Adenovirus vectors differentially modulate proliferation of pituitary lactotrophs in primary culture in a mitogen and infection time-dependent manner

Zhenhua Wang1,2, Tetsuo Mitsui1, Maho Ishida1 and Jun Arita1

1Department of Physiology, Interdisciplinary Graduate School of Medicine and Engineering, University of Yamanashi, Yamanashi 409-3898, Japan
2Department of Physiology, College of Basic Medicine, China Medical University, Shenyang 110001, People's Republic of China

Abstract

Adenoviruses are powerful, widely utilized vectors for gene transfer. Limitations to their application, however, have not been well described. We used rat pituitary lactotrophs in primary culture as a model for studying how adenovirus vector infection modulates mitogen-induced proliferation and the activities of mitogen signaling pathways. Infection with adenovirus vectors expressing β-galactosidase (βgal) raised basal proliferative levels and blocked fetal bovine serum (FBS)-induced proliferation of lactotrophs, but did not influence the changes in proliferation induced by forskolin, IGF-I, and bromocriptine. The βgal-expressing adenoviruses did not alter the inhibitory action of 17β-estradiol (E2) in the presence of IGF-I; however, they blocked the stimulatory action of E2 in the presence of dextran-coated charcoal-striped serum or forskolin. An adenovirus expressing no protein failed to block FBS-induced proliferation, but was effective in modulating basal proliferative levels and the stimulatory actions of E2. The increased basal proliferative level and the blockade of FBS-induced proliferation were transient, and lost 5 days after infection while the blockade of the stimulatory action of E2 in the presence of forskolin persisted. Adenovirus infection raised basal protein levels of the phosphorylated forms of cAMP response element-binding protein (pCREB) and ERK1/2 and increased the proportion of pCREB-immunoreactive lactotrophs. Adenoviruses also altered estrogen-induced responses in mRNA expression of several estrogen-responsive genes in a gene-specific manner. The results demonstrate that an adenovirus vector differentially interferes with lactotroph proliferation in response to various mitogens. Our results suggest that the effects of the adenovirus that are independent of the genes transferred must be considered when performing adenoviral gene transfer in the primary cultures of normal cells.


Introduction

In recent years virus-mediated gene transfer has been widely applied in primary cultures of normal cells, and to cells in vivo in which conventional methods of gene transfer are unsuccessful. Together with lentiviruses and adeno-associated viruses, replication-defective adenoviruses that are genetically modified by deletions within the viral genome to allow insertion of foreign genes, are the most commonly used viral vectors (Yeh & Perricaudet 1997, Seth 2000). Adenoviruses have a high efficiency of gene transfer, are able to transfer in nonproliferating cells, resulting in sustained high expression of transferred genes, and are easily prepared with high viral titers. Therefore, in addition to experimental studies, adenovirus vectors have been used for gene therapy in human diseases (Crystal et al. 1994). However, adenovirus-mediated gene transfer has disadvantages too. The first generation of adenovirus vectors induces immunological responses when infected in vivo (Dai et al. 1995, Yang et al. 1995, Lieber et al. 1997b). This has only been partially resolved in improved adenovirus vectors (Lieber et al. 1997a, Chirmule et al. 1998).

Adenovirus infection and gene transfer in vivo also interfere with specific cell functions in cultured cells. Adenovirus vectors enhance cell survival and inhibit apoptosis in endothelial (Ramalingam et al. 1999, Zhang et al. 2004) and lung epithelial cells in culture (Flaherty et al. 2004). However, the effect of adenovirus vector infection is poorly understood in endocrine cells. Castro et al. (1997) reported that infection of the primary cultures of anterior pituitary cells with an adenovirus expressing β-galactosidase (βgal), at multiplicities of infection (MOIs) up to 5, did not affect hormone secretion. However, infection at a MOI of 20 increased basal secretion and conversely decreased secretagogue-stimulated secretion of adrenocorticotropin (ACTH) and luteinizing hormone. Adenoviruses expressing green fluorescent protein and βgal also increased the basal secretion of adrenocortical steroids such as cortisol, 17-hydroxyprogesterone, and 11-deoxycorticisol and decreased their ACTH-stimulated secretion in bovine
increase in [3H]thymidine uptake is observed in adenovirus-infected cells, suggesting that the increased basal secretion by adenoviruses is caused by an increased cell number.

Understanding how adenovirus vectors interfere with specific cell functions and intracellular signaling will assist with designing experiments that use these vectors appropriately. Additionally, knowing the limitations of adenovirus vectors will aid with correctly interpreting experimental data. Therefore, in this study, we used the rat pituitary lactotroph in the primary culture as a model for studying how adenovirus vector infection modulates cell proliferation. We found that the first-generation E1/E3-deleted adenovirus vectors differentially modulated lactotroph proliferation in a mitogen and infection-time-dependent manner. In addition, we measured the activities of mitogen signaling pathways in adenovirus-infected cells and correlated them with changes in cell proliferation.

Materials and Methods

Plasmids and adenovirus vectors
Adenovirus vectors were produced using the Adeno-X Expression and Adeno-X Tet-On Expression Systems according to the manufacturer’s protocol (Clontech Laboratories). The adenovirus vector Ad-βgal, which expresses β-gal under the control of the human cytomegalovirus immediate early gene (CMV) promoter, was purchased from Clontech Laboratories. Ad-Tet-On, an adenovirus expressing CMV-driven rtetR-VP16, and Ad-TRE/βgal, an adenovirus expressing β-gal in a tetracycline-regulated manner, have been described in detail previously (Ishida et al. 2007a). An adenovirus expressing firefly luciferase driven by the CMV promoter (Ad-luc) was produced by inserting luciferase cDNA of pGL3-Basic into the NheI/XbaI sites in the pShuttle vector. To produce an adenovirus carrying the rat prolactin (PRL) promoter but no cDNA for exogenous proteins (Ad-emp), a pGL3 vector containing a 3.2-kb fragment of the rat PRL promoter (Ishida et al. 2007b) was digested with HindIII and XbaI to delete luciferase cDNA and then cloned into the NotI/BamHI sites in pTRE-Shuttle2-NruI.MCS.SmaI (Ishida et al. 2007a). These modified pShuttle and pTRE-Shuttle2 vectors were digested with I-CeuI and PstI, and the genes of interest were inserted into a site created with the same restriction enzymes in Adeno-X viral DNA, a replication-defective adenoviral genome. Adeno-X viral DNA was transfected into HEK293 cells using the FuGene 6 Transfection Reagent (Roche Diagnostics) for virus production, and the recombinant adenoviruses were further propagated by serial infection in HEK293 cells. The adenoviruses obtained were purified with the Adeno-X virus purification kit (Clontech Laboratories) and concentrated with the Centricon Centrifugal Filter Unit (Millipore, Bedford, MA, USA). Adenovirus vector titers (infectious units, ifu) were determined using the Adeno-X Rapid Titer Kit (Clontech Laboratories).

Cell culture and viral infection
Experiments were approved by the Ethical Committee of Animal Experiments of the University of Yamanashi. Seven-week-old female Wistar rats, purchased from Japan SLC (Shizuoka, Japan), were used to obtain anterior pituitary cells for the primary culture. Anterior pituitary cells were dispersed as described previously (Kawashima et al. 2002). A 100 μl aliquot of a cell suspension containing 2·0·10^6 pituitary cells was combined with the adenovirus vectors at appropriate MOIs in a 1:1 mixture of Dulbecco’s modified Eagle’s medium (DMEM) and Ham’s Nutrient Mix F-12 without phenol red, containing 15 mM HEPES, penicillin, and streptomycin (DMEM/F12). The cells were then plated in poly-D-lysine-coated 35 mm culture dishes. The cells were then subsequently allowed to attach to the surface of the dishes in a humidified CO2 incubator for 1 h. The pituitary cells were then flooded with 2 ml DMEM/F12 containing 500 ng/ml insulin and preincubated at 37 °C in a humidified atmosphere of 5% CO2 and 95% air for 1 day. After 1 day of preincubation, the pituitary cells were washed with DMEM/F12 to remove adenoviruses and cultures were initiated with a serum-free, chemically defined medium (Kawashima et al. 2002). In experiments using Ad-TRE/βgal and Ad-Tet-On, doxycycline was added at a concentration of 10 μg/ml to induce the reverse Tet repressor protein and thereby β-gal protein. Cultured pituitary cells were treated with the following mitogens: forskolin (1 μM), insulin-like growth factor-I (IGF-I; 30 ng/ml), bromocriptine (10 nM), 17β-estradiol (E2; 1 nM), fetal bovine serum (FBS; 5%), and dextran-coated charcoal-stripped (DCC) horse serum (10%). For labeling proliferating pituitary cells, the cultured cells were treated with 200 μM 5-bromo-2’-deoxyuridine (BrdU) for 3 h prior to the end of culture.

Immunostaining
To determine the BrdU-labeling index, pituitary cells labeled with BrdU were detached from culture dishes at the end of culture by trypsin treatment. The cells were attached to poly-D-lysine-coated glass slides by centrifugation using a cytcentrifuge and fixed with ice-cold methanol. The pituitary cells, attached to glass slides, were double immunostained for BrdU and PRL as described previously (Kawashima et al. 2002). Immunostained slides were covered with PermaFluor (Immunon, Pittsburgh, PA, USA) and examined using a fluorescence microscope equipped with a dual-band mirror unit for fluorescein isothiocyanate (FITC) and Texas Red. A total of 1000 PRL-immunoreactive cells were examined in randomly chosen fields for each slide to determine the BrdU-labeling index, which was the percentage of cells immunoreactive for both PRL and BrdU of the total PRL-immunoreactive cells counted.

For double immunostaining for phosphorylated cAMP response element-binding protein (pCREB) and PRL, the pituitary cells cultured on dishes were fixed with paraformaldehyde, permeabilized with TBS containing 1% Tween 20...
and treated with 3% hydrogen peroxide. After blocking nonspecific binding by using a blocking reagent supplied in the tyramide signal amplification (TSA) fluorescence system (Perkin-Elmer, Boston, MA, USA), the pituitary cells were immunostained for pCREB and PRL as described previously (Ishida et al. 2007a). Fluorescence images of pCREB and PRL labeled with FITC–tyramide and AlexaFluor 594 respectively were captured with a digital CCD camera and stored in a personal computer. The pCREB immunoreactivity was quantified by measuring the fluorescence intensity of nuclear FITC with Photoshop imaging software (Adobe Systems), and a cell with a fluorescence intensity higher than a level of 60–85 was defined as a pCREB-immunoreactive cell. The pituitary cells immunoreactive for both pCREB and PRL were counted in a total of 200 PRL-immunoreactive cells.

βgal histochemistry

Pituitary cells cultured on dishes were fixed with 3% paraformaldehyde in 0.1 M phosphate buffer for 10 min and rinsed with TBS containing 1% Tween 20 (TBST). The pituitary cells were permeabilized with TBS containing 1% Tween 20 for 30 min and incubated with PBS containing 5 mM K$_4$Fe(CN)$_6$, 5 mM K$_3$Fe(CN)$_6$, 2 mM MgCl$_2$, and 1 mg/ml X-gal at 37 °C for 1 h.

Immunoblotting

The pituitary cells, cultured at a cell number of 9 × 10$^5$ cells/35 mm dish, were lysed with 100 μl ice-cold lysis buffer composed of 20 mM Tris, pH 7.6, 137 mM NaCl, 1% Nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS, 2 mM phenylmethylsulfonyl fluoride, 1 mM sodium orthovanadate, and protease inhibitor cocktail (aprotinin, bestatin, leupeptin, and pepstatin A) (sc-24948, Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 15 min at 4 °C. The cell lysates were placed on ice for 30 min in microcentrifuge tubes and vortexed every 10 min, and whole-cell extract of soluble protein was obtained by centrifugation at 17 500 g for 15 min at 4 °C. The cell lysates were counted in a total of 200 PRL-immunoreactive cells.

Quantitative real-time PCR (qRT-PCR)

Expression levels of mRNAs for estrogen-responsive genes were determined by qRT-PCR as described previously (Ishida et al. 2007a). Total pituitary RNA was isolated using the RNasy Mini Kit (Qiagen) and treated with RNase-free DNase I (Qiagen). RNA was reverse transcribed using an oligo (dT)$_2$0 primer and ReverTra Ace reverse transcriptase (Toyobo, Osaka, Japan). Reverse transcriptase reactions were amplified on an ABI Prism 7500 Sequence Detection System (Applied Biosystems, Foster City, CA, USA) using the SYBR Green Real-Time PCR Kit (Toyobo). The primers used are shown in Table 1. The mRNA levels were calculated by the 2$^{-ΔΔCt}$ method using acid ribosomal phosphoprotein P0 (Arbp) as an endogenous reference. The fold change in gene expression of the estradiol treated group was determined relative to the vehicle-treated control in each experiment, and the mean of the fold changes obtained from three independent experiments was then calculated.

Statistical analysis

Experiments were replicated at least three times with separate batches of cell preparations. Differences between groups were analyzed using one-way analysis of variance (ANOVA). The mean ± SEM was calculated.

Table 1 The primers used for quantitative real-time PCR to determine mRNA levels of estrogen-responsive genes

<table>
<thead>
<tr>
<th>Primer</th>
<th>Forward/Reverse</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pim3</td>
<td>Forward</td>
</tr>
<tr>
<td>Plag1</td>
<td>Forward/Reverse</td>
</tr>
<tr>
<td>Stat5a</td>
<td>Forward/Reverse</td>
</tr>
<tr>
<td>Serpine1</td>
<td>Forward/Reverse</td>
</tr>
<tr>
<td>c-myc</td>
<td>Forward/Reverse</td>
</tr>
<tr>
<td>Arbp</td>
<td>Forward/Reverse</td>
</tr>
</tbody>
</table>

Sequence (5′→3′)

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pim3</td>
<td>GGACAGAGTCTCAAGCCTTTATT</td>
</tr>
<tr>
<td>Plag1</td>
<td>CACTTTGAACTTATAAGCCACC</td>
</tr>
<tr>
<td>Stat5a</td>
<td>TTTCGCCAGAAGCTCTTGTGC</td>
</tr>
<tr>
<td>Serpine1</td>
<td>GTTGTACACTAACAGAGAAGCTTC</td>
</tr>
<tr>
<td>c-myc</td>
<td>CACCTGCACCTGCTACCTACA</td>
</tr>
<tr>
<td>Arbp</td>
<td>TTTCACGCAGTGGAAGGTT</td>
</tr>
</tbody>
</table>


www.endocrinology-journals.org

Downloaded from Bioscientifica.com at 10/26/2018 05:24:42AM via free access
Results

Adenovirus vectors differentially modulate mitogen-induced proliferation of lactotrophs

The ability of adenoviruses to express exogenous protein was verified in the primary cultures of the anterior pituitary cells using βgal histochemistry. After a 3-day infection at an MOI of 5 with Ad-βgal, an adenovirus vector encoding the βgal gene downstream of the CMV promoter, many cells exhibited βgal activity (Fig. 1A and B). The βgal activity was also seen when cells were co-infected with Ad-Tet.On and Ad-TRE/βgal, an adenovirus encoding the βgal gene downstream of the tetracycline-responsive expression cassette and treated with the tetracycline analog doxycycline (Fig. 1C).

To determine the effects of adenovirus vector infection on lactotroph proliferation, pituitary cells were infected with no virus, Ad-βgal (5 MOIs), or a combination of Ad-TRE/βgal and Ad-Tet.On (Ad-TRE/βgal + Ad-Tet.On) (2.5 MOIs each). The MOIs were chosen based on our previous study (Ishida et al. 2007a). The cells were then treated with various mitogens known to act on lactotrophs and labeled with BrdU to detect proliferating cells. Infection with Ad-βgal or Ad-TRE/βgal + Ad-Tet.On significantly raised basal levels of lactotroph proliferation in vehicle-treated cells (P < 0.05), an effect that was consistently observed in many experiments shown in Figs 2 and 3. The magnitude of the increase in basal proliferation was always greater in Ad-TRE/βgal + Ad-Tet.On-infected cells than in Ad-βgal-infected cells. Treatment for 24 h with forskolin, an adenylyl cyclase activator, and IGF-I significantly increased lactotroph proliferation in adenovirus-uninfected cells (P < 0.05). The degree of increase was similar in Ad-βgal- or Ad-TRE/βgal + Ad-Tet.On-infected cells (Fig. 2A and B). Dopamine secreted by hypothalamic neurons inhibits a variety of lactotroph functions in vivo and in vitro (Ben-Jonathan & Hnasko 2001). Treatment with bromocriptine, a dopaminergic agonist, significantly decreased lactotroph proliferation in adenovirus-uninfected cells (P < 0.05). Similar decreases were also seen in the adenovirus-infected cells (Fig. 2C). However, the stimulatory action of FBS on lactotroph proliferation, normally seen in adenovirus-uninfected cells, was completely lost in Ad-βgal- or Ad-TRE/βgal + Ad-Tet.On-infected cells (P > 0.05) (Fig. 2D). Dose–response studies revealed that the action of FBS on proliferation was blocked by Ad-βgal infection at MOIs of more than 2 (Fig. 2E).
coated charcoal-treated (DCC) serum alone or in combination with cells were challenged by 24-h treatment with vehicle, 10% dextran-20, and cells were treated with IGF-I alone or in combination with E2.

The pituitary cells were infected with Ad-TRE/bgal gene, and Ad-emp, an adenovirus expressing no exogenous protein. An increase in basal proliferation as observed in Ad-βgal, an adenovirus carrying the luciferase gene instead of the βgal gene, and Ad-emp, an adenovirus expressing no exogenous protein. An increase in basal proliferation as observed in Ad-βgal or Ad-TRE/βgal + Ad-Tet.On-infected cells was also seen in Ad-luc- or Ad-emp-infected cells (P<0.05; Fig. 4A). Like Ad-βgal and Ad-TRE/βgal + Ad-Tet.On, both Ad-luc and Ad-emp blocked the stimulatory action of E2 in the presence of DCC serum or forskolin (P<0.05) (Fig. 4A and B).

Adenovirus vector modulation of lactotroph proliferation via viral proteins and nonspecific transgene proteins

To examine whether the changes in lactotroph proliferation caused by Ad-βgal and Ad-TRE/βgal + Ad-Tet.On were due to either a specific effect of βgal, a common effect of expressed exogenous proteins, or an effect of adenovirus-derived proteins, we measured lactotroph proliferation in cells infected with Ad-luc, an adenovirus carrying the luciferase gene instead of the βgal gene, and Ad-emp, an adenovirus expressing no exogenous protein. An increase in basal proliferation as observed in Ad-βgal or Ad-TRE/βgal + Ad-Tet.On-infected cells was also seen in Ad-luc- or Ad-emp-infected cells (P<0.05; Fig. 4A). Like Ad-βgal and Ad-TRE/βgal + Ad-Tet.On, both Ad-luc and Ad-emp blocked the stimulatory action of E2 in the presence of DCC serum or forskolin (P<0.05) (Fig. 4A and B).
However, FBS-induced proliferation was lost in Ad-luc-infected cells (P<0.05) but not in Ad-emp-infected cells (Fig. 4C).

**Infection time-dependent adenovirus vector modulation of lactotroph proliferation**

In the above-mentioned experiments, adenoviruses were infected for 3 days to determine their effect on lactotroph proliferation. We next asked whether the modulation of proliferation by adenovirus infection was a function of infection time. Time-course studies revealed that the increase in basal proliferation by Ad-βgal or Ad-TRE/βgal+Ad-Tet-On peaked at 3 days after infection and was lost at 5 days, indicating a transient effect on basal proliferation (Fig. 5A). In addition, the blockade of FBS-induced proliferation by these adenoviruses observed at 3 days after infection was also lost at 5 days (Fig. 5B). FBS induced a dramatic proliferative effect in adenovirus-infected cells at this time point (P<0.05). By contrast, the blockade of the stimulatory action of E2 in the presence of forskolin by Ad-βgal or Ad-TRE/βgal+Ad-Tet-On persisted even at 5 days after infection (P<0.05; Fig. 5C).

**Adenovirus infection-induced changes in the activities of mitogenic signaling pathways**

We next determined whether the increased basal proliferation of lactotrophs caused by adenovirus infection was accompanied by changes in the activities of mitogenic signaling pathways in Ad-βgal- and Ad-emp-infected cells. The western blot analysis revealed that expression levels of cyclin D1 protein were increased in these adenovirus-infected cells (Fig. 6A), consistent with the results on proliferation as determined by BrdU labeling. Phosphorylated forms of CREB and ERK1/2 also showed an increase in Ad-βgal or Ad-emp–infected cells compared with uninfected cells while CREB and ERK1 were not changed in either group. There was a modest increase in the expression level of phosphorylated AKT. Because it was possible that the change in pCREB in cell lysates from total pituitary cell populations as determined by western blotting did not necessarily correctly reflect a change in lactotrophs, we determined lactotroph-specific CREB activation in adenovirus-infected cells using quantitative double immunostaining for pCREB and PRL (Ishida et al. 2007a). The percentage of pCREB-immunoreactive lactotrophs was markedly increased by forskolin as a positive control in uninfected cells (Fig. 6B). Infection with Ad-βgal or Ad-emp significantly increased the percentage of pCREB-immunoreactive lactotrophs in vehicle-treated cells (P<0.05).

**Adenovirus infection-induced changes in the expression of estrogen-responsive genes**

The differential modulation by adenovirus vector infection of the stimulatory and inhibitory actions of estrogen on lactotroph proliferation may be accompanied by changes in the response of estrogen-responsive genes to estrogen. We therefore determined E2–induced mRNA expression levels of estrogen-responsive genes to estrogen.

![Figure 6](image-url) Changes in the activities of mitogenic signaling pathways in adenovirus vector-infected cells. (A) Pituitary cells were uninfected or infected with Ad-emp or Ad-βgal at 5 MOIs. After 3 days, cell lysates were collected for the western blot analysis to determine basal levels of cyclin D1 protein and the phosphorylated forms of CREB, ERK1/2, and AKT. β-Actin served as a loading control. (B) After 3 days of infection, cells were treated with either vehicle or 1 μM forskolin for 3 h, fixed with paraformaldehyde, and double-immunostained for PRL and phosphorylated CREB (pCREB). The percentage of pCREB-immunoreactive lactotrophs is expressed as a fold increase relative to the uninfected, vehicle-treated group. Data are means±S.E.M. of triplicate determinations from a representative experiment. *Significantly different from the corresponding vehicle-treated groups at P<0.05; #significantly different from the uninfected, vehicle-treated group at P<0.05.
genes in Ad-βgal- and Ad-emp-infected cells. The estrogen-responsive genes were chosen on the basis of the results in preliminary studies and included the estrogen-upregulated genes Pim3, Stat5a, and c-myc and the estrogen-downregulated genes Plag1 and Serpine1. The E2 responsiveness in their mRNA expression levels were verified in uninfected cells (Fig. 7). E2-induced fold changes in mRNA levels of Pim3 and Plag1 were not altered by either Ad-emp or Ad-βgal infection (P > 0.05). Both adenovirus vectors markedly enhanced the E2 responsiveness of Stat5a mRNA (P < 0.05) while they attenuated the responsiveness of Serpine1 and c-myc mRNAs (P < 0.05).

Discussion

Adenovirus vectors have been successfully used to transfer a gene to endocrine cells, in particular those from the anterior pituitary in vitro and in vivo (Castro et al. 1997, Lee et al. 1999, 2001, Neill et al. 1999, Southgate et al. 2000, Smith–Arica et al. 2001). We used pituitary lactotrophs cultured in serum-free medium as a model and evaluated the undesirable effects of adenovirus vector infection at a low MOI of 5 on cell proliferation. This study demonstrates that infection of lactotrophs with adenovirus vectors expressing βgal markedly increases basal proliferation of lactotrophs. Since a similar increase in basal proliferation by adenovirus vectors is also observed in adrenocortical cells in the primary culture (Alesci et al. 2002), it is likely that adenovirus vectors generally stimulate basal proliferation in endocrine cells in the primary culture. In contrast to the stimulation of basal proliferation, the adenovirus vectors differentially inhibited mitogen-induced proliferation in a mitogen–dependent manner. The adenovirus vectors did not alter the actions of forskolin, IGF-I, and bromocriptine alone, and E2 in combination with IGF-I. They attenuated or blocked the actions of FBS and E2 alone, and E2 in combination with DCC serum. Interestingly, the stimulatory action of E2 in the presence of forskolin, normally observed in uninfected cells, was reversed to inhibition in adenovirus-vector-infected cells. In addition, the modulation of lactotroph proliferation by the adenovirus vectors was infection-time dependent. The increased basal proliferation and the blockade of FBS action were transient and lost at 5 days after infection, while the blockade of the stimulatory action of E2 in combination with forskolin persisted at the same infection time. The results of modulation of lactotroph proliferation by adenovirus vector infection illustrate the importance of considering whether the mitogen used or the infection time is susceptible to adenovirus-mediated interference. For example, in lactotrophs in primary culture, adenovirus vectors can be used in studies using forskolin, IGF-I, and bromocriptine alone, and E2 in combination with IGF-I as mitogens without reservation. In order to apply adenovirus vectors in studies using FBS as a mitogen, FBS must be administered not earlier than 5 days after infection to avoid confounding of the results by the adenovirus.

It is possible that the modulation of lactotroph proliferation, observed in cells infected with adenovirus vectors expressing βgal, is due to a specific effect of βgal expression, a nonspecific effect of exogenous protein expression by the transferred gene, or an effect of adenovirus-derived proteins. We addressed this possibility by using Ad-luc and Ad-emp. Modulation of basal proliferation and proliferation stimulated by estrogen in combination with DCC serum or forskolin, as observed in βgal-expressing adenovirus vector-infected cells, was also seen by infection with Ad-luc and Ad-emp. These results indicate that the viral proteins and not the transgene proteins are responsible for these changes. By contrast, the blockade of FBS-stimulated proliferation was seen following infection with Ad-luc but not with Ad-emp, indicating that the FBS action on lactotroph proliferation is blocked by the expression of an exogenous transgene protein. This study yielded three key observations. First, adenovirus infection modulates mitogen-induced proliferation in a mitogen–dependent manner. Secondly, adenovirus-mediated modulations are caused by both adenovirus-derived proteins and exogenous transgene proteins. Finally, a time course exists for modulation of mitogen activity following adenovirus infection, as exemplified by the difference in the persistence of the changes in basal and estrogen-stimulated proliferation. These findings suggest that the adenovirus-mediated modulation of lactotroph proliferation involves a complicated, multi-factorial mechanism.

Figure 7 Differential effects of adenovirus vector infection on E2-induced mRNA expressions of pituitary estrogen-responsive genes. The pituitary cells were uninfected or infected with Ad-emp or Ad-βgal at 5 MOIs, and, 2 days later, were treated with vehicle or 1 nM E2 for 4 h. Total RNAs were collected for determination of mRNA levels of Pim3, Plag1, Stat5a, Serpine1, and c-myc by means of quantitative real-time PCR. The mRNA levels of the genes were normalized to those of Arbp as an endogenous reference and responses to estrogen were expressed as a fold change in mRNA levels relative to the vehicle-treated group. Data are means ± S.E.M. of triplicate experiments. *Significantly different from the corresponding uninfected groups at P < 0.05.
The identities of the adenovirus-derived proteins that are responsible for the modulation of increased basal proliferation and estrogen-stimulated proliferation are currently undetermined. Adenoviruses have many genes that are capable of promoting transformation, cell proliferation, and transcriptional activity in infected cells (Leppard 1997). An E4 gene product, the E4-orf6/7 protein, may be responsible for the increased basal proliferation of lactotrophs. This is likely because the adenovirus vectors used in this study are human adenovirus serotype 5 with deletions of the early genes E1 and E3. Transcription of E2F-1, a member of the E2F transcription factor family that plays a pivotal role in the regulation of cell proliferation (Stevens & La Thangue 2003) is positively regulated by E2Fs themselves (Hsiao et al. 1994, Johnson et al. 1994), Schaley et al. (2000) have shown that E4-orf6/7 interacts directly with a heterodimer consisting of E2F and the DP protein and induces the binding of E2F to the E2F-1 promoter in infected cells. This leads to stimulation of E2F-1 promoter activity and protein expression. Thus, E4-orf6/7 may stimulate basal proliferation of lactotrophs via E2F induction.

We observed that the increased basal proliferation of lactotrophs by Ad-emp or Ad-βgal was accompanied by increased basal levels of phosphorylated CREB and ERK in pituitary cells. Additionally, the result of a greater proportion of pCREB-immunoreactive lactotrophs, demonstrated by double immunostaining for pCREB and PRL, verifies that pCREB is increased in lactotrophs. Furthermore, the elevated pCREB is not due to an increase in cell number resulting from increased basal proliferation, but to more pCREB per cell. Consistent with these findings, protein kinase A activity and CREB binding in human umbilical vein endothelial cells are stimulated by a βgal-expressing adenovirus vector (Zhang et al. 2005). ERK is activated by adenovirus vectors in a kidney-derived epithelial cell line (Tibbles et al. 2002) and lung epithelial cells (Flaherty et al. 2004). Both the protein kinase A/CREB and ERK signaling pathways are implicated in the regulation of lactotroph proliferation (Suzuki et al. 1999, Fernández et al. 2003, Yamakawa & Arita 2004, Chaturvedi & Sarkar 2005, Ishida et al. 2007). Thus, CREB and ERK activation are likely to be involved in the increased basal proliferation induced by adenovirus vectors, although it is unknown whether the relationship is parallel or causal.

Estrogen regulates both normal and cancer cell proliferation in estrogen-responsive tissues via estrogen receptor-mediated expression of genes that regulate cell proliferation (Weisz & Bresciani 1993, Dickson & Lippman 1995). We investigated whether the influence of adenovirus vectors on estrogen effects was mediated by a change in the response to estrogen of estrogen-responsive genes. We measured, in adenovirus-infected cells, E2-induced expression levels of various estrogen-responsive pituitary cell genes that regulate proliferation (Hennighausen et al. 1997, Obaya et al. 1999, Deneen et al. 2003, Durand et al. 2004, Abdollahi 2007). The E2 responsiveness of mRNA levels of the estrogen-upregulated gene Pim3 and the estrogen–downregulated gene Plagl1 was not altered by adenovirus vectors, the response of the upregulated gene c-myc and downregulated gene Serpine1 was attenuated, and that of Stat5α was enhanced. These results are the first to demonstrate changes in expression of estrogen-responsive genes by adenovirus vector infection. In addition, they suggest that the differential response of the stimulatory and inhibitory actions of estrogen to adenovirus infection is caused by differences in the response to adenovirus vectors among the various estrogen-responsive genes.

Adenovirus vectors are a promising means for gene transfer in normal cells in the primary culture. Ideally, the viral vector by itself should not affect the function of the target cells. We have shown in this study that adenovirus vectors modulate proliferation of pituitary lactotrophs in the primary culture more extensively than was originally thought. The modulation is dependent both on the mitogen and the infection time. The effects result both from viral proteins and delivered gene products. Because interfering modulation by adenovirus vectors may confound the results of experiments using them, a complete understanding of the interactions between adenovirus vectors and target cells is critical to the successful application of adenovirus vectors as gene transfer tools.

Declaration of Interest

The authors declare that there is no conflict of interest that would prejudice the impartiality of this scientific work.

Funding

This work was supported by the Ministry of Education, Science, and Culture of Japan (grant-in-aid for Scientific Research 17590198, 19590224).

Acknowledgements

The authors are grateful to Dr A F Parlow and the NIDDK for providing anti-PRL antibody and Ms W Takahashi for her technical assistance.

References


