A novel selective 11β-hydroxysteroid dehydrogenase type 1 inhibitor prevents human adipogenesis

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Abstract

Glucocorticoid excess increases fat mass, preferentially within omental depots; yet circulating cortisol concentrations are normal in most patients with metabolic syndrome (MS). At a pre-receptor level, 11β-hydroxysteroid dehydrogenase type 1 (11β-HSD1) activates cortisol from cortisone locally within adipose tissue, and inhibition of 11β-HSD1 in liver and adipose tissue has been proposed as a novel therapy to treat MS by reducing hepatic glucose output and adiposity. Using a transformed human subcutaneous preadipocyte cell line (Chub-S7) and human primary preadipocytes, we have defined the role of glucocorticoids and 11β-HSD1 in regulating adipose tissue differentiation. Human cells were differentiated with 1-0 μM cortisol (F), or cortisone (E) with or without 100 nM of a highly selective 11β-HSD1 inhibitor PF-877423. 11β-HSD1 mRNA expression increased across adipocyte differentiation (P<0.001, n = 4), which was paralleled by an increase in 11β-HSD1 oxo-reductase activity (from nil on day 0 to 5.9±1.9 pmol/mg per h on day 16, P<0.01, n = 7). Cortisone enhanced adipocyte differentiation; fatty acid-binding protein 4 expression increased 312-fold (P<0.001) and glycerol-3-phosphate dehydrogenase 47-fold (P<0.001) versus controls. This was abolished by co-incubation with PF-877423. In addition, cellular lipid content decreased significantly. These findings were confirmed in the primary cultures of human subcutaneous preadipocytes. The increase in 11β-HSD1 mRNA expression and activity is essential for the induction of human adipogenesis. Blocking adipogenesis with a novel and specific 11β-HSD1 inhibitor may represent a novel approach to treat obesity in patients with MS.

Introduction

Glucocorticoid excess (Cushing’s syndrome) causes visceral obesity, insulin resistance, diabetes mellitus, dyslipidaemia, hypertension and premature vascular mortality; as such it represents an excellent paradigm for patients with the metabolic syndrome (MS). However, Cushing’s syndrome is rare and circulating glucocorticoid levels are usually normal or even slightly reduced in obese patients (Fraser et al 1999). At a cellular level, glucocorticoids within adipose tissue, specifically in omental depots, can be generated from inactive circulating cortisone (in humans) or 11-dehydrocorticosterone (in rodents) through the oxo-reductase activity of 11β-hydroxysteroid dehydrogenase type 1 (11β-HSD1; Bujalska et al 1997b). Transgenic mice with adipose tissue-targeted 11β-HSD1 overexpression develop visceral obesity, insulin resistance, hyperlipidaemia and hypertension without altering circulating glucocorticoids (Masuzaki et al 2001, 2003), while mice with 11β-HSD1 overexpression in the liver develop MS without obesity (Paterson et al 2004). Conversely, global deletion of 11β–HSD1 caused reduced visceral fat accumulation and improved insulin sensitivity on a high fat diet (Kotelevtsev et al 1997, Morton et al 2004). At a molecular level, glucocorticoids exert potent effects upon adipose tissue; in mature rat adipocytes, dexamethasone decreases glucose uptake and oxidation (Olefsky 1975, De et al 1981). Glucocorticoids increase lipolysis by up-regulating the expression of the rate-limiting enzyme hormone-sensitive lipase (Slavin et al 1994), as well as lipogenic enzyme, lipoprotein lipase (Yang et al 1993). In preadipocytes, glucocorticoids are essential for terminal adipogenesis (Hauner et al 1987) and limit cell proliferation (Tomlinson et al 2002). The process of cellular differentiation is a highly synchronized cascade of regulated differentiation-dependent gene expression. Genes such as retinoblastoma proteins that regulate the cell cycle (Richon et al 1997) are followed by adipogenic transcription factors, such as peroxisome proliferator-activated receptor γ (PPARγ) and CCAAT enhancer-binding protein (Rosen & MacDougald 2006). Mature adipocytes express late differentiation genes involved in lipid metabolism and lipid transport including...
glycerol-3-phosphate dehydrogenase (G3PD) and fatty acid-binding protein 4 (FABP4; Hotamisligil et al. 1996); many of these genes are regulated by glucocorticoids (Wu et al. 1996, Rosen & MacDougall 2006). Previously, we have shown that non-selective inhibition of 11β-HSD1 can prevent human adipocyte differentiation in vitro (Bujalska et al. 1999). The potential for therapeutic intervention has been tested in rodent models where selective 11β-HSD1 inhibitors lower plasma glucose, improve insulin sensitivity and in some studies reduce body weight in mice (Alberts et al. 2002, Kershaw et al. 2005). However, the potency of these inhibitors has been variable and there are no data on efficacy in human tissue. We report the development of a selective inhibitor against human 11β-HSD1, PF-877423 (Pfizer Global R&D, La Jolla, CA, USA) and the effect of this compound upon adipogenesis in a well-characterised differentiating human subcutaneous preadipocyte cell line (Darimont et al. 2003, Qiao et al. 2005) and in primary cultures of subcutaneous human preadipocytes.

Research design and methods

Recombinant protein assay

Wild-type recombinant human 11β-HSD1 protein (24–292) was used for studying the inhibitor kinetics. Radio-labelled [1,2-3H]-cortisone was purchased from American Radiolabeled Chemicals Inc (St Louis, MO, USA). NAD (reduced form; NADPH), glucose-6-phosphate (G6P) and G6P dehydrogenase (G6PD) were purchased from Sigma–Aldrich. All the concentrations reported in the following section are final in the assay buffer. In addition, the enzyme concentrations represent the active concentrations that were determined by active-site titration using a tight-binding inhibitor. The experimental data were fitted by using the non-linear regression analysis software, Grafit (Leatherbarrow (2001) GraFit Version 5, Erithacus Software Ltd, Horley, UK).

The measurement of the in vitro 11β-HSD1 activity was performed in a 100 mM triethanolamine buffer (pH 8.0), containing 200 mM NaCl, 0.02% n-dodecyl β-D-maltoside, 5% glycerol and 5 mM β-mercaptoethanol. A typical reaction for the determination of enzyme activity comprised the following: 5 nM enzyme pre-incubated for at least 30 minutes in the assay buffer in the presence of 500 μM NADPH in round-bottom 96-well plates (Costar cat # 3365). Next, the reaction was initiated by adding a regenerating system (consisting of 2 mM G6P, 1 U/ml G6PD and 6 mM MgCl2) and labelled 3H-cortisone as substrate. After an incubation period (30–40 min), 100 μl of the assay mixture were transferred to a second empty round-bottom 96-well plate and mixed with an equal volume of dimethylsulphoxide (DMSO) to quench the reaction. Then, a 15 μl aliquot of the assay solution was loaded into a C-18 column (Polaris C18-A, 50 × 4. 6 mm, 5 μ, 180 Å, Varian; Polaris, Palo Alto, CA, USA connected to an automated High-Throughput Liquid Chromatography instrument (Cohesive Technologies, HTLC, Franklin, MA, USA). The radioactive material from the column was detected with a β-RAM model 3 Radio-HPLC detector (IN/US, Tampa, FL, USA). Substrate and product peaks were separated by using an isocratic mixture of 38:62 methanol to water (v/v) at a flow rate of 1. 0 ml/min. Under these experimental conditions, the retention time for cortisone and cortisol were 4.5 and 5.5 min respectively. The initial reaction velocities recorded were in the linear range and were determined by measuring the peak area for cortisol formation with time.

Recombinant protein kinetic analysis

The inhibition of 11β-HSD1 by PF-877423 was analysed by fitting to the equation described below (Equation (1); Morrison 1969) and provided an accurate measurement for the value of \( K_{i, app} \) at a fixed concentration of cortisone

\[
V_i = V_o \left(1 - \frac{[E_o] + [I_o] + K_{i, app} - \sqrt{([E_o] + [I_o] + K_{i, app})^2 - 4[E_o][I_o]}}{2[E_o]} \right)
\]  

where \([E_o]\) and \([I_o]\) are the active enzyme and inhibitor concentration respectively; \(V_i\) and \(V_o\) are the rates of cortisone reduction in the presence or in the absence of inhibitor respectively. Four \(K_{i, app}\) values were determined by varying the cortisone concentration while keeping the concentration of NADPH constant at 500 μM in the assay buffer. The true inhibition constant, \(K_m\), for PF-877423 was then obtained by plotting the \(K_{i, app}\) values versus the cortisone concentration, \([C_o]\), and fitting the data using Equation (2) for a competitive inhibitor

\[ K_{i, app} = K_m \left(1 + \frac{[C_o]}{K_m} \right) \]  

where \(K_m\) was the Michaelis–Menten constant for cortisone.

HEK293 and Chubb-S7 cell culture

HEK293 cells stably transfected with human 11β-HSD1 (HEK293T1) or 11β-HSD2 (HEK293T2) cDNA as described previously (Bujalska et al. 1997a) were used to study the specificity of inhibitor PF-877423 upon 11β-HSDs. Cells were cultured in minimum essential medium (MEM) media supplemented with 10% fetal bovine serum (FBS) and 1% non-essential fatty acids; for experiments, cells were seeded into 24-well tissue culture dishes and maintained in MEM with 10% FBS until confluence.

The Chub-S7 cell line was derived from human subcutaneous adipose tissue (Darimont et al. 2003) by co-expression of human telomerase reverse transcriptase and papillomavirus E7
oncoprotein (HPV-E7) genes. The cell line has an unlimited life span and the capacity to accumulate lipid without chromosomal alteration. Confluent Chub-S7 cells were cultured in 75 cm² TC flasks in DMEM/Ham's F-12 medium supplemented with 10% FBS. For experiments, cells were seeded in 24-well plates at density 10⁵ cells/well. Chub-S7 were differentiated (up to 21 days) according to Hauner et al. (1987) with 166 nM human insulin (1–9278 Sigma), 1 μM PPARγ agonist (GW1929, Camlab, Cambridge, UK) and, where specified, with the addition of 1 μM cortisol (F). For inhibition studies, Chub-S7 cells were differentiated with 0.5 μM cortisone (E) and an optimised concentration of the selective 11β-HSD1 inhibitor, PF-877423, at a final concentration of 100 nM.

Human subcutaneous stromal-vascular cell differentiation

Confluent human subcutaneous stromal-vascular (s–v) cells (96-well plates) were obtained from Zen-Bio Inc. (Research Triangle Park, NC, USA) and allowed to recover overnight at 37 °C 5% CO₂. The following day, day 0, differentiation was initiated by incubating cells in differentiating media, DM2(E) (Zen-Bio Inc.), which consists of DMEM/Ham’s F-12 medium containing FBS, i-glutamine, penicillin/streptomycin, insulin, indomethacin, 3-isobutyl-1-methylxanthine with 1 μM E and vehicle (DMSO not exceeding 0.1%). To define the effect of the PF-877423 inhibitor upon the differentiation of primary preadipocytes, cultures were supplemented with 300 nM PF-877423 (in DMSO). Control cells cultured without E. Media were replenished and PF-877423 or vehicle treatment repeated every 2–3 days, except when 11β-HSD1 activity was assessed.

Measuring lipid content in human subcutaneous s–v cells

Lipogenesis in human subcutaneous s–v cells was measured as triglyceride accumulation. Cells were carefully washed with PBS and lysed in situ by adding 50 μl/well of Hecameg (10% solution in water – Calbiochem, Nottingham, UK). After gentle shaking at room temperature for 10 min, 200 μl triglyceride (Infinity) reagent (Thermo DMA, Louisville, CO, USA) was added to each well. Plates were read after 10–20 min at 580 nm. Activity has been expressed as cortisol production in pg/ml per 24 h.

Total RNA extraction and RT reaction

Total RNA was extracted from cells using TriReagent (Sigma) according to the manufacturer's protocol. Integrity and concentration of RNA were assessed by electrophoresis and spectrophotometry respectively. Reverse transcription was carried out using AMV and random primers at 37 °C for 1 h. All RT reagents were purchased from Promega.

PCR

Expression studies were carried out using gene specific primers for human 11 β-HSD1, hexose 6-phosphate dehydrogenase (H6PDH), glucocorticoid receptor α (GRα, FABP4), G3PD, glucose transporter 4 (GLUT-4) and PPARγ. All primers were designed using primer 3 software (http://frodo.wi.mit.edu/cgi-bin/primer3/primer3.cgi) and their sequences from 5’ to 3’ are shown in Table 1. Primers for 18S as internal control were purchased from Ambion (Quantum RNTAM Classic 18S Internal Standard #1716; Ambion). PCRs were carried out at 95 °C for 30 s, 60 °C for 30 s and 72 °C for 30 s in 20 μl final volume for 30 cycles.

Real-time PCR

Quantitative mRNA expression levels of 11β-HSD1, H6PDH, GRα, FABP4, G3PD, GLUT-4 and PPARγ2 were measured by real-time PCR using an ABI 7500 system (Perkin–Elmer, Biosystems, Warrington, UK). PCR was performed in 25 μl reactions on 96-well plates. Reactions contained TaqMan universal PCR master mix (Applied Biosystems, Foster City, CA, USA), 900 nmoi primers, 100–200 nmoi TaqMan probe and 25–50 ng cDNA. All reactions were correlated to expression

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of ribosomal 18S rRNA (provided as a pre-optimized mix; Perkin–Elmer) as an internal reference. All target gene probes were labelled with the fluorescent label FAM and the 18S probe with the fluorescent label VIC. Reactions were as follows: 50 °C for 2 min, 95 °C for 10 min, and then 40 cycles of 95 °C for 15 s and 60 °C for 1 min. Data were analysed according to the manufacturer’s guidelines and were obtained as Ct values (the cycle number at which logarithmic PCR plots cross a calculated threshold line) and used to determine ΔCt values (ΔCt of the target gene minus Ct of the internal reference, 18S). Primers and probes for 11β-HSD1, H6PDH and G3PD were designed using PrimerExpress 1.0 software (Applied Biosystems). Sequences from 5’ to 3’ are shown in Table 2. Expression assay kits were purchased from Applied Biosystems to measure the gene expression of GRα, GLUT-4, PPARg2 and FABP4.

### Statistical analysis

Where data were normally distributed, unpaired Student’s t-test was used to compare single treatments with control. If normality tests failed, then non-parametric tests were used. One-way ANOVA on ranks was used to compare multiple treatments (SigmaStat 3.1, Systat Software Inc., Point Richmond, CA, USA). Results were expressed as mean values ± S.D. or S.E.M. values and a P value of < 0.05 was accepted as statistically significant. Statistical analysis on real-time PCR data was performed on mean ΔCt values and not on fold changes.

### Results

**Kinetics of PF-877423 upon recombinant 11β-HSD1 protein**

The potency for PF-877423 was strongly affected by the presence of the substrate in the assay buffer (Fig. 1): $K_{\text{app}}^i$ values increased at high cortisone concentration, suggesting that the inhibitor behaved as a reversible and competitive inhibitor against cortisone. Fitting the experimental data using equation (2) provided a value of 0.2 ± 0.04 and 333.4 ± 109.2 nM for the inhibition constant, $K_i$, and the apparent Michaelis–Menten constant, $K_m$, respectively.

**Specificity of PF-877423**

11β-HSD enzyme assays on HEK293T1 and HEK293T2 cells showed total abolition of dehydrogenase (12.4 ± 1.0 vs 0.2 ± 0.01, % cortisol to cortisone conversion, mean ± S.D.) and oxo-reductase (34.7 ± 0.6 vs 0.4 ± 0.1, % cortisone to cortisol conversion, mean ± S.D.) activities of 11β-HSD1 following incubation with 100 nM PF-877423 for 24 h (Fig. 2A), but PF-877423 had no effect on 11β-HSD2 activity (63.6 ± 4.0 vs 62.2 ± 4.4, % cortisol to cortisone conversion, mean ± S.D., control versus PF-877423 respectively; Fig. 2B). No toxic effects of PF-877423 were observed up to 10 μM concentrations using a commercially available assay kit (CellTiter 96 Aqueous, Promega; data not shown).

### Table 1 Primer sequences for various human genes for the PCR

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward</th>
<th>Reverse</th>
</tr>
</thead>
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<tr>
<td>HSD11B1</td>
<td>ACCAGAGATGCTCCAAAGGAA</td>
<td>ATGCTTCCATGTGCTCTGCTT</td>
</tr>
<tr>
<td>H6PDH</td>
<td>AGAAACGAGACAGCTCTCCAC</td>
<td>GCTGCTGGAAAAAGAACCA</td>
</tr>
<tr>
<td>Grz</td>
<td>TCGACCATGGTCCAGAGAAC</td>
<td>TTTCGGAACCAAGGGGATTT</td>
</tr>
<tr>
<td>GLUT-4</td>
<td>GCCATTGTATCGGCATTCT</td>
<td>CTACCCCTCCTGCTCTGAAG</td>
</tr>
<tr>
<td>PPARγ1</td>
<td>TCTCTGCATGGAAGACACC</td>
<td>GATTATGACACATCCCCAC</td>
</tr>
<tr>
<td>PPARγ2</td>
<td>GGATTCCTCTGACTGACATAC</td>
<td>GATTATGACACATCCCAAC</td>
</tr>
<tr>
<td>G3PD</td>
<td>GAAGAATCTGGAAGGCAGAAGAT</td>
<td>CACCGGCACCTATCATTCTT</td>
</tr>
<tr>
<td>FABP4</td>
<td>CATCAGTGTGAATGCGATG</td>
<td>ATGCGAACATCAGATCCAGGT</td>
</tr>
</tbody>
</table>

Primers were designed using Primer3 software (http://frodo.wi.mit.edu/cgi-bin/primer3).

### Table 2 Primer and probe sequences for various human genes for the real-time PCR

<table>
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<th>Gene</th>
<th>Forward</th>
<th>Reverse</th>
<th>Probe</th>
</tr>
</thead>
<tbody>
<tr>
<td>HSD11B1</td>
<td>AGGAAACGCTCATGGGAGGACTAG</td>
<td>ATGGTGAATATCATCATGAAAAAGATTC</td>
<td>CATGCTCATTCTCAACCACTCA</td>
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<tr>
<td>H6PDH</td>
<td>GGGCCTATGGAACCTCCAA</td>
<td>GACCCACTGTTCTCAGTCTC</td>
<td>CCCTGACCGCTATCATGACAC</td>
</tr>
<tr>
<td>PPARγ2</td>
<td>AGAAAGAGCCGCGCGTTCTC</td>
<td>TCGGTGAGAGCACCACATC</td>
<td>AGGGGCCAACACAGACTTGACACAT</td>
</tr>
<tr>
<td>G3PD</td>
<td>CCAATCGGTATCCGCGGAGAT</td>
<td>TCGTCTAACCCTCTTTAAGAGATATG</td>
<td>AGGGCCATCGTAAGGGCAAACCGC</td>
</tr>
</tbody>
</table>

Primers and probes were designed using PrimerExpress software (Applied Biosystems, UK).
11β-HSD1 is essential for human adipogenesis · IJ BUJALSKA and others 301

Characterisation of chub-S7 cells

At confluence (day 0), Chub-S7 cells did not accumulate lipid droplets (Fig. 3A); however, they readily underwent adipogenesis (shown as oil red O staining) when cultured for 21 days in chemically-defined, serum-free media (166 nM insulin, 1 μM PPARγ agonist and 1 μM F; Fig. 3B). As demonstrated by conventional PCR, confluent undifferentiated Chub-S7 cells expressed GRβ H6PDH and PPARγ1 mRNA but not 11β-HSD1, PPARγ2, GLUT-4, G3PD or FABP4 mRNA (Fig. 3C). In the differentiated Chub-S7 cells, increased expression of adipogenic markers including G3PD and FABP4 was observed. This process resulted in an increase in 11β-HSD1, GLUT-4 and PPARγ2 mRNA levels (Fig. 3D).

Across differentiation, 11β-HSD1 o xo-reductase activity increased significantly; from nil on day 0 to 0.4±0.2 on day 3, 3.5±0.7 on day 5, 8.4±0.14 on day 7, 10.5±1.9 on day 9 and 5.9±1.9 on day 16 (pmol/mg per h, mean±s.d., n=7, all P<0.01 versus previous time point; Fig. 4A). Conventional PCR findings were endorsed and quantified by real-time PCR. Expression of 11β-HSD1 mRNA increased 2.9-fold on day 5, 3.6-fold on day 7, 3.4-fold on day 9 (P<0.01) and 38.1-fold on day 16 (P<0.001) when compared with day 3, n=4 (Fig. 4B).

We observed a transient increase in H6PDH mRNA levels (11β-HSD1 co-factor provider) – 2.9-fold on day 3, 3.5-fold on day 5, 3.7-fold on day 7, 3.4-fold on day 9 and 0.6-fold on day 16 versus day 0, P<0.01 (Fig. 4C) – but there was no significant change in GRβ mRNA during Chub-S7 differentiation (Fig. 4D).

Significant increases in differentiation markers FABP4 (2-fold on day 5 (P<0.01), 38-fold on day 7, 142-fold on day 9 and 870-fold on day 16 versus day 3, P<0.001) and G3PD (4.5-fold on day 7 (P<0.01), 22-fold on day 9 and 380-fold on day 16 versus day 5, P<0.001) were also observed (Fig. 5A and B respectively). When compared with day 7, the expression of adipocyte-specific genes including GLUT-4 and PPARγ2 also increased – 2.1-fold on day 9 and 9.8-fold on day 16, P<0.01 (GLUT-4) and 1.3-fold on day 7 and 2.2-fold on day 16, P<0.01 (PPARγ2; Fig. 5C and D respectively).

Glucocorticoid metabolism and adipogenesis in Chub-S7 cells incubated with PF-877423

Chub-S7 cells differentiated for 10 days with 500 nM cortisone showed increased 11β-HSD1 o xo-reductase activity: 14.6±2.3 (E) versus 3.4±1.3 (control), pmol/mg per h mean±s.e.m., P<0.001 (Fig. 6A), and mRNA expression (14.1-fold versus control; Fig. 6B). Co-incubation with 100 nM PF-877423 abolished this effect: 14.6±2.3 (E) versus 1.3±1.1 (E+PF-877423) vs 0.6±0.5 (PF-877423) pmol/mg per h, mean±s.e.m. (Fig. 6A) and 14.1-fold (E) versus 1.2-fold (E+PF-877423), 11β-HSD1 activity and mRNA respectively (Fig. 6B). Differentiated Chub-S7 cells with E showed increased expression of the adipogenic markers FABP4 (31.2-fold versus control, P<0.001) and G3PD (47-fold versus control, P<0.001) – an effect that was completely abolished by co-incubation with PF-877423 (1.3- and 0.7-fold, FABP4 and G3PD respectively; Fig. 6C and D). The change in adipogenesis following incubation with the 11β-HSD1-specific inhibitor was confirmed visually through staining the cells with oil red O after 21 days of differentiation. A marked increase in the number of red-stained cells was observed in cells differentiated with E or F but not in the presence of PF-877423 (Fig. 6E).

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Human subcutaneous s–v cells differentiation

Human subcutaneous s–v cells differentiated with E + PF-877423 had significantly lower 11β-HSD1 oxo-reductase activity compared with cells differentiated with E at any time point studied (day 6: 154 ± 8 vs 5387 ± 182; day 9: 128 ± 1 vs 5489 ± 230; day 14: 174 ± 18 vs 4041 ± 106; day 20: 409 ± 27 vs 10443 ± 78; day 22: 330 ± 7 vs 11218 ± 193 pg/ml per 24 h, mean ± s.d., P < 0.001, n = 3, E or E + PF-877423 treated respectively (Fig. 7A)). Lipid content in cells differentiated with E and PF-877423 was significantly lower than in cells differentiated with E only and similar to undifferentiated cells (day 16, 0.25 ± 0.03 vs 0.20 ± 0.01; day 20, 0.3 ± 0.02 vs 0.20 ± 0.01; day 22, 0.27 ± 0.01 vs 0.19 ± 0.01; OD (500/660 nm); mean ± s.d., P < 0.01, n = 3, E or E + PF-877423 treated respectively; Fig. 7B).

Discussion

Numerous studies (Hauner et al. 1989, Gregoire et al. 1991, Wolf 1999), including our own (Bujalska et al. 1999, 2002b), have...
defined the permissive role of GCs in inducing adipocyte differentiation. Here, we extend those observations to a novel transformed human adipocyte cell line that should greatly enhance in vitro human-based adipocyte research studies. Within 5 days of incubation in chemically defined media comprising insulin, PPAR agonist and glucocorticoid, impressive differentiation was observed in Chub-S7 cells as assessed by markers including FABP4, G3PD and adipocyte-specific genes such as GLUT-4 and PPARγ2. Adipogenesis was an ongoing process up to 16 days in culture with intracellular lipid stores confirmed by oil red O staining. No significant changes were observed in GR mRNA expression levels across Chub-S7 cell differentiation. P values: **P<0.01, ***P<0.001.

**Figure 5** Gene expression analysed by real-time PCR of adipogenic markers; (A) FABP4: (log scale) 2-fold increase on day 5, 38-fold on day 7, 142-fold on day 9 and 870-fold on day 16 versus day 3; (B) G3PD: 4.5-fold on day 7, 22-fold on day 9 and 380-fold on day 16 versus day 5; (C) PPARγ2: 1.3-fold on day 9 and 2.2-fold on day 16 versus day 7; and (D) GLUT-4: 2.1-fold on day 9 and 9.8-fold on day 16 versus day 7 across Chub-S7 cell differentiation.

In support of these observations, in this study, the rise in H6PDH mRNA levels probably explained the increase in 11β-HSD1-mediated oxo-reductase activity between days 3 and 9 in addition to an increase in 11β-HSD1 mRNA levels. Furthermore, decreased H6PDH mRNA expression seen on day 16 might account for the decline in 11β-HSD1 oxo-reductase activity despite high mRNA. The impressive increase in 11β-HSD1 expression across adipocyte differentiation has been noted recently in mouse 3T3-L1 cells (Kim et al. 2007), as it has been done in other differentiating cell systems including osteoblasts (Eijken et al. 2005). In both cases cells have been incubated with glucocorticoids that themselves positively regulate 11β-HSD1 (Bujalska et al. 1997b); it is not possible from these or previous data sets to conclude whether or not the increased 11β-HSD1 expression is glucocorticoid mediated or a manifestation of the differentiation process per se.

Nevertheless, 11β-HSD1 expression, resulting in a functionally active oxo-reductase, was clearly linked to the differentiation phenotype. Incubation of Chub-S7 cells with inactive steroid, cortisol, induced a degree of adipocyte differentiation similar to that observed with cortisol. This could be explained on the basis of 11β-HSD1-mediated oxo-reductase activity with a $K_m$ for cortisol in Chubb-S7 cells of 100 nM which is similar to the reports in other 11β-HSD1-expressing cell systems (Monder & Lakshmi 1989, Ricketts et al. 1998) including studies on the recombinant enzyme ($K_m = 333$ nM; Walker et al. 2001, Shafqat et al. 2003). Evaluation of the competitive 11β-HSD1 inhibitor, PF-877423, indicated selectivity for the type 1 11β-HSD isoform with a $K_i = 0.2$ nM in the recombinant assay and IC50~5 nM in Chubb-S7 cells. Selectivity of the inhibitor towards 11β-HSD1 isoform but not 11β-HSD2 is crucial as patients with impairment in the latter are presented with severe hypertension, (White et al. 1997).
PF-877423 selectivity was undertaken using a transformed cell line expressing human 11β-HSD2 enzyme where no inhibitory effect was observed. Incubation of cells with 100 nM PF-877423 completely abolished the cortisone induction of FABP4, G3PDH and 11β-HSD1 itself in Chubb-S7 cells. We also demonstrated that the transformed cell line data were consistent with data in primary human subcutaneous primary human subcutaneous preadipocytes. While incubation with cortisone yielded a less impressive effect on adipogenesis in human subcutaneous primary human subcutaneous preadipocytes when compared with Chubb-S7 cells (this might reflect a more advanced adipogenic lineage of primary cultures), inhibition of 11β-HSD1 activity nevertheless reduced the ability of human subcutaneous preadipocytes to differentiate and accumulate lipid.

Whilst the cell line that we have used is subcutaneous in origin and the expression of 11β-HSD1 is higher in omental human preadipocytes (Bujalska et al. 1997b) and therefore we predict that the impact upon omental cells would be more pronounced. We anticipate that this would not lead to preferential loss of subcutaneous fat. Unfortunately, omental cell lines are not available for study and in vivo human clinical studies have not been performed.

Previously, non-selective 11β-HSD inhibitors have been shown to diminish human adipocyte differentiation in vitro (Bujalska et al. 1999) and increase insulin sensitivity in man (Walker et al. 1995), but a lack of isozyme selectivity can cause water retention and hypertension. Since then, patents have been filed on compounds that report to be selective 11β-HSD1 inhibitors. An aryl sulphonamide derivative has been shown to reduce insulin levels and improve glucose tolerance when administered to rodents for 7 days (Alberts et al. 2002). Similar data have been reported for an adamantyl triazole that also reduced body weight and the progression of atherosclerosis in mice (Hermanowski-Vosatka et al. 2005). Transgenic mouse models have established 11β-HSD1 as a novel therapeutic target in this regard; global deletion of 11β-HSD1 results in improved

Figure 6 Chub-S7 cells differentiated with 0.5 µM E and with or without the selective 11β-HSD1 inhibitor PF-877423 at 100 nM. All experiments were carried out on differentiated cells on day 10, n=3. (A) 11β-HSD1 oxo-reductase activity (cortisone to cortisol conversion): 14.6±2.4 (E) versus 1.2±1.1 (E+PF-877423) vs 3.4±1.3 (control), pmol/mg per h mean±s.e.m., P<0.001, (B) 11β-HSD1 mRNA expression and adipogenic markers (C) FABP4, (D) G3PD (log scale, P<0.001 and P<0.001 respectively) and (E) Chub-S7 cells differentiated for 21 days and stained with oil red O: 1) control: 166 nM insulin, PPARγ agonist, 100 nM PF-877423, 2) 0.5 µM E, 3) 0.5 µM E+100 nM PF-877423 and 4) positive control (differentiation with 0.5 µM F). P values: **P<0.01, ***P<0.001.
glucose tolerance through reduced gluconeogenesis and hepatic glucose output (Kotelevtsev et al. 1997, Morton et al. 2001), while targeted overexpression of 11β-HSD1 in adipose tissue recapitulates features of the MS including central adiposity (Masuzaki et al. 2005). In the liver of db/db mice (a model of type 2 diabetes), GR and 11β-HSD1 mRNA expression positively correlated with blood insulin and glucose (Liu et al. 2005). Inhibition of GR and 11β-HSD1 expression either with GR antagonist (RU486; Liu et al. 2005) or by chronic activation of liver X receptor (Liu et al. 2006) attenuated the phenotype of type 2 diabetes in mice.

In humans, the situation is less clear with regard to a role for 11β-HSD1 in the pathogenesis of MS. Some authors have argued for a primary overexpression of 11β-HSD1 in affected patients akin to that observed in some animal models (Paulmyer-Lacroix et al. 2002, Rask et al. 2002) However, while the expression of 11β-HSD1 might be increased, at least in subcutaneous adipose tissue in obese patients with MS and type 2 diabetes (Tomlinson et al. 2002, Alberti et al. 2007) no such increase was observed in omental adipose tissue. Furthermore, based on urinary cortisol/cortisone metabolite ratios and plasma cortisol generation curves following oral cortisol acetate that primarily reflects hepatic 11β-HSD1 expression, a reduction, not an increase, in 11β-HSD1 expression was observed at least in subjects with simple obesity (Stewart et al. 1999, Tomlinson et al. 2004). We have argued that obesity is not primarily a state of 11β-HSD1 overexpression, but that the fall in hepatic 11β-HSD1 activity with increased visceral adiposity might serve as a protective mechanism to offset hepatic glucose output and further adiposity (Valsamakis et al. 2004). Failure of such a switch off in 11β-HSD1 expression might be a factor that determines the onset and persistence of hyperglycaemia in obese patients with type 2 diabetes mellitus (Valsamakis et al. 2004). Further studies are indicated to define the exact role of 11β-HSD1 in the pathophysiology of human MS. Irrespective of the outcome of these studies, selective inhibitors such as PF-877423 offer a real advance in the prevention and treatment of diabetes in subjects with obesity with the potential added benefit of inhibiting adipocyte differentiation. A reduction in adipogenesis specifically within omental depots is likely to further improve the metabolic phenotype of these patients. Clinical studies characterising a novel, potent (K_i 0.2 nM) and fully selective 11β-HSD1 inhibitor preventing lipogenesis in vitro will further our understanding of the role of local glucocorticoid metabolism in human adipose tissue.

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Disclosure

I J B, L L G, J W T and C D declare no conflict of interest. J E, A N F and P A R are employed by Pfizer. P M S is on the advisory board for Pfizer Global R&D.

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