β-HSD1) or cortisol (active GC; $P=0.008$).

**Western blot analyses**

Western blot analysis of rabbit and human tissues, using the anti-human 11β-HSD1 antibody, indicated a high level of cross-species reactivity. A protein band of 34 kDa, the expected size of 11β-HSD1, was identified in both the human and rabbit liver samples (Fig. 6A). Furthermore, this binding could be blocked using primary antibody pre-adsorbed with the immunising peptide (Fig. 6B) indicating that these protein bands were specific to 11β-HSD1. No band was observed in the rabbit kidney negative control tissue (Fig. 6A). A 34 kDa band was also observed on Western blot analysis of ciliary body tissue (Fig. 6A) that could be blocked by use of primary antibody preincubated with immunising peptide.

**Discussion**

11β-HSD1 is a tissue-specific prereceptor regulator of the GC responses (Tomlinson et al. 2004). Its importance as a possible therapeutic target in the management of human obesity and insulin resistance is supported by the development of selective 11β-HSD1 inhibitors that improve glucose tolerance in diabetic mice (Alberts et al. 2002, 2003, Barf et al. 2002). In the human eye, the isozyme has been implicated in the pathogenesis of ocular surface disease and glaucoma, the leading causes of worldwide blindness (Stokes et al. 2000, 2003, Rauz et al. 2003a,b).

In human ocular tissues, 11β-HSD1 is localised to the basal cells of the corneal epithelium (Rauz et al. 2001), NPE (Stokes et al. 2000, Rauz et al. 2001, Suzuki et al. 2001) and possibly the trabecular meshwork (Stokes et al. 2000). To explore the expression and activity of this isozyme further in the anterior segment ocular tissues, we turned to an animal model – the NZWAR. This appeared to be the most appropriate model for our studies, as it has been most widely

The conjunctiva and the cornea are of paramount importance in providing optical clarity and protecting the eye from traumatic insult. The constantly proliferating and differentiating corneal epithelium is known to express SGK1 in the limbal (proliferating) region and both SGK1 and 11β-HSD1 in the basal (differentiated) layer. In order to investigate the autocrine regulation of the cortisone–cortisol shuttle within the GR-rich ocular surface, we examined specific layers of the NZWAR cornea and conjunctiva.

Immunohistochemistry demonstrated 11β-HSD1 expression in the conjunctival and corneal epithelium and, contrary to human studies, the corneal endothelium. Cultured conjunctival fibroblasts did not interconvert cortisone and cortisol, and oxo-reductase and dehydrogenase activity was also absent in naïve cultured corneal stromal keratocytes. In contrast, however, cultured corneal epithelial cells, confirmed by cytokeratin-3 staining, expressed 11β-HSD1 protein as seen by our immunocytochemistry studies, endorsing findings in earlier human tissues (Stokes et al. 2000, Rauz et al. 2001). In addition, these cells demonstrated predominant 11β-HSD1 oxo-reductase (cortisone → cortisol) activity. Natural and synthetic GCs are known to impair wound healing through interactions with the GR. Our studies confirmed that the rate of corneal epithelial cell proliferation was reduced when the cells were coincubated with the active GC, cortisol. More importantly, however, corneal epithelial cell proliferation was reduced by 50% when primary cultures were coincubated with inactive GC, cortisone. These data suggest that the local generation of cortisol from cortisone by 11β-HSD1 may be a critical feature of corneal epithelial renewal. It is possible that selective 11β-HSD1 inhibitors may improve epithelial stability in eyes vulnerable to epithelial defects. Furthermore, as 11β-HSD1 is potently regulated by cytokines and active GC is a key component of immune regulation (Thieringer et al. 2001, Freeman et al. 2005, Zhang et al. 2005), the expression of this isozyme in the conjunctival and corneal epithelia and the autocrine regulation of cortisol may be an integral feature of the ocular surface protective barrier. Further studies are now necessary to tease apart the exact role of 11β-HSD1 in ocular surface renewal, protection and immune regulation.

The role of 11β-HSD1 in corticosteroid receptor-mediated sodium transport in the NPE is an enigma, as in classical tissues such as the kidney and colon, sodium transport is dependent on the mineralocorticoid, aldosterone.
11β-HSD2 protects the MR from cortisol excess (by inactivation to cortisone). Nevertheless, in the human intraocular environment, high ratios of AH cortisol:cortisone indicate a functional intraocular 11β-HSD1 (cortisone → cortisol) (Rauz et al. 2001, 2003b), endorsed by a reduction in IOP (therefore AH secretion) after systemic inhibition of 11β-HSD1. Furthermore, both 11β-HSD1 and -2 activities have been shown in whole rat eye homogenates (Stokes et al. 2000) and abnormal urinary corticosteroid metabolites indicative of 11β-HSD dysregulation are also seen in patients with primary open-angle glaucoma compared with controls (Stokes et al. 2003).

From these data, it is postulated that the presence of a cortisol-generating system within the human eye may have a twofold role: (i) a short-term physiological role, centred on the sodium-transporting NPE linked to the secretion of AH, maintaining a normotensive, intra-ocular environment, where inhibition of the preceptor 11β-HSD1 could result in the suppression of AH formation and a fall in IOP and (ii) a long-term pathological role related to secondary cortisol interactions with the GR within the trabecular meshwork, contributing to aqueous outflow resistance (and raised or uncontrolled IOP) in individuals susceptible to pressure spikes after the use of GCs and the pathogenesis of glaucoma.

There are several ways this hypothesis may be tested. First, by a clinical study designed to show a reduction in intraocular circulating AH cortisol paralleling the reduction in IOP after treatment with systemic carbenoxolone (a non-selective inhibitor of the 11β-HSDs), we confirm the inhibition of intraocular 11β-HSD1 and therefore AH production. This study is not possible as serial AH sampling in the healthy eye would not merit ethical approval. An alternative is to move to an established in vitro model for evaluating NPE pathophysiology, the ODM-2 NPE cell line (Martin-Vasallo et al. 1989, Civan et al. 1996, Carre et al. 2000, Rauz et al. 2003). Our preliminary studies, however, failed to demonstrate 11β-HSD1 oxo-reductase or dehydrogenase activity (S Rauz & E A Walker, unpublished observations) despite the expression at a mRNA level (Rauz et al. 2001). One reason for this anomaly is that ODM-2 is a simian virus-transformed cell line and the transformation process may alter the molecular and cellular characteristics of the cell including 11β-HSD1 activity. It has also been well documented that there is a switch in isozyme expression from 11β-HSD1 to -2 in proliferating cell lines in culture (Rabbit et al. 2002, 2003).

Finally, we considered the anatomy of the ciliary body, which is a highly complex tissue consisting of an outer NPE layer that communicates with the PE forming an anatomical and physiological syncytium overlying a vascular stroma. We hypothesised that 11β-HSD1 function in the NPE is dependent on interactions with the PE and the stroma. By taking precedence from the methodology described in earlier studies establishing a cortisone-activating enzyme in hepatic tissue lysates (Lakshmi & Monder 1988) and ocular homogenates (Stokes et al. 2000), we progressed onto the use of whole ciliary body explants dissected from an animal model for our in vitro analyses described in this present study. In this manner, an attempt was made to maintain the functional syncytium that was confirmed by histological analysis. In order to minimise the preferential 11β-HSD1 dehydrogenase activity in vitro that limits the use of tissue

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**Figure 5** Corneal epithelial cell number estimation using colorimetric cell proliferation assay (CellTitre96 One Solution Proliferation assay; Promega) coincubated with glucocorticoids (GCs). Cell number estimations were performed at 0, 48 and 96 h of culture in serum-containing medium. The optical density at 490 nm reflects the number of intact viable cells. Proliferation was reduced by 50% 96 h after incubation with active (cortisol) and inactive (cortisone) GC (n=3 rabbits; P=0.008).

**Table 1** Western blot analyses of NZWAR and human tissues using (A) the anti-human 11β-HSD1 and (B) anti-human 11β-HSD1 preadsorbed with immunising peptide. A protein band of 34 kDa, the expected size of 11β-HSD1, was identified in the human liver, NZWAR liver and ciliary body tissue (A) that could be blocked using primary antibody preadsorbed with the immunising peptide. No 34 kDa band was observed in NZWAR kidney.
lysates, we carefully dissected ciliary body tissue so that there was minimal cellular destruction.

Our data have confirmed the expression of 11β-HSD1 isozyme to the outer NPE layer. For the first time, we have confidently defined the absence of either isozyme within the inner PE, as the NZWAR is devoid of pigment and thus cannot mask visualisation of the secondary antibody. Most importantly, enzyme assays on the intact ciliary body tissue have endorsed predominant oxo-reductase (cortisone→cortisol) activity and hence this activity is reduced when incubated with an 11β-HSD1 inhibitor. As immunohistochemistry confirmed the absence of 11β-HSD2 expression in the NPE and PE, the minimal dehydrogenase activity was attributed to surgical dissection and tissue necrosis driving the isozyme in a dehydrogenase direction and possibly to 11β-HSD2 dehydrogenase activity from the vascular fraction of the stroma. These data provide evidence that an active 11β-HSD1 is a feature of ciliary body physiology, providing a rationale for further characterisation of the NZWAR as a model to evaluate selective topical 11β-HSD1 inhibitors as a putative treatment for glaucoma.

In summary, the distribution of 11β-HSD1 in the NZWAR eye resembles that of the human eye. This isozyme activates cortisone to cortisol in both ciliary body tissue and corneal epithelium. The NZWAR provides a suitable in vivo model for the further evaluation of 11β-HSD1 activity in the eye, especially its role in the regulation of IOP and corneal epithelial physiology.

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