Nuclear receptor co-regulator Krüppel-like factor 9 and prohibitin 2 expression in estrogen-induced epithelial cell proliferation in the mouse uterus

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Abstract

Estrogen, acting through its cognate receptor estrogen receptor-α (ESR1), is a critical regulator of uterine endometrial epithelial proliferation. Although the dynamic communication between endometrial stromal (ST) and epithelial cells is considered to be an important component in this process, key molecular players in particular compartments remain poorly defined. Here, we used mice null for Krüppel-like factor 9 (KLF9) to evaluate the contribution of this nuclear protein in ST-epithelial interactions underlying proliferative effects of estrogen. We found that in ovariectomized mice administered estradiol-17β (E2) for 24 h, KLF9 null mutation resulted in lack of E2-induced proliferative response in all endometrial compartments. We demonstrated a negative association between KLF9 expression and nuclear levels of ESR1 transcriptional corepressor prohibitin (PHB) 2 in uterine ST and epithelial cells of E2-treated wild-type (WT) and KLF9 null mice. In early pregnancy uteri of WT mice, the temporal pattern of KLF9 transcript levels was inversely associated with that of PHB2. Deletion of KLF9 up-regulated uterine PHB2 expression and increased PHB2 nuclear localization in endometrial ST and epithelial cells, with no effects on the expression of the related PHB1. In the human endometrial ST cell line treated with E2 for 24 h, KLF9 siRNA targeting augmented PHB2 transcript and increased nuclear PHB2 protein levels, albeit this effect was not to the extent seen in vivo with KLF9 null mutants. Our findings suggest a novel mechanism for control of estrogen-induced luminal epithelial proliferation involving ST KLF9 regulation of paracrine factor(s) to repress epithelial expression of corepressor PHB2.

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Introduction

Estrogen (E) control of cell proliferation is a complex process that is subject to regulation at many levels. The nuclear receptor/transcription factor estrogen receptor-α (ESR1) is the key regulatory participant, transducing E action by binding the ligand to form a complex that, upon homodimerization, interacts with various co-regulators to target E-responsive gene promoters, leading to transcriptional activation, and the synthesis of gene products that modify cellular phenotype and behavior (Tsai & O'Malley 1994, Hall et al. 2001). Nuclear receptor co-regulator proteins, acting as coactivators or corepressors contribute to ESR1 transactivity and exert their effects by direct or indirect interactions with ESR1 (McKenna & O’Malley 2002, Lonard & O’Malley 2006, Green & Carroll 2007). Examples of co-factors that directly interact with ligand-bound ESR1 at its activation function domains include the forkhead protein FOXA1 and the p160 protein family members NCOA7/ERA160/NCOA1, NCOA2, and NCOA3 (Yahata et al. 2001, Hu et al. 2002, Dutertre & Smith 2003, Wang et al. 2007). Co-regulators that influence ESR1 transactivation of target genes indirectly alter chromatin structure by facilitating histone acetylation/deacetylation and methylation/demethylation, and by post-translational modifications of other co-regulators and transcription factors within the transcriptional complex (Stenoien et al. 2001, McKenna & O’Malley 2002, Green & Carroll 2007). The reports on the formation of an extranuclear complex between ligand-bound ESR1 and NCOA3 (Zheng et al. 2005) and the increased transcription of ESR1 target genes upon MAPK phosphorylation of NCOA3 (Font de Mora & Brown 2000) provide evidence for the increasingly complex and less predictable mechanisms underlying co-regulator participation in ESR1 signaling.

We previously reported that the nuclear protein Krüppel-like factor 9 (KLF9), previously designated basic transcription element-binding protein-1 (Imatake et al. 1992) and a member of the Sp family of transcription factors (Suske et al. 2005) may function as a co-regulator of steroid hormone receptor signaling in the uterine endometrium (Zhang et al. 2002, 2003). We found
that KLF9 exerts its effects on progesterone receptor (PGR) and ESR1 actions by distinct mechanisms. KLF9 promotes PGR transactivation through functional and physical interactions with PGR-B and to a lesser extent, PGR-A at progesterone (P)-responsive gene promoters (Zhang et al. 2003, Velarde et al. 2006). By contrast, KLF9 can inhibit ESR1 transactancy in vitro in a high-estrogen environment by promoting estradiol-dependent down-regulation of ESR1 expression through enhancement of the association of ESR1 to GC-rich motifs within its promoter (Velarde et al. 2007). Based on these studies, we concluded that KLF9 may influence both PGR and ESR1 genomic pathways to favor P-induced cell differentiation (Velarde et al. 2007). Nonetheless, while the latter presented an attractive model integrating KLF9 in the opposing actions of E and P in the uterus, it did not account for our earlier findings that in the endogenous E-dominated environment of early pregnancy, uterine endometrial cells of Klf9 null mice exhibited delayed (by 24 h) and attenuated proliferation relative to wild-type (WT) counterparts (Velarde et al. 2005). The decreased numbers of implanting embryos in Klf9 null mutants suggested that the altered pattern of proliferation of endometrial cells with Klf9 ablation resulted in developmental asynchrony between the uterine luminal epithelium (LE) and the implantation-ready embryo, leading to subfertility (Simmen et al. 2004, Velarde et al. 2005). Thus, KLF9 may function as a positive regulator of ESR1 signaling to influence cell proliferation, with important consequences on pregnancy outcome.

Prohibitin (PHB) 2, also designated as repressor of estrogen receptor activity (REA), is a 37 kDa protein exhibiting high homology to the putative tumor suppressor protein PHB1 (Montano et al. 1999). PHB1 and PHB2 are highly conserved proteins in eukaryotic cells, with similar functions as inhibitors of cellular proliferation (Mishra et al. 2006). Unlike PHB1, however, which represses signaling of numerous steroid hormone receptors (Wang et al. 2004, Mussi et al. 2006, Gamble et al. 2007, He et al. 2007), PHB2 demonstrates selective repression of ER activity (Montano et al. 1999), possibly in a cell-specific context (Wang et al. 2004), and cooperates with the chicken ovalbumin upstream promoter binding transcription factors I and II to decrease transcription (Kurtev et al. 2004). PHB2 inhibits ESR1 transcriptional activity by competing with p160 co-regulators such as NCOA1 and NCOA3 for binding to ESR1 in the presence of E (Montano et al. 1999, Delage-Morroux et al. 2000, Wang et al. 2004), and by recruiting class I and class II histone deacetylases (Kurtev et al. 2004). Whereas genetic deletion of both Phb2 alleles resulted in embryonic lethality, heterozygous mice displayed phenotypes that are characteristic of overactivated ESR1 signaling including uterine epithelial hyperplasia coincident with higher expression levels of E-responsive genes, enhanced mammary gland morphogenesis, and delayed mammary gland involution (Park et al. 2005, Mussi et al. 2006). These collective results indicate that PHB2 functions as an important mediator of E action in vivo.

In the present study, we show that endometrial epithelial cells of ovariectomized (Ovx) mice with Klf9 null mutation were unresponsive to E2-induced proliferation. We hypothesized that stromal (ST) KLF9 may mediate E2 effects on uterine epithelial cell proliferation by influencing the expression of specific ESR1 co-regulators. We demonstrate that the KLF9-mediated increase in proliferation with E2 was negatively associated with uterine Phb2, but not Phb1, expression and with nuclear localization of PHB2 in all endometrial compartments. We further show that this negative linkage between KLF9 and nuclear PHB2 also occurs during the physiological condition of early pregnancy and in an E2-treated human endometrial ST cell line (HESC). Our results suggest that the repression of epithelial PHB2 expression involving ST KLF9 signaling may be a necessary component in the paracrine regulation of uterine epithelial proliferation by estrogen.

Materials and Methods

Animals and treatments

Experiments were carried out in accordance with the protocols approved by the Institutional Animal Care and Use Committee of the University of Arkansas for Medical Sciences. WT and Klf9 mutant (KO) mice lines were propagated and genotyped as described previously (Morita et al. 2003, Simmen et al. 2004). For steroid hormone treatment studies, ~8-week-old WT and KO female littermates were subjected to bilateral ovariectomy. Two weeks later, mice were treated with vehicle (sesame oil; 0:25 ml) or 17β-estradiol (E2, 125 ng in 0:25 ml sesame oil), and uteri were collected after 24 h. Uterine tissues from WT and KO mice at day post-coitum (dpc) 2.5, 3.5, and 4.5 were isolated as described previously (Velarde et al. 2005). The presence of vaginal plug was considered 0-5 dpc.

Immunohistochemistry

Paraffin-embedded uteri from Ovx WT and KO females (n=3–5 mice per genotype per treatment) were serially sectioned, dewaxed in xylene, and rehydrated through a graded alcohol series. Tissue sections were treated with 3% H2O2 to quench endogenous peroxidase activity. Antigen unmasking was performed by boiling the sections in Citra (Biogenex, San Ramon, CA, USA) in a microwave oven for 105 s at power 10 and then for 10 min at power 1. After blocking with horse IgG (Vectastain Elite ABC kit, Vector Laboratories, Inc., Burlingame, CA, USA) for 30 min, sections were incubated with the following antibodies to detect expression of proteins: a) mouse monoclonal antibody to proliferating cell nuclear antigen (PCNA) (PC-10, Dako, Carpinteria, CA, USA) at 1:500 dilution; b) rabbit polyclonal antibody to ESR1 (MC-20, Santa Cruz Biotechnology, Inc.) at 1:500 dilution; and c) rabbit polyclonal antibody to PHB2 (Bethyl Laboratories Inc., Montgomery, TX, USA) at 1:250 dilution. Incubations with anti-PCNA and ESR1 were carried out for 1 h at room temperature, whereas incubation with anti-PHB2 antibody was performed overnight at room
temperature. Following incubation with secondary antibody (horse anti-mouse IgG at 1:1600 dilution or goat anti-rabbit IgG at 1:2000 dilution) for 30 min, sections were stained with 3,3'-diaminobenzidine tetra-hydrochloride (Dako) and counterstained with hematoxylin. Control sections were processed similarly with the omission of the primary antibody. A total of 1000 ST, 300 luminal epithelial, and 200 glandular epithelial cells were counted on average from at least three randomly selected fields (200× magnification) per slide; one to two slides/mouse with n=3–5 mice per treatment group per genotype, were evaluated. Results are expressed as % nuclear-immunopositive cells ((number of nuclear positively staining cells/number of total cells counted)×100).

RNA isolation and quantitative RT-PCR

Total cellular RNA was prepared from whole uteri or cells by TRIzol reagent (Invitrogen). Integrity of isolated RNAs was confirmed using the RNA 6000 NanoLabChip kit (Agilent Biotechnologies, Palo Alto, CA, USA). RNA samples were reverse transcribed using random primers and a cDNA synthesis kit (Applied Biosystems, Foster City, CA, USA). SYBR Green quantitative RT-PCR was performed as described previously (Velarde et al. 2006). Primer sets were designed to flank an intron to prevent amplification of genomic DNA, using PrimerExpress (Applied Biosystems). Synthetic oligonucleotides were obtained from Integrated DNA Technologies, Inc. (Coralville, IA, USA). The sense and antisense primers for the mouse genes, and the resultant PCR product sizes (in parentheses) were: Klf9, 5'-CGT TGC CCA CTG TGT GAG AA-3' and 5'-TTG ATC ATG CTG -3' (92 bp); Phb2/Rea, 5'-AGC AGG AAC AGC AC AGA AGA-3' and 5'-CGG AGC TTG ATA TAG CCA GGA T-3' (103 bp); and Phb1, 5'-GCC CGG TCC TTT GAC GAC ATT CCA AAC CAA TTA-3' and 5'-TGTGAT ATT GAC TTG CTG CAA GTC T-3' (101 bp). For each sample, gene expression was normalized to that of cyclophilin A (Ppia) (sense: 5'-AGA TGC CAG GAC CTG TAT GCT T-3'; antisense: 5'-TGT GCC AGG GTG GTG ACT TTA-3') as internal reference. Human primers (sense and antisense respectively) for KLF9, PHB2/REA, and the internal reference gene RPL7 were as follows: KLF9, 5'-TGG CTG TGG GAA AGT CTA TGG-3' and 5'-CTC GTC TGA GCG GGA GAAA CT-3' (124 bp); PHB2/REA, 5'-GAA CAG CGG CAG AAA ATT GTG-3' and 5'-CGA ATC TGG CGA AGT TTG ATG T-3' (105 bp), and RPL7, 5'-TCGCTG GCC AGA AAC CCT TAA-3' and 5'-GCT TCC TCC TTG CCT TGC G-3' (110 bp).

Cell culture, transfection with siRNAs, immunofluorescence, and western blots

The HESC was a generous gift from Dr Graziella Krikun (Yale University, New Haven, CT, USA). The cell line was maintained in phenol red-free DMEM and Ham’s F12 (1:1 vol/vol)-medium supplemented with 10% charcoal-stripped calf serum and 1% antibiotic/antimycotic solution in an atmosphere of 5% CO2, as described previously (Krikun et al. 2004). Transfection with Klf9 or scrambled (non-specific) siRNAs (Dharmacon, Waltham, MA, USA) at a final concentration of 50 nM was performed using Lipofectamine 2000 according to the manufacturer’s protocol when cells were ~60% confluent. After 6 h, the transfection mix was replaced with the medium containing 2% charcoal-stripped calf serum. Cells were incubated for an additional 24 h in 2% charcoal-stripped calf serum with added E2 (10 nM) and collected for RNA analyses. Immunofluorescence was performed essentially as described previously (Velarde et al. 2007). In brief, cells were seeded on sterile 22 mm glass cover slides at a density of 1-8 x 10^5 cells per well and grown overnight. After transfection with siRNAs, cells were incubated in the medium containing E2 (10 nM) in DMSO for 24 h. Cells were fixed in 4% paraformaldehyde for 10 min and permeabilized in 0-1% Triton-PBS for 45 min, with 1×PBS washes after each step. Cells were sequentially incubated in goat serum blocking solution (Vectastain Elite ABC kit) for 30 min and with anti-PHB2 polyclonal antibody (1:250 dilution) overnight. After incubation with biotinylated secondary antibody (Vectastain Elite ABC kit), fluorescence signals were developed, visualized, and quantified for percentage of nuclear staining cells (relative to total number of cells; ~250 per treatment group; Velarde et al. 2007). Western immunoblots were performed (Velarde et al. 2007) using anti-rat KLF9 antibody generated in-house (Zhang et al. 2002) and anti-α-actinin antibody (Santa Cruz Biotechnology) for loading control.

Data analysis

Values are presented as mean±s.e.m. Data were analyzed using SigmaStat (SPSS Science, Chicago, IL, USA) and evaluated for differences between groups by Student’s t-test or two-way ANOVA, followed by Tukey’s test. P≤0.05 were considered statistically significant.

Results

Uterine endometrial cells are unresponsive to E2-induced proliferation with Klf9 null mutation

To evaluate the proliferative response of uterine endometrial cells to E as a function of KLF9 expression status, age-matched Ovx WT and Klf9 KO littermates were subcutaneously administered E2 in sesame oil or sesame oil alone (control), and corresponding uteri were isolated after 24 h. Tissue sections from WT and KO mice were immunostained for the proliferation marker PCNA (Fig. 1A), and the numbers of immunopositive cells in each endometrial compartment were counted. In WT mice, E2 administration increased PCNA expression in LE, glandular epithelium (GE), and ST cells by at least fourfold over those of corresponding control cells (Fig. 1B). Basal and E2-induced
PCNA expression was significantly greater for LE than for GE and ST. By contrast, PCNA immunoreactivity in all endometrial compartments of E2-treated Klf9 null mice did not rise above those of corresponding control cells.

Since E2 predominantly signals through ESR1 to exert its proliferative effects in the uterus (Dupont et al. 2000), the nuclear levels of this protein in endometrial compartments were evaluated by immunohistochemistry as a function of KLF9 status. Ovx oil-treated WT mice showed robust ESR1 expression in the uterine endometrium. E2 treatment had no effect on nuclear levels of this protein for all cell types, in the presence or absence of KLF9. The percentages of cells positive for nuclear ESR1 in LE from control (15.10 ± 7.43%) and E2-treated (22.91 ± 5.91%) WT mice did not differ from those of LE in corresponding control (10.67 ± 2.25%) and E2-treated Klf9 null mice (11.86 ± 2.34%). Similarly, ST cells from WT (control: 45.02 ± 10.38%; E2-treated: 38.51 ± 6.19%) and Klf9 null (control: 35.86 ± 11.59%; E2-treated: 37.78 ± 6.27%) mice and GE cells from WT (control: 29.21 ± 11.42%; E2-treated: 36.23 ± 13.56%) and Klf9 null (control: 21.44 ± 6.65%; E2-treated: 36.23 ± 13.56%) mice showed no differences in percentages of cells positive for nuclear ESR1.

Figure 1 Nuclear PCNA levels in uterine endometrial cells of oil (vehicle) and E2-treated ovariectomized WT and Klf9 null mice. (A) Representative PCNA immunostaining of glandular epithelium (GE), luminal epithelium (LE), and stromal (ST) compartments are shown at 200× magnification. (B) The percentages of nuclear staining cells are presented as mean ± s.e.m. (n=3–5 mice per treatment per genotype). Significant differences were identified by two-way ANOVA, followed by Tukey’s test. Means with different superscripts differed at P<0.05.

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KLF9 is negatively associated with uterine PHB2 expression and nuclear localization

To understand the mechanism by which ablation of Klf9 expression (predominantly in the ST; Simmen et al. 2004) resulted in the lack of E2-mediated proliferative response in all endometrial compartments, we examined the possibility that KLF9 negatively regulates the expression of the selective ESR1 transcriptional corepressor PHB2 (Montano et al. 1999). Uterine tissues from control (oil) and E2-treated Ovx WT and KO mice were quantified for Phb2 mRNA abundance by qPCR. Control (oil-treated) WT and KO mice showed comparable levels of Phb2 transcripts (Fig. 2A). E2 administration appeared to decrease (\( P < 0.10 \)) Phb2 expression in WT mice, relative to corresponding control, and this trend was reversed with loss of Klf9 (Fig. 2A). The expression of the related gene Phb1 was not affected by E2 or by KLF9 expression status (Fig. 2B). Similarly, E2 had no effect on Klf9 transcript levels (Fig. 2C).

We evaluated the presence of PHB2 protein in uterine tissue sections by immunohistochemistry to determine its spatial distribution pattern, as a function of E2 treatment and KLF9 expression. Using a previously characterized anti-PHB2 antibody (Montano et al. 1999), we found immunoreactive PHB2 in both cytoplasmic and nuclear compartments of GE, LE, and ST cells (Fig. 3A), consistent with the reported localization of this protein in the mitochondrion and nucleus (Fusaro et al. 2003, Mishra et al. 2006). Since E-dependent interactions between PHB2 and ESR1 occur predominantly in the nucleus to inhibit ESR1 transactivity, nuclear PHB2 expression in endometrial GE, LE, and ST cells was determined in control and E2-treated mice. In oil-treated Ovx WT and KO mouse uteri, the amount of nuclear PHB2 (measured as %PHB2-immunopositive cells) were significantly greater in LE (\( \sim 20\% \)) than in GE (10%) and ST (1%) cells (Fig. 3B). With E2 treatment, the levels of nuclear PHB2 in GE and ST cells of WT uteri did not differ from those of oil-treated controls whereas in LE cells, nuclear PHB2 levels showed a trend to decrease (Fig. 3B), following that seen for corresponding mRNA in whole uteri (Fig. 2A). Loss of Klf9 reversed the possible inhibitory effect of E2 on nuclear PHB2 expression in LE cells to the level of control (non-E2 treated) cells, and increased the percent of nuclear PHB2-immunopositive GE and ST cells, with the latter demonstrating a most dramatic (\( \sim 40\text{-fold} \)) enhancement (Fig. 3B).

Uterine PHB2 and KLF9 expression are inversely associated in early pregnancy

We have previously shown that during the E-dominated condition of early pregnancy at dpc 2-5, uterine endometrial cells of Klf9 null mice had negligible BrdU labeling relative to those of WT mice; however, KO cells displayed peak BrdU labeling 1 day later at dpc 3-5, similar to the level of WT cells at dpc 2-5 (Velarde et al. 2005). To determine whether the delayed proliferative response to E2 in Klf9 null uteri was associated with a transient increase in Phb2 expression, the same WT and KO uterine tissues previously analyzed for proliferation status (Velarde et al. 2005) were evaluated for Phb2 mRNA abundance. In WT mice, Phb2 expression was low at dpc 2-5, significantly increased at dpc 3-5, and remained at that level at dpc 4-5 (Fig. 4A). By contrast, Phb1 mRNA abundance in WT mice (relative to WT dpc 2-5) progressively decreased at early pregnancy (Fig. 4B). The large variations in Phb1 abundance reflect its very low levels of expression, relative to Phb2. The pattern of Phb2 mRNA expression was inversely associated with
that of Klf9 (Fig. 4C), the latter concordant with the peak of proliferation of luminal epithelial cells at dpc 2.5 and subsequently decreasing at dpc 3.5 and 4.5 (Velarde et al. 2005). In KO mice, Phb2 gene expression was significantly higher than for WT mice at dpc 2.5 and 4.5, while at dpc 3.5, no difference in Phb2 expression levels was noted between genotypes (Fig. 4A). Phb1 expression was not affected by the loss of Klf9 expression (Fig. 4B).

Next, immunoreactive PHB2 was localized in uteri of WT and KO mice at dpc 2.5, 3.5, and 4.5. Representative immunostained uterine sections from WT and KO mice at dpc 2.5 are shown (Fig. 5A). In WT mice, immunoreactive PHB2 was present in all endometrial cell types and in the myometrium (not shown), and was localized to both cytoplasmic and nuclear compartments (Fig. 5B). The percentage of nuclear PHB2-positive cells in WT mice was higher in GE and ST cells at dpc 3.5 and 4.5 than at dpc 2.5 (Fig. 5C), following the temporal pattern observed for corresponding mRNA in whole uteri (Fig. 4A). Phb1 expression was not affected by the loss of Klf9 expression (Fig. 4B).

Figure 3 PHB2 levels in uterine endometrial cells of oil (vehicle) and E2-treated ovariectomized WT and Klf9 null mice. (A) Representative PHB2 immunostaining of GE, LE, and ST cells of E2-treated WT and Klf9 null (KO) mice are shown at 400× magnification to demonstrate nuclear localization of immunoreactive PHB2. Negative control was tissue from E2-treated WT mice processed similarly except for omission of primary antibody. (B) The percentages of PHB2 nuclear staining cells are presented as mean ± S.E.M. (n=3–5 mice per treatment per genotype). Significant differences were identified by two-way ANOVA, followed by Tukey’s test. Means with different superscripts differed at P<0.05.

To evaluate if KLF9 directly inhibits PHB2 expression in an E2-dominated environment as suggested in vivo (above), a HESC was used in siRNA targeting of human KLF9 mRNA. PHB2 expression in siKlf9 mRNA-transfected cells, relative to those transfected with non-specific (scrambled) siRNAs, was subsequently determined at the transcript and nuclear protein levels. Transfection of HESC with a pool of Klf9 siRNAs (Dharmacon) followed by E2 treatment diminished Klf9 gene expression by at least 70%, relative to similarly treated cells transfected with scrambled siRNAs (Fig. 6A); this level of knockdown at the transcript level resulted in a comparable decrease in KLF9 protein levels, as determined by western blots (Fig. 6B). The decrease in Klf9 expression significantly (P=0.009), albeit modestly increased (by 20%) PHB2 transcript levels (Fig. 6A). Immunofluorescence studies with anti-PHB2 antibody confirmed the up-regulated expression of PHB2 at the level of the protein (Fig. 6C). Relative to scrambled siRNA, Klf9 siRNA increased the percentage of nuclear-localized PHB2 in E2-treated HESC (Fig. 6D).
Parallel studies using anti-ESR1 antibody showed no effect of KLF9 status on the nuclear levels of this protein (data not shown), consistent with the findings for uterine ST cells of Ovx, E2-treated Klf9 null mice (described in text, above).

**Discussion**

Results from the present study suggest the participation of the transcription factor KLF9 in paracrine signaling to regulate E2-induced proliferation of uterine endometrial epithelial cells. Using Ovx, E2-treated Klf9 null mutant mice and corresponding WT littermates as controls, we showed that the ST KLF9 regulation of the proliferative response of endometrial LE cells to E2 is negatively associated with uterine Phb2 expression and nuclear localization of PHB2 in endometrial epithelial cells. We confirmed the physiological relevance of the negative linkage between KLF9 and PHB2 by demonstrating that the transiently lower proliferative response of uterine endometrial epithelial cells of Klf9 null mutants relative to WT counterparts at dpc 2.5 (Velarde et al. 2005) corresponded with increased expression of nuclear PHB2 expression in these cells. Finally, we demonstrated that in the E2-treated HESC, a reduction in KLF9 expression resulted in increased transcript and nuclear protein levels for PHB2. Taken together, our results suggest a working model whereby E2 control of epithelial cell proliferation can be dynamically influenced by ST KLF9 (Fig. 7). By attenuating expression of ST ESR1 corepressor PHB2 that can interact with ESR1 coactivators such as NCOA1 (Montano et al. 1999) and histone deacetylases (Kurtev et al. 2004), KLF9 may promote ESR1-mediated transactivation of yet unknown paracrine factor(s) to repress epithelial expression of PHB2, leading to increased proliferative responsiveness to E2.

The present study provides the first direct evidence for KLF9 involvement in E2-induced proliferation of uterine endometrial cells. Our previous studies using early pregnant mouse uteri hinted at this possibility, since maximal BrdU labeling of endometrial epithelial cells occurred at the E-dominated uterine environment of dpc 2.5 (Velarde et al. 2005), coincident with the highest uterine ST Klf9 expression (this study). Since uterine expression of Klf9 is predominantly ST and is lacking in luminal epithelial cells (Simmen et al. 2004), the absence of proliferative response to E2 of the latter endometrial compartment in Ovx, E2-treated Klf9 null mice suggests paracrine control by an E2-induced ST-derived growth-regulatory factor(s) whose synthesis may involve functional KLF9/ESR1 interactions. The latter is in agreement with previous reports that ST-localized ESR1 mediates the mitogenic effects of E2 on neighboring LE cells (Cooke et al. 1997) and that KLF9 can influence ESR1 transactivity in endometrial epithelial cells (Velarde et al. 2007). Although the identity of this putative KLF9/ESR1-regulated paracrine factor(s) is currently unknown, our results suggest that this factor(s) may be involved in the inhibition of PHB2 expression in luminal epithelial cells to allow these cells, which express

![Figure 4](https://www.endocrinology-journals.org)
ESR1, to optimally respond to E2. Interestingly, the negative relationship between PHB2 and KLF9 was not extended to the related Phb1 gene, whose gene product also functions as a repressor of ESR1 signaling (Mishra et al. 2006, He et al. 2007). However, we cannot exclude the possibility that an association between KLF9 and PHB1 expression may be manifest at the level of PHB1 protein.

To further investigate the molecular interplay between ESR1 and KLF9 in ST cells, we evaluated the effect of Klf9 knockdown as a function of E2 treatment on PHB2 expression using the HESC. Since loss of KLF9 expression in vivo was associated with enhanced expression of nuclear PHB2 in ST (by 40-fold), more so than in GE and LE (by threefold) cells, it is possible that KLF9 regulation of PHB2 levels in ST may be distinct from that in LE. Our findings that abrogation of Klf9 gene expression resulted in increased PHB2 transcript and nuclear PHB2 protein levels confirm the negative association of KLF9 and PHB2 in vivo. However, the in vitro effects were modest and did not recapitulate the two- to threefold (pregnancy) and 40-fold (Ovx/E2 model) increases in PHB2 expression seen in vivo. Albeit this difference may be a function of the limited time of exposure to E2 in vitro, the in vivo data were also observed 24 h post-E2, raising the strong possibility of a requirement for ST and LE communication (paracrine signaling) in this E2-mediated process.

Our studies indicate that the regulation of uterine PHB2 expression involving KLF9 is complex and could occur at the levels of RNA, protein, and cellular localization. Indeed, the lack of a clear-cut association between nuclear PHB2 and KLF9 expression (this study) with LE proliferation status...
Figure 6. PHB2 levels in human endometrial stromal cells as a function of KLF9 expression status. HESC were incubated in phenol-red free DMEM/Ham’s F12 medium containing 10% charcoal-stripped serum and E2 (10 nM) in the presence of siRNA to scrambled (negative control) mRNA or siRNA to KLF9 (50 nM). (A) Harvested cells were analyzed for KLF9 and PHB2 transcripts by qPCR and normalized to control gene Rpl7. Results (mean ± S.E.M.) relative to cells treated with scrambled siRNA shown are representative of two to three independent experiments, with each experiment conducted in triplicates. (B) Western blots of whole cell lysates prepared from HESC cells as a function of Klf9 knockdown. Cells transfected with siRNA to scrambled sequence (negative control) or to Klf9 were subjected to western blots using anti-KLF9 or anti-α-actinin antibodies. Each lane represents an independent experiment. (C) HESC treated with E2 for 24 h were immunostained for PHB2/REA and counterstained for DAPI. Immunopositive cells were visualized using fluorescent antibodies. Overlay of anti-PHB2 (red) and DAPI (blue) staining cells showed nuclear localization of PHB2. (D) The percentages of nuclear PHB2 staining cells were expressed relative to the total number of cells counted. Results (mean ± S.E.M.) were normalized to those of scrambled siRNA-treated cells and were from two independent experiments, with each experiment performed in triplicates. Differences between groups were determined by t-test (P=0.06).
(Velarde et al. 2005) at dpc 3.5 and 4.5, in contrast to that seen at dpc 2.5, and the loss of the inverse association of KLF9 and nuclear PHB2 levels at dpc 4.5 in LE as well as in GE and ST, suggests the additional contributions of progesterone, other KLF9-related proteins that may compensate for the loss of KLF9 function (Simmen et al. 2004) and possibly, the attaching embryo in the control of LE proliferation at early pregnancy. Since control of PHB2 levels may constitute part of a homeostatic mechanism to allow for the appropriate proliferative response of uterine cells to E2, further investigations into the function and molecular regulation of PHB2 in the uterine endometrium by KLF9 in concert with other nuclear steroid receptor co-regulators should constitute an important area for future research.

In summary, our studies define a potential mechanism whereby control of E-induced endometrial epithelial cell proliferation can be dynamically regulated by ST KLF9 through its indirect inhibition of nuclear PHB2 expression in epithelial cells. Results suggest the participation of KLF9 in E2-dependent synthesis of an ST-derived paracrine factor(s) that mediates this epithelial mitogenic response to estrogen. While the global significance of KLF9 regulation of ESR1 signaling mediated by PHB2 in luminal epithelial cells is currently unclear, the sub-fertility phenotype of Klf9 null mice (Simmen et al. 2004) suggests physiological relevance of KLF9 during the E2-dominated period of early pregnancy prior to implantation (i.e., predecidual ST) and underscores the delicate control of nuclear coactivator and corepressor expression to achieve the requisite uterine response to estrogen for successful pregnancy.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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