Interleukin 10 deficiency attenuates induction of anti-TSH receptor antibodies and hyperthyroidism in a mouse Graves' model

Ikuko Ueki¹,², Norio Abiru², Kentaro Kawagoe¹ and Yuji Nagayama¹

¹Department of Molecular Medicine, Atomic Bomb Disease Institute and ²Division of Immunology, Endocrinology and Metabolism, Department of Medical and Dental Sciences, Graduate School of Biomedical Sciences, Nagasaki University, 1-12-4 Sakamoto, Nagasaki 852-8523, Japan

(Correspondence should be addressed to Y Nagayama; Email: nagayama@nagasaki-u.ac.jp)

Abstract

Experimental Graves'-like hyperthyroidism can be induced in susceptible mouse strains by repetitive immunizations with recombinant adenovirus expressing the human full-length TSH receptor (TSHR) or its A-subunit. Previous studies have shown that splenocytes from immunized mice produce interferon (IFN)-γ and interleukin (IL) 10 in response to antigen stimulation in an in vitro T cell recall assay. Although IFN-γ is now well known to be essential for disease induction, the role(s) played by IL10 are unknown. Therefore, this study was conducted to clarify the significance of endogenous IL10 in the pathogenesis of experimental Graves' disease using IL10 deficient (IL10⁻/⁻) mice. Our results show that T cell response was augmented when estimated by their antigen-specific secretion of the key cytokine IFN-γ, but B cell function was dampened, that is, anti-TSHR antibody titers were decreased in IL10⁻/⁻ mice, resulting in a lower incidence of Graves' hyperthyroidism (54% in IL10⁺/+ vs 25% in IL10⁻/⁻). Thus, in addition to IFN-γ, these data clarified the role of IL10 for optimizing anti-TSHR antibody induction and eliciting Graves' hyperthyroidism in our Graves' mouse model.

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Introduction

Graves' disease is a thyroid-specific autoimmune disease characterized by overproduction of thyroid hormones and thyroid enlargement by agonistic anti-TSH receptor (TSHR) autoantibody. Graves'-like hyperthyroidism can be experimentally induced in susceptible mouse strains (e.g. BALB/c) by repetitive immunizations with recombinant adenovirus expressing the human full-length TSHR or its A-subunit (Nagayama et al. 2002, Chen et al. 2003). In these models, an in vitro T cell recall assay demonstrates the antigen-specific secretions of interferon (IFN)-γ and interleukin (IL) 10 from splenocytes of immunized BALB/c mice (Nagayama et al. 2004b, Saitoh et al. 2007), suggesting the importance of these two cytokines in the disease pathogenesis. Indeed our previous study clearly demonstrates the resistance of IFN-γ deficient BALB/c mice to induction of Graves’ hyperthyroidism (Nagayama et al. 2004a). This finding indicates that Graves' hyperthyroidism is an autoantibody-dependent but T helper type 1 (Th1)-mediated autoimmune disease. Our other studies, showing suppression of development of Graves' disease by Th2 polarization induced by parasite infection or IL4, support this notion (Nagayama et al. 2003, 2004b).

Regarding IL10, it was originally discovered as cytokine synthesis inhibitory factor produced by Th2 cells and is now well known to have pleotropic actions on various immune cells (Moore et al. 2001, Asadullah et al. 2003). For example, IL10 has, on the one hand, a regulatory (suppressing) effect on T cells but, on the other hand, activates B cells and induces class switching and secretion of antibody. Overall, IL10 is an immunosuppressive cytokine because IL10 deficient (IL10⁻/⁻) mice spontaneously develop autoimmune diseases (Kuhn et al. 1993). However, the effects of IL10 on autoimmune diseases so far reported vary: exogenous IL10 suppresses (Pennline et al. 1994, Bai et al. 1997, Bettelli et al. 1998, Ma et al. 1998, Batteux et al. 1999) or augments (Ishida et al. 1994, Zhang et al. 2001) and IL10 deficiency inhibits (Poussin et al. 2000, Johansson et al. 2001) or enhances (Poussin et al. 2000, Johansson et al. 2001) different autoimmune diseases. Although we have previously shown that exogenous IL10 prevents BALB/c mice from developing Graves’ disease (Saitoh et al. 2005), the consequence of genetic disruption of IL10 gene on this disease has not yet been evaluated in mice.

Therefore, this study was conducted to clarify the significance of endogenous IL10 using IL10⁻/⁻ mice. Our data demonstrate that IL10 deficiency dampens induction of Graves’ hyperthyroidism by suppressing induction of anti-TSHR antibodies. Thus, in addition to IFN-γ, IL10 is a cytokine critical for the pathogenesis of Graves’ hyperthyroidism in mice.
Materials and Methods

Mice

Wild-type (wt) IL10<sup>+/+</sup> BALB/c mice were purchased from Charles River Japan Laboratory, Inc. (Tokyo, Japan) and IL10<sup>−/−</sup> BALB/c mice from Jackson Laboratory, Inc. (Bar Harbor, ME, USA). IL10<sup>+/+</sup> and IL10<sup>−/−</sup> mice were crossed to generate IL10<sup>C</sup>/K<sup>C</sup> mice, which were then intercrossed to produce IL10<sup>C</sup>/<sup>−/−</sup> littermate mice. Both males and females were used for the current studies. All the mice were bred in the specific pathogen-free animal facility at Nagasaki University. Animal care and all experimental procedures were performed in accordance with the Guidelines for Animal Experimentation of Nagasaki University with approval of the Institutional Animal Care and Use Committee.

Experimental protocols

Construction, amplification, purification of non-replicative recombinant human adenovirus expressing the human TSHR A-subunit (Ad-TSHR289), and determination of the viral particle concentration were described previously (Nagayama et al. 2002, Chen et al. 2003).

Mice were i.m injected in the quadriceps with 100 µl PBS containing 10<sup>10</sup> particles of Ad-TSHR289 on two occasions at 3-week intervals. Blood samples and thyroid tissues were obtained 2 weeks after the second immunization.

Thyroxine and anti-TSHR antibody measurements

Serum-free thyroxine (T<sub>4</sub>) concentrations were measured with a RIA kit (DPC-free T<sub>4</sub> kit; Diagnostic Products, Los Angeles, CA, USA) and expressed as ng/dl. The normal range was defined as the mean ± 3 S.D. of control untreated mice.

Anti-TSHR antibodies in mouse sera were determined by a flow cytometric assay with Chinese hamster ovary cells stably expressing the full-length human TSHR, as described previously (Saitoh et al. 2007). This assay measures the titers of anti-TSHR antibodies recognizing the native TSHR expressed on the cell surface. The data were expressed as percentage mean fluorescent index (MFI) compared with the mean values from control untreated mice. The normal range was defined as the mean ± 3 S.D. of control mice.

Cytokine assays

Splenocytes were cultured (triplicate aliquots) at 5 × 10<sup>5</sup> cells/well in a 96-well round bottomed culture plate in the presence or absence of 10 µg/ml TSHR289 protein as described previously (Saitoh et al. 2007). The culture supernatants were collected 4 days later. The concentrations of IFN-γ and IL10 were determined with Bio-Plex Suspension Array System (Bio-Rad). Cytokine production was expressed as pg/ml using a standard curve of recombinant mouse cytokines.

Flow cytometry

Single-cell suspensions of splenocytes were prepared from spleens, and red cells were lysed in the ammonium chloride buffer. The cells were resuspended in PBS and stained with phycoerythrin or FITC-conjugated anti-CD4 (H129.19), anti-CD8 (53-6.7), anti-CD19 (1D3), anti-CD25 (7D4), anti-CD44 (IM7), and anti-CD62L (MEL-14) (PharMingen, San Diego, CA, USA or eBioscience, San Diego, CA, USA) and analyzed on a FACSCanto II flow cytometry using FACS Diva software (BD Biosciences, San Diego, CA, USA).

Thyroid histology

Thyroid histology was examined with hematoxylin and eosin (H&E) staining of formalin-fixed tissue sections.

Statistical analysis

Levels of T<sub>4</sub>, anti-TSHR antibodies and cytokines, and incidences of hyperthyroidism were analyzed by t-test or by χ<sup>2</sup> test respectively. A ‘P’ value of <0.05 was considered statistically significant.

Figure 1  (A) Serum-free T<sub>4</sub> concentrations and (B) anti-TSHR antibody titers in IL10<sup>+/+</sup> and IL10<sup>−/−</sup> BALB/c mice immunized twice with Ad-TSHR289. Free T<sub>4</sub> and anti-TSHR antibody levels were determined 2 weeks after the second immunization. Data are shown for individual mice. The horizontal broken lines indicate the normal upper limits of free T<sub>4</sub> and anti-TSHR antibodies. ND, not determined.

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Results

To clarify role(s) played by endogenous IL10 in the pathogenesis of Graves’ hyperthyroidism in a mouse model, induction of anti-TSHR immune responses and ensuing development of Graves’ hyperthyroidism was compared in the littermates of IL10+/+ and IL10−/− mice.

As previously reported (Chen et al. 2003), injection of Ad-TSHR289 induced increased T₄ in more than half (14 out of 20, 53.8%) of wt (IL10+/+) BALB/c mice (Fig. 1A). However, increased T₄ was observed only in seven out of 28 (25%) IL10−/− mice. Thus, IL10 deficiency significantly suppressed induction of hyperthyroidism (P<0.01 by χ² test). Levels of T₄ were also significantly different between the two groups (1.50±0.77 in IL10+/+ vs 0.88±0.35 ng/ml in IL10−/− mice, mean±s.d., P<0.01 by t-test).

Regarding anti-TSHR antibodies (Fig. 1B), the antibody titers were also significantly lower in IL10−/− mice than IL10+/+ mice (MFI: 765.7±774.0 in IL10+/+ vs 331.8±344.2 in IL10−/− mice, mean±s.d., P<0.05 by t-test).

A T cell recall assay, in which antigen-specific splenocyte secretion of IFN-γ in vitro was determined, was then performed as a measure of T cell activation (Ueki et al. 2011).

Discussion

Since the first description of Graves’ mouse models using recombinant adenovirus expressing the full-length human TSHR in 2002 and the receptor A-subunit in 2003 by our groups (Nagayama et al. 2002, Chen et al. 2003), we have tried to clarify the pathogenesis of this disease using various

Table 1 Comparison of lymphocyte subsets in splenocytes of immunized IL10+/+ and IL10−/− mice

<table>
<thead>
<tr>
<th>Subset</th>
<th>IL10+/+</th>
<th>IL10−/−</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD4+</td>
<td>30.7±2.0</td>
<td>21.6±4.9*</td>
</tr>
<tr>
<td>CD4+CD25+</td>
<td>6.4±0.44</td>
<td>6.0±1.45</td>
</tr>
<tr>
<td>CD4+CD44−CD62L+/CD4+ naive</td>
<td>6.1±1.2</td>
<td>4.9±0.8</td>
</tr>
<tr>
<td>CD4+CD44+CD62L+/CD4+ activated</td>
<td>56.1±5.3</td>
<td>57.1±5.9</td>
</tr>
<tr>
<td>CD4+CD44+CD62L−/CD4+ memory</td>
<td>34.8±4.6</td>
<td>37.9±3.9</td>
</tr>
<tr>
<td>CD8+</td>
<td>12.5±2.1</td>
<td>17.0±4.8</td>
</tr>
<tr>
<td>CD19+</td>
<td>38.3±5.6</td>
<td>29.6±3.8*</td>
</tr>
</tbody>
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*Significantly lower than IL10+/+ mice (P<0.05 by t-test, n=3).
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and myasthenia gravis (Zhang antibody-mediated disease including lupus (Ishida (EAE; Bettelli incidence of Graves’ hyperthyroidism, in IL10 decreased anti-TSHR antibody titers, resulting in a lower incidence of Graves’ hyperthyroidism, in IL10^{-/-} mice.

Since our recent study revealed that not only antibody-producing but also antigen-presenting abilities of B cells are crucial for development of Graves’ hyperthyroidism (Ueki et al. 2011), it is possible that B cell’s antigen-presenting ability may also be impaired in IL10^{-/-} mice. Furthermore, lower percentage of CD19^+ B cells in IL10^{-/-} mice is likely attributed to lack of positive regulation by IL10.

These results are generally consistent with previous studies showing that T cells from IL10^{-/-} mice exhibit enhanced antigen-specific response probably due to lack of the regulatory function of IL10 on T cells, but B cells from IL10^{-/-} mice show impaired function due to lack of IL10’s ability to stimulate B cells (Bettelli et al. 1998, Poussin et al. 2000).

In the previous studies, exogenous IL10 administration or transgenic/gene transfer-mediated IL10 overexpression suppresses cell-mediated autoimmune diseases such as auto-immune neuritis (Bai et al. 1997), collagen-induced arthritis (Ma et al. 1998), experimental autoimmune encephalitis (EAE; Bettelli et al. 1998), diabetes (Pennline et al. 1994), and Hashimoto’s thyroiditis (Batteux et al. 1999), but enhances antibody-mediated disease including lupus (Ishida et al. 1994) and myasthenia gravis (Zhang et al. 2001). On the other hand, IL10^{-/-} mice are more susceptible to EAE (Bettelli et al. 1998) and collagen-induced arthritis (Johansson et al. 2001), but resistant to myasthenia gravis (Poussin et al. 2000) and anti-type II collagen antibody-induced arthritis (Johansson et al. 2001). Thus IL10 appears to suppress cell-mediated autoimmune diseases but to accelerate antibody-mediated autoimmune diseases (not necessarily Th2 diseases) including Graves’ disease. In other words, the regulatory function of IL10 on T cells likely dominates over its stimulatory function on B cells in cell-mediated autoimmune diseases, while IL10’s stimulatory ability to B cells appears to be dominant in antibody-mediated autoimmune diseases.

However, a difference between our data and the others is that not only IL10 gene disruption (this study) but also gene transfer-mediated IL10 overexpression (Saitoh et al. 2005) has also suppressed Graves’ hyperthyroidism in our mouse Graves’ model. Although the reason(s) for this difference is unclear, it is not surprising that the pharmacological levels of exogenous IL10 and the physiological levels of endogenous IL10 have distinct functions, as we have previously shown for IL4 (Nagayama et al. 2003, 2004a).

Overexpression of IL10 has recently been shown to induce CD4^+ CD25^+ regulatory T cells (Goudy et al. 2003), which are well known to negatively regulate immune response (Sakaguchi 2005). Indeed our recent antibody-mediated depletion study has demonstrated a regulatory role of CD4^+ CD25^+ T cells in Graves’ hyperthyroidism (Saitoh & Nagayama 2006). Although the percentages of CD4^+ CD25^+ regulatory T cells have not been studied in our previous report (Saitoh et al. 2005) and they were the same between IL10^{-/-} and IL10^{-/-} mice in this study, the effect of IL10 deficiency on T cell’s regulatory function may be worth scrutinizing.

Overall, this study, together with our previous study (Nagayama et al. 2004a), identified IFN-γ and IL10 as critical cytokines for inducing anti-TSHR antibodies and eliciting Graves’ hyperthyroidism in our mouse model. It may be worth noting here that, clinically, the role of IL10 for the pathogenesis of Graves’ disease can also be suspected by studies on IL10 gene polymorphisms (Khalilzadeh et al. 2010) and on serum IL10 concentrations in patients with autoimmune thyroid diseases (Takeoka et al. 2004).

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Author contribution statement

Y N conceived and designed the experiments, and wrote the manuscript; and I U, N A, and K K performed the experiments and analyzed the data.

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