Calcineurin expression and activity is regulated by the intracellular redox status and under hypertension in human neutrophils

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Abstract

Calcineurin (protein phosphatase 2B) (CN) comprises a family of serine/threonine phosphatases that play a pivotal role in signal transduction cascades in a variety of cells, including neutrophils. Angiotensin II (Ang II) increases both activity and de novo synthesis of CN in human neutrophils. This study focuses on the role that intracellular redox status plays in the induction of CN activity by Ang II. Both de novo synthesis of CN and activity increase promoted by Ang II were downregulated when cells were treated with L-buthionine-(S,R)-sulfoximine, an inhibitor of synthesis of the antioxidant glutathione. We have also investigated the effect of pyrrolidine dithiocarbamate and phenazine methosulfate, which are antioxidant and oxidant compounds, respectively, and concluded that the intracellular redox status of neutrophils is highly critical for Ang II-induced increase of CN expression and activity. Results obtained in neutrophils from hypertensive patients were very similar to those obtained in these cells on treatment with Ang II. We have also addressed the possible functional implication of CN activation in the development of hypertension. Present findings indicate that downregulation of hemoxygenase-1 expression in neutrophils from hypertensive subjects is likely mediated by CN, which acts by hindering translocation to the nucleus of the transcription factor NRF2. These data support and extend our previous results and those from other authors on modulation of CN expression and activity levels by the intracellular redox status.

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Introduction

Redox regulation of cellular processes is currently a focus of intense research (Finkel 2011). The intracellular redox status is determined by the balance between reactive oxygen species (ROS) and the cellular antioxidant defenses. Although high concentrations of ROS are catatonic, they are now known to function as signal transducing molecules at lower concentrations, modulating indirectly the activity of many enzymes and transcription factors. Classical regulation of the activity of these proteins by phosphorylation is subjected to modulation by ROS. In general, ROS increase net phosphorylation by inhibiting phosphatases and only some kinases can be directly modified by them (Paravicini & T ouyz 2006).

Calcineurin (CN) is a Ca2+/calmodulin-dependent serine/threonine phosphatase. Since its first proposed biological role as a putative inhibitor of calmodulin-dependent phosphodiesterase, CN has gradually revealed itself as a modulator of the immune response. CN is the main target of the immunosuppressive drugs cyclosporine A (CsA) and tacrolimus, and it is considered nowadays as a major player in Ca2+-dependent eukaryotic signal transduction pathways (Lawrence et al. 2011). In this context, CN regulates the expression of a number of immune–response genes through dephosphorylation of a family of transcription factors known as nuclear factors of activated T cells (NF-ATs; Rusnak & Mertz 2000). CN belongs to the class of metallophosphatases containing iron and zinc in the binuclear center at their active site. During the past two decades, several studies have shed light on possible mechanisms of CN activity modulation by ROS. It has been described that superoxide is a very potent CN-inhibiting species, acting by altering the redox state of iron at the CN active site (Namgaladze et al. 2002). Although CN function is particularly important in brain, cartilage, and
lymphocytes, we have shown that neutrophils also exhibit high CN expression (Carballo et al. 1999). Neutrophils, in addition to their classically assigned role in host defense against microbial pathogens, play a key function in the genesis of inflammatory diseases through the release of ROS (Segel et al. 2011).

Hypertension is considered nowadays to be an inflammatory disease, and increasing knowledge on hypertension has led to appreciation of the relevant role in this process of circulating leukocytes in general, and of neutrophils in particular (Sela et al. 2004, Tsukimori et al. 2007). Angiotensin II (Ang II) is a potent vasoconstrictor and regulator of cellular immune responses through a CN-dependent pathway (Nataraj et al. 1999).

Previous work from our group indicates that neutrophils are highly responsive to Ang II in the context of superoxide anion production and upregulation of CN activity and synthesis (El Bekay et al. 2003). In neutrophils from healthy and hypertensive subjects, we have reported that expression of the antioxidant enzyme hemoxygenase-1 (HO1 (HMOX1)) becomes induced at the mRNA and protein levels with a parallel increase in enzyme activity on treatment with prostaglandin (Alba et al. 2008). The present work focuses on the role that an altered intracellular redox status plays in the induction of CN activity. We have also investigated expression levels of CN in normal human neutrophils under treatment with Ang II, as well as in neutrophils from hypertensive patients, and we have analyzed the possible modulating role of CN in hypertension.

Materials and Methods

Reagents

Dextran T-500 and [γ32P]ATP were obtained from Amersham Pharmacia Biotech. Ficoll-Paque was from BioWhittaker. CsA was kindly supplied by Dr S F Borel (Sandoz Ltd., Basel, Switzerland). Glutathione (GSH), Ang II, t-buthionine-(S,R)-sulfoximine (BSX), disopropyl fluorophosphate (DFP) soybean trypsin inhibitor, leupeptin, aprotinin, and goat antirabbit IgG conjugated to HRP were purchased from Sigma. The synthetic peptide used as a substrate for CN was purchased from Peninsula Laboratories. 2-Mercaptoethanol, SDS, acrylamide, N,N'-methylenebisacrylamide, and blidding polyvinylidene difluoride (PVDF) transfer membranes were purchased from Bio-Rad. Rabbit antobody CN IgG was kindly provided by C B Klee (National Institutes of Health, Bethesda, MD, USA). Antibodies to HO1 (sc-10789) and Nrf2 (sc-722) were from Santa Cruz Biotechnology (Santa Cruz, CA, USA).

Isolation and culture of human neutrophils

Human peripheral blood neutrophils were isolated from fresh heparinized blood of human donors by dextran sedimentation followed by Ficoll-Paque gradient centrifugation, and hypotonic lysis of residual erythrocytes (Carballo et al. 1999). Neutrophils were washed twice in Hank’s balanced salt solution and then suspended at a density of 10⁷ cells/ml in RPMI 1640 medium supplemented with 10% FCS plus gentamicin, penicillin, and streptomycin at 50 mg/ml each. Before all stimulations, neutrophil suspensions were pre-incubated at room temperature with 1 mM DFP (to minimize proteolysis) for 5 min (Gilbert et al. 2002). Neutrophils were incubated at 37°C for the times indicated in each experiment. Under these conditions, the cells viability ranged between 90 and 97%, as estimated by the lactic dehydrogenase release assay (Gualberto et al. 1998).

Subjects

Patients were eligible as hypertensive if they were >18 years old, suffered from moderate-to–high hypertension (systolic blood pressure, >140 mmHg; diastolic blood pressure, >90 mmHg; both measured for 24 h of Holter monitoring during unrestricted daily life), and they had not undergone any previous treatment with antihypertensive drugs or other compounds. Patients with diabetes mellitus, fasting glucose in serum >120 mg/dl, total cholesterol levels >240 mg/dl, or other pathologies were excluded. The criteria for normotensive subjects were to bear systolic and diastolic blood pressure values, <120 and <80 mmHg respectively. The general characteristics of all subjects used in this work are shown in Table 1. The Universidad de Sevilla Ethics Committee approved this study, and each subject gave informed consent before its undertaking.

CN activity assay

CN activity was measured as described previously (Carballo et al. 1999). Briefly, neutrophils (1×10⁷ cells/ml) were incubated in Krebs–Ringer–HEPES buffer (KR–HEPES) containing 118 mM NaCl, 4.75 mM KCl, 1.18 mM H2PO4, 1.18 mM MgSO4, 1.25 mM CaCl₂, 10 mM glucose, and 25 mM HEPES (pH 7.4), in the presence or absence of the drugs as indicated in the text. After incubation, the cells were

| Table 1 General characteristics of normotensive and hypertensive subjects |
|---|---|---|
| **Sex** | **Normotensives** | **Hypertensives** | **P value** |
| | Five men; five women | Five men; five women | |
| **Age (years)** | 40–50 | 40–50 | NS |
| **Weight (kg)** | 70.2 ± 3.4 | 83.6 ± 4.3 | NS |
| **Systolic blood pressure (mmHg)** | 119.3 ± 6.3 | 170.1 ± 14.0 | ≤0.001 |
| **Diastolic blood pressure (mmHg)** | 75.5 ± 4.5 | 101.4 ± 7.9 | ≤0.001 |
| **Body mass index (kg/m²)** | 24.2 ± 3.4 | 28.9 ± 2.2 | ≤0.05 |
harvested and lysed on ice in 60 μl of lysis buffer (50 mM Tris–HCl pH 7.5, 1 mM EDTA, 0.1 mM EGTA, 1% Triton X-100, 0.5 mM dithiothreitol (DTT), 50 μg/ml soybean trypsin inhibitor, 50 μg/ml phenylmethylsulphonyl fluoride, 10 μg/ml leupeptin, and 10 μg/ml aprotinin). A reaction mixture containing 2 μM 32P-labeled phosphopeptide, 500 nM okadaic acid (added to inhibit PP-1 and PP-2 type phosphatases), 150 μM trifluoperazine, and 50 μg cell lysate protein were incubated in a total volume of 60 μl of assay buffer (20 mM Tris–HCl pH 7.5, 100 mM NaCl, 6 mM MgCl2, 0.5 mM DTT, 0.1 mM CaCl2, and 0.1 mg/ml BSA) for 15 min at 30 °C. Subsequently, the reaction was stopped by the addition of 0.5 ml of 100 mM potassium buffer, pH 7-0, containing 5% trichloroacetic acid. The reaction mixture was passed through a 500 μl column of activated Dowex cation-exchange resin, and free inorganic phosphate was quantitated in the eluate by scintillation counting. Data are expressed as the number of picomoles of 32PO4 released in 15 min/mg protein.

GSH determination

Neutrophils were lysed in 315 μl of the phosphate–EDTA buffer containing 100 mM sodium phosphate (pH 8.0) and 5 mM EDTA, and 85 μl of 25% HPO3, which was used as a protein precipitant. The total homogenate was centrifuged at 4 °C at 100 000 g for 30 min to obtain the supernatant for the assay of GSH. This was performed by a modification of the method of Hissin & Hilf (1976). To 100 μl of the 100 000 g supernatant, 565 μl Tris–HCl–EDTA buffer, containing 200 mM Tris–HCl pH 8.2 and 5 mM EDTA, was added together with 35 μl of 1 mg/ml O-phthalaldehyde in 100% methanol. After thorough mixing and incubation at room temperature for 15 min, the solution was transferred to a quartz cuvette. Fluorescence at 420 nm was determined at an activation wavelength of 350 nm.

Western blotting analysis of CN, HO1, and NRF2 protein levels

Total cell lysates and nuclear extracts were prepared basically as described (Carballo et al. 1999). Western blotting analysis of HO1 (HMOX1) protein levels was performed on total cell lysates as described (Alvarez-Maqueda et al. 2004). To verify even protein loading, the blots were subsequently stripped and reprobed with polyclonal antibodies against GAPDH at a 1:1000 dilution. Blots were probed with rabbit polyclonal antibodies to HO1 at a 1:10000 dilution. NRF2 levels were analyzed on nuclear extracts by using the anti-Nf-κB IgG at a 1:5000 dilution. CN levels were measured using rabbit antibovine CN IgG at a 1:1000 dilution. Band intensities were quantitated densitometrically using the Scion Image software (Scion, Inc., Frederick, MD, USA) and corrected for differences in GAPDH levels, in the case of HO1 and CN, and for differences in nuclear proteins stained with Ponceau Red, in the case of NRF2.

Results

Effect of Ang II and intracellular GSH levels on the induction of CN activity in human neutrophils

In a previous work, we have evidenced that Ang II increases ROS production with an associated activation of MAPK. Besides, we found that Ang II increases both activity and de novo synthesis of CN in human neutrophils with a maximum stimulation being obtained at a 100 nM concentration of this hormone (El Bekay et al. 2003). Experiments in this work were thus addressed to analyze whether the increase of CN synthesis and activity promoted by Ang II in neutrophils was modulated by the intracellular redox state. Given that reduced GSH plays an important role in control of the latter, neutrophils were treated with BSX, a specific inhibitor of γ-glutamylcysteine synthase (Cuzzocrea et al. 1998), in order to depress GSH levels.

Figure 1 illustrates CN activity levels measured in neutrophils previously incubated with 10 or 20 μM BSX...
and subsequently treated with 100 nM Ang II. We previously tested that BSX effectively depressed GSH content in neutrophils (Fig. 1A). Treatment with this inhibitor resulted in a time-dependent loss of the ability of Ang II to enhance CN activity, this returning to basal levels (i.e. those found in the absence of Ang II) after 18 h of incubation in the presence of BSX (Fig. 1B and C). However, no in vitro effect of BSX alone was found on CN activity (Fig. 1D). Figure 1D illustrates that the addition of GSH to these cells was able to reverse the inhibitory effect elicited by prior BSX treatment on Ang II-mediated increase of CN activity. As shown, after 4 h of treatment with 10 mM GSH, CN activity was increased twofold by Ang II, even in the presence of BSX. By contrast, GSH addition was unable to enhance by itself Ang II-dependent CN induction.

In a previous work, we found that Ang II treatment induced the synthesis of a new CN protein (El Bekay et al. 2003). Here we have studied the effect of GSH depletion on Ang II-induced CN synthesis. With this purpose, we used a CN antibody, prepared from bovine CN, which for a long time has shown a very good reactivity to human CN (Aperia et al. 1992). By using immunological methods, we observed that the levels of both CN subunits, 19 and 59 kDa, were clearly increased 5 h after Ang II addition to neutrophils.

Figure 1 Depletion of endogenous GSH on treatment with BSX inhibits Ang II-induced CN activation in human neutrophils. Neutrophils (6×10⁶ cells/ml) were incubated either alone or in the presence of BSX at 10 μM (B) or 20 μM (A and C) for 4 (A) or 4 or 18 h (B and C) at 37 °C, as indicated. Further, the cells were incubated in the absence or presence of 100 nM Ang II for 5 h. Some cell preparations were incubated in the presence of 10 mM GSH for 2, 4, or 10 h (D). A control including 100 nM Ang II or BSX is shown. The cells were lysed and GSH levels or CN activity was assayed as described in Materials and Methods section. Results are expressed as picomoles of Pi released/min. Mean ± S.E.M. values from three separate experiments each performed in triplicate are presented. Statistical significance: * P<0.001, for BSX-treated vs BSX-untreated, ** P<0.001, for Ang II-treated vs Ang II-untreated; * P<0.01, for BSX- and Ang II-treated vs Ang II-treated and BSX-untreated. $ P<0.01, for Ang II-, BSX-, and GSH-treated vs Ang II- and BSX-treated and GSH-untreated.
It was also noticeable that Ang II-dependent de novo CN synthesis was decreased when the neutrophils were treated with 20 mM BSX for 20 h, but that the amount of CN protein was higher than that found in untreated cells. This fact was in contrast with the observation that BSX decreased CN activity down to basal levels, which could indirectly suggest a positive role of GSH on the catalytic center of CN (Fig. 1B).

In a new series of experiments, we analyzed whether environmental stress induced by other specific redox reagents, added to neutrophils a short time (ca. 1 h) before Ang II, could modify Ang II-dependent CN activation. First, we assayed the thiol-containing antioxidant, pyrrolidine dithiocarbamate (PDTC), and the effect we observed was opposite to that obtained with GSH, in the sense that CN activity decreased down to basal levels, which could indirectly suggest a positive role of GSH on the catalytic center of CN (Fig. 1B).

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**Effect of hypertension on the induction of CN activity in human neutrophils**

Hypertension is characterized by high intrarenal levels of Ang II (Navar et al. 2011). Thus, subsequent experiments using this hormone were carried out in human neutrophils from hypertensive patients compared with cells from healthy subjects. As shown in Fig. 4, neutrophils from hypertensive patients exhibited an increased basal expression of CN with respect to normotensive neutrophils. These high levels of CN decreased when neutrophils from hypertensive patients were incubated at 37 °C for 15 h in the absence, but not in the presence, of 100 nM Ang II. The results obtained in human neutrophils incubated with Ang II were thus very similar to those obtained in untreated neutrophils from hypertensive patients.

**GSH-related enzyme levels in neutrophils from hypertensive patients**

Taking into account the importance of GSH in modulation of CN activity, we studied the activity of GPx and GR, two enzymes responsible for intracellular levels of GSH in human neutrophils. The activity data obtained in hypertensive patients were normalized to those found in normotensive subjects and are shown in Fig. 5. It was observed that levels of GPx activity were ca. 22% lower in neutrophils from hypertensive patients than from healthy subjects (P < 0.001). By contrast, GR activity underwent a slight, but not significant, decrease in neutrophils from hypertensive patients compared with normotensive ones.
Implication of CN in oxidative stress in hypertension

Hypertension is also characterized by an increased oxidative stress status (Drummond et al. 2011). In this context, HO1 is an inducible antioxidant enzyme that decreases the cellular levels of heme, a pro-oxidant group, and elevates those of bilirubin, an antioxidant (Gonzales et al. 2006). So, it is currently considered as a crucial enzyme to counteract intracellular oxidative stress. We have recently described that HO1 mRNA and protein expression becomes reduced together with its enzyme activity in human normal neutrophils on their treatment with Ang II (Alba et al. 2008). In this work, we set to investigate whether the down-regulatory action of Ang II on HO1 expression was mediated by CN. With this purpose and because of the difficulty to use standard DNA transfection and micro-injection techniques on neutrophils in vitro, as they are small, short-lived, and terminally differentiated cells (Haynes & Fletcher 1990, Gaines et al. 2005), we used cyclosporine, a specific inhibitor of CN activity (O’Keefe et al. 1992, Ho et al. 1996). Cells thus were preincubated with CsA before subsequent treatment with Ang II and 15-deoxy-D12,14-prostaglandin J2 (15dPGJ2), the latter being a well-known inducer of HO1 expression in human lymphocytes (Alvarez-Maqueda et al. 2004). Figure 6A shows that Ang II prevented induction of HO1 protein expression by 15dPGJ2. However, this negative effect exerted by Ang II on HO1 expression was reversed when CN was previously inhibited with CsA.

HO1 expression becomes activated by pro-oxidant reagents via translocation to the nucleus of the transcription factor nuclear factor erythroid 2-related factor 2 (NRF2; Alam et al. 1999), which has emerged as a critical regulator of the cellular response to oxidative stress. In a previous work, we found that Ang II treatment before 15dPGJ2 addition significantly prevented nuclear accumulation of Nrf2 (Alba et al. 2008). We have now studied the possible participation of CN in Nrf2 expression. As shown in Fig. 6B, it was interesting to observe that inhibition of CN also prevented the inhibition exerted by Ang II on the increase of Nrf2 nuclear levels. These results indicated that the negative effects exerted by Ang II on HO1 expression and Nrf2 nuclear levels in human neutrophils were mediated, at least in part, through CN activation.

Finally, we compared the capacity to express HO1 by neutrophils isolated from healthy subjects and hypertensive patients. Basal levels of HO1 protein were undetectable in cells from both sources (Fig. 7). However, when neutrophils were stimulated in vitro with 15dPGJ2, synthesis of HO1 was clearly detected, although it was noteworthy that its expression levels in the presence of the prostaglandin were markedly reduced in neutrophils isolated from hypertensive patients as compared with cells from healthy subjects. By contrast, when CN was inhibited by CsA, HO1 expression was maximally induced by 15dPGJ2 in both groups.

Discussion

CN is a phosphatase implicated in signal transduction, whose function is particularly important in brain and lymphocytes and is susceptible to redox regulation. We have previously...
described that CN is also present in neutrophils (Carballo et al. 1999), and that these cells are highly responsive to Ang II, a hormone that promotes the synthesis of superoxide anions and increases CN synthesis and activity (El Bekay et al. 2003). Present studies have been addressed to analyze whether CN activation promoted by Ang II in human neutrophils was modulated by the redox intracellular status.

Given that GSH is the main intracellular thiol implicated in the redox balance, we analyzed how depletion of GSH could affect the induction of CN by Ang II. With this purpose, we pretreated human neutrophils with BSX, a specific GSH synthesis inhibitor, before Ang II addition. We observed that depletion of intracellular GSH decreased the ability of Ang II to enhance CN activity and protein levels. We also investigated whether this inactivation was reversible and found that addition of GSH to neutrophils increased their CN activity.

It has been described that CN has 1–2 thiol groups essential for its full catalytic activity (Wagner & Mutus 1991). Thus, the latter can be diminished on exclusion of reducing agents, such as GSH, and the reversal of CN inhibition exerted by BSX, after incubation with GSH, may be partially due to the ability of GSH to reduce thiols. Therefore, the intracellular levels of GSH seem to be critical for Ang II–promoted induction of CN expression and activity. Other authors have described similar increases in CN activity in fibroblasts on incubation with two other thiol-reducing agents, N-acetylcysteine and lipoic acid (Sommer et al. 2000). The reversibility of CN inhibition by GSH is especially important in the emerging field of redox signaling. In general, reversibility is a major criterion for a particular enzyme modification to be associated with a physiological regulatory mechanism. The reversible oxidative modification of proteins, like formation of cysteine sulenic acid or cyclic sulfenyl-amine (Salmeen et al. 2003), is opposed to non-specific oxidative damage to proteins, such as protein carbonyl formation or cysteine oxidation to sulfonic acid (Droge 2002). Thus, rapid recovery of CN activity after removal of an oxidative signal indicates that CN redox regulation can participate in physiological signaling.

We have also checked the effects of another antioxidant, PDTC, on CN activity. However, when this compound was added to neutrophils before Ang II addition, CN activity was drastically canceled. This compound possesses antioxidative properties by the virtue of its ability to scavenge hydroxyl radicals and superoxide anions, as well as by chelating heavy
metals that can cause the production of hydroxyl radicals by catalyzing the Haber–Weiss reaction. Our results are in agreement with those reported by Sommer et al. (2000), who found that PDTC increased the activity of purified CN but inhibited most of the CN activity present in fibroblast lysates likely by chelating an exogenous metal(s) needed for its activity. By metal abstraction, PDTC inactivates copper/zinc superoxide dismutase (Kelner et al. 1989), a stabilizing factor in intact cells, which protects CN from inactivation (Wang et al. 1996). It has also been described that PDTC prevents activation of the transcription factor NF-AT in T cells, possibly by inhibiting CN phosphatase activity (Saccani et al. 1999).

CN belongs to the class of metallophosphatases containing iron and zinc in the binuclear center at their active site. Namgaladze et al. (2002) and Ghosh et al. (2003) have demonstrated that only under certain oxidation states in the dinuclear metal center of CN this enzyme retain catalytic activity. Indeed, Fe$^{2+}$/Zn$^{2+}$ is the active form of the enzyme with very high activity, and its oxidized Fe$^{3+}$/Zn$^{2+}$ form is the inactive form of the enzyme. Although, it was shown previously by Yu et al. (1997) that Fe$^{3+}$/Zn$^{2+}$ was the active form of CN.

Another approach used in the present work to expand current knowledge on regulation of CN by Ang II was based on the hypertension model, a status characterized by high levels of Ang II (Navar et al. 2011) and increased oxidative stress (Drummond et al. 2011). Besides, it is increasingly evident that the neutrophils are involved in this disease. This led us to address CN expression in neutrophils from normal and hypertensive subjects, where we obtained similar results. Next, by taking into account the importance of GSH for CN activity, we studied the activity of GPx and GR, two enzymes that modulate the intracellular levels of GSH, in neutrophils from normotensive and hypertensive subjects. This molecule has multiple roles both as an antioxidant and as a cosubstrate for the detoxification of peroxides by GPx. As a result of the action of this enzyme, GSH is oxidized to GSSG, the accumulation of which in tissues causes a variety of noxious effects. Therefore, GSSG must be reduced back to GSH in the reaction catalyzed by GR.

We found a highly significant decrease in GPx activity in neutrophils from hypertensive subjects and a slight, yet not significant, increase in GR activity. Similar results have been obtained in other cells and in animal models of hypertension (Gomez-Amores et al. 2006, 2007, Amirkhizi et al. 2010). In the hypertensive condition, the GSH/GSSG ratio is lower than in the healthy status (Gomez-Amores et al. 2006), probably due to associated oxidative stress. An explanation of GPx reduced activity may be that reactive aldehydes interact with the selenocysteine residue of GPx via a Michael-type addition reaction, resulting in the depletion of enzyme activity (Kondo et al. 2001). Another possibility may be a compensatory response by the hypertensive subjects to ameliorate GPx activity in order to maintain GSH levels. In this sense, these levels of GSH in hypertensive subjects could be sufficient to maintain CN activity.

In the present work, we have also aimed at finding a possible functional implication for the high CN activity in hypertension. In recent years, endogenously produced CO, released from heme by the activity of hemoxygenases, has been shown to exhibit vasorelaxant properties. These are constitutive (HO2) or inducible (HO1) isoforms able to catalyze the rate-limiting step in the metabolic conversion of pro-oxidant heme to endogenous antioxidants (biliverdin and bilirubin) and CO. HO1 expression is induced by a series of stimuli that have in common their ability to elicit oxidative stress and exhibit anti-inflammatory, anti-proliferative, anti-apoptotic, and antioxidant effects on the vasculature, in addition to protecting against atherosclerosis development (Loboda et al. 2008).

Our group has previously shown that HO1 synthesis is induced by 15dPGJ2 in neutrophils, and that HO1 expression (at the mRNA and protein levels) and activity are down-regulated in both neutrophils from healthy subjects treated with Ang II and in untreated neutrophils from hypertensive patients (Alba et al. 2008). As Ang II is able to induce both CN synthesis and activity in human neutrophils, we have examined in this work the possibility that the Ang II effect observed on HO1 might be mediated by CN. We found that inhibition of CN by CsA did abolish the effect of Ang II on HO1 synthesis, which allowed us to infer that Ang II exerts its effect on HO1 expression in a CN-mediated fashion. It is noteworthy that results similar to those found in Ang II–treated neutrophils were obtained in cells from hypertensive patients. The expression of HO1 and/or the application of exogenous CO have been shown to confer protection in several models of cardiovascular injury or disease, including hypertension and atherosclerosis, and in HO1 deficiency it has been related to extensive endothelial damage (Ryter et al. 2006).

The main transcription factor responsible for HO1 synthesis is NRF2. When CN was inhibited by CsA, a decrease in Nrf2 nuclear translocation was found in this work. We can thus infer that CN prevents HO1 induction by decreasing the translocation to the nucleus of the transcription factor NRF2. The regulation of NRF2 is currently an issue of great interest. It has been proposed that the Nrf2 system may be a potential target for the development of indirect antioxidants aimed at the prevention/treatment of a wide array of human diseases (Jung & Kwak 2010). Dissociation of Nrf2 from Keap1 is the primary mechanism for Nrf2 translocation. The increase in the cellular oxidative status provokes the disruption of Nrf2–Keap1 interaction and thereby increases nuclear translocation of Nrf2. It has been demonstrated that inducers of NRF2-dependent transcription inhibit Keap1-dependent Nrf2 degradation exerted by the ubiquitin–proteasome pathway (Zhang & Hannink 2003). In mouse peritoneal macrophage cells an electrophilic agent, diethylmaleate, has been shown to facilitate nuclear accumulation of Nrf2 by suppressing...
Keap1 activity (Itoh et al. 2003). How CN may hinder Nrf2 nuclear translocation in our neutrophil preparations is unknown. It can be speculated that CN might affect the redox status of Keap 1 and thereby decrease its dissociation from Nrf2 or promote its degradation.

Finally, we have also compared the capacity to express HO1 by neutrophils isolated from healthy subjects and hypertensive patients. HO1 expression in the presence of the inducer 15dPGJ2 was markedly lower in neutrophils obtained from patients. HO1 expression in the presence of the inducer from Nrf2 or promote its degradation.

redox status of Keap 1 and thereby decrease its dissociation unknown. It can be speculated that CN might affect the nuclear translocation in our neutrophil preparations is

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Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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