The role of interleukin 1β in the anti-adipogenic action of macrophages on human preadipocytes

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Abstract

When adipose tissue accumulates in obesity, the ability of preadipocytes to differentiate permits a hyperplastic expansion of functional adipocytes that preserves insulin sensitivity. Adipose infiltration by macrophages is associated with an adipogenic deficit and the appearance of inflamed, insulin-resistant hypertrophied adipocytes. Interleukin 1β (IL1β) has been reported to account for the anti-adipogenic action of macrophages in a mouse model. Using the THP-1 human macrophage cell line and human primary preadipocytes, our objective was to determine whether IL1β was necessary for the ability of conditioned medium from THP-1 macrophages (THP-1-MacCM) to: i) stimulate human preadipocyte inhibitor of κB kinase β (IKKβ) and ii) inhibit human adipocyte differentiation. IL1β is present in THP-1-MacCM, and THP-1-MacCM or IL1β (500 pg/ml; its concentration in THP-1-MacCM) acutely stimulated IKKβ phosphorylation and inhibitor of κB (IkB) degradation in preadipocytes. IL1β was sufficient to inhibit adipogenesis on its own, and this was blocked by SC-514, an IKKβ inhibitor, as has been reported for THP-1-MacCM. IkB degradation by IL1β-immunodepleted THP-1-MacCM was attenuated, whereas IKKβ phosphorylation and the inhibition of adipocyte differentiation were unchanged. Therefore, in contrast to what has been suggested for mouse cell models, IL1β is not required for the ability of MacCM to inhibit adipogenesis in human cell models.

Key Words

- preadipocyte
- differentiation
- macrophage
- interleukin 1β
- cytokine
- inflammation

Introduction


Determining the precise identity of anti-adipogenic macrophage factors has been challenging. One candidate is interleukin 1β (IL1β), a member of the IL1 family of cytokines. It is synthesized in an inactive form, and...
becomes active as an inflammatory cytokine when processed by caspase 1 within the inflammasome, as recently reviewed (Tack et al. 2012). IL1β expression is increased in obese mouse models, and it is associated with metabolic inflammation. It is sufficient to induce insulin resistance in adipocytes, interference with its action improves insulin sensitivity, and its genetic deletion results in a more favorable metabolic phenotype. Within adipose tissue, IL1β is produced by macrophages and, to a lesser extent, by adipose cells (Tack et al. 2012).

IL1β binds to the IL1 type 1 receptor to activate pro-inflammatory intracellular signaling molecules, such as inhibitor of κB kinase β (IKKβ), that can block the adipocyte differentiation program (Gregoire et al. 1998, Tanti & Jager 2009). IKKβ is also activated in human preadipocytes by medium conditioned by human THP-1 macrophages (THP-1-MacCM), and this medium also suppresses the adipogenic response (Yarmo et al. 2010). Furthermore, IL1β was recently proposed to be the factor underlying the anti-adipogenic action of MacCM in a mouse model (Lu et al. 2010). Therefore, our objective was to determine whether IL1β was responsible for the ability of human THP-1-MacCM to: i) stimulate human preadipocyte IKKβ and ii) inhibit human adipocyte differentiation.

Materials and methods

Culture of THP-1 macrophages and preparation of THP-1-MacCM

THP-1 monocytes (ATCC, Manassas, VA, USA) were grown in RPMI-1640 medium (Life Technologies) containing 2 mM L-glutamine, 4.5 g/l HEPES, 1 mM sodium pyruvate, 0.05 mM β-mercaptoethanol, and supplemented with 10% fetal bovine serum (FBS; Life Technologies) and antibiotics (100 U/ml penicillin and 0.1 mg/ml streptomycin; Life Technologies), designated THP-1 growth medium. Conditioned medium was prepared as described previously (Constant et al. 2006). Briefly, cells were resuspended at a density of 1 × 10⁶ cells/ml in THP-1 growth medium and differentiated into macrophages with the addition of 100 nM phorbol 12-myristate 13-acetate (PMA; Sigma–Aldrich) for 24 h. The medium was replaced with serum-free THP-1 growth medium (without PMA), and conditioned by THP-1 macrophages for 24 h. The conditioned medium (THP-1-MacCM) was collected and centrifuged at 150 g for 5 min. The supernatant was collected and either used directly for preadipocyte studies or frozen in aliquots at −20 °C for future use.

The anti-adipogenic effect of THP-1-MacCM was the same in either case. This standardized approach yielded THP-1-MacCM with stable properties from batch to batch. Individual batches of THP-1-MacCM were used for each preadipocyte experiment. The medium never exposed to THP-1 cells (control medium) was similarly processed to control for medium manipulations. The heat treatment of the control medium and THP-1-MacCM consisted of boiling for 15 min, followed by centrifugation (12 000 g, 5 min) of the cooled medium. The supernatants were collected and frozen for future use.

The control medium and THP-1-MacCM were fractionated by filtration through Amicon 3 or 50 kDa molecular-weight cut-off filters (EMD Millipore, Billerica, MA, USA) according to the manufacturer’s instructions. The filtrates and retenates were collected (<3, ≥3 or <50 kDa, ≥50 kDa). Unfractionated media and pooled fractions were also prepared and used as controls.

For IL1β immunodepletion experiments, the control medium and THP-1-MacCM were incubated for 1 h at room temperature in the presence or absence of 10 μg/ml mouse IgG (Jackson ImmunoResearch Laboratories, West Grove, PA, USA) or human IL1β antibody (R&D Systems, Minneapolis, MN, USA). After incubation for 30 min at room temperature with protein G-agarose, antibody–antigen complexes were removed from the medium by centrifugation. The supernatants were used in adipogenesis and cell stimulation studies.

Isolation and culture of human abdominal subcutaneous preadipocytes

Abdominal subcutaneous adipose tissue samples were obtained from 21 weight-stable patients (14 females and seven males) undergoing elective abdominal surgery (approved by the Ottawa Hospital Research Ethics Board), aged 53 ± 10 years (mean ± S.D.), with a BMI of 30 ± 9 kg/m² (mean ± S.D.; range 22–45), and were processed as described previously (Constant et al. 2006). There was no detectable effect of BMI, gender, or age on the anti-adipogenic effect of THP-1-MacCM on preadipocytes. The types (and numbers) of surgery were: abdominal liposuction (10) for cosmetic reasons; repair of incisional hernia (2); gastric bypass (1) for weight management; hysterectomy (3) for endometrial hyperplasia, fibroids, and ovarian cancer; abdominoplasty (1) for cosmetic reasons; pelvic pouch repair (1) originally constructed for previous ulcerative colitis; ileostomy closure (1) post-resolution of previous diverticulitis; rectosigmoid resection (1) for colon cancer; and partial liver resection (1) for benign
hepatic cyst. Medications used include: statins (2); proton pump inhibitors (3); escitalopram (2); t-thyroxine replacement (1); estrogen/progesterone replacement (1); allopurinol (1); enalapril (1); hydrochlorothiazide (1); carbamazepine (1); olanzapine (1); and lithium (1).

Patients on insulin or glucocorticoid steroids were excluded. Briefly, adipose tissue was removed, dissected to remove blood vessels, and digested with collagenase CLS type 1 (600 U/g tissue; Worthington Biochemical Corporation, Lakewood, NJ, USA). Stromal preadipocytes, obtained following progressive size filtration and centrifugation, were grown in DMEM (Life Technologies) supplemented with 10% FBS, antibiotics, and 50 U/ml nystatin (EMD Millipore) (designated preadipocytes) supplemented with 10% FBS, antibiotics, and 50 U/ml nystatin (EMD Millipore) (designated preadipocyte growth medium). They were either cryopreserved until use, or directly passaged, up to a maximum of three times. Preadipocytes were seeded at a density of 3×10^4 cells/cm² and grown to confluence prior to adipocyte differentiation or acute stimulation.

### Adipocyte differentiation

Preadipocytes were placed in the control medium or THP-1-MacCM (with or without heat treatment, fractionation, or immunodepletion) supplemented with 10% FBS and the following adipogenic inducers: 5 µg/ml insulin (Roche); 0.5 µM dexamethasone (Steraloids, Newport, RI, USA); 0.25 mM isobutylmethylxanthine; and 100 µM indomethacin (both from Sigma), a peroxisome proliferator-activated receptor γ (PPARγ) agonist at this concentration (Lehmann et al. 1997), for 2 weeks. This protocol for human primary preadipocytes was based on that for Poietics Human Subcutaneous Preadipocytes, in which there were no subsequent medium changes during the duration of differentiation (Lonzagroup Ltd., Allendale, NJ, USA). Where indicated, 50 µg/ml SN-50 (481480; EMD Millipore) or water vehicle was present. For some experiments, preadipocyte growth medium was supplemented with adipogenic inducers and with indicated doses of recombinant human IL1β (#201-LB-005; R&D Systems) in the presence or absence of 100 µM SC-514 (#401485; EMD Millipore) or vehicle (0.1% DMSP), or with the indicated doses of recombinant human tumor necrosis factor α (TNFα; #210-TA-010; R&D Systems). After 2 weeks of differentiation, cultures were photographed with a digital camera (Coolpix 995; Nikon, Mississauga, ON, Canada) mounted on a microscope (Eclipse TS-100; Nikon). Cells were washed and triacylglycerol (TG) was extracted and quantified spectrophotometrically, as described previously (Constant et al. 2006). The remaining protein was solubilized in Laemmli buffer (Laemmli 1970) and processed for the determination of protein concentration by the modified Lowry method (Bio-Rad), with BSA as the standard, followed by immunoblot analysis.

### Acute cell stimulation

Preadipocytes were stimulated for 30 min with or without 500 pg/ml IL1β in preadipocyte growth medium, or with the control medium or THP-1-MacCM, immunodepleted of IL1β or not, and supplemented with 10% FBS; all media contained adipogenic inducers. Cellular protein was solubilized in Laemmli buffer (Laemmli 1970) containing 1 mM sodium orthovanadate, 50 mM sodium fluoride, 5 mM sodium pyrophosphate, and 5 mM EGTA, and processed for the determination of protein concentration and immunoblot analysis, as described.

### Quantification of macrophage-secreted factors

IL1β and platelet-derived growth factor-BB (PDGF-BB) were measured in the control medium and THP-1-MacCM using ELISA Kits, according to the manufacturer’s instructions (R&D Systems). Wnt-5A was assessed by immunoblotting the concentrated control medium and THP-1-MacCM, using an Amicon 3 kDa molecular-weight cut-off filter unit (EMD Millipore).

### Immunoblot analysis

Equal amounts of solubilized cellular protein (10–20 µg depending on the experiment), or concentrated control medium or THP-1-MacCM (for wnt-5A), were resolved by SDS–PAGE and transferred to a nitrocellulose membrane (Bio-Rad). Non-specific binding sites were blocked by 5% non-fat milk, and the membranes were probed with the following antibodies directed against: adipocyte fatty acid-binding protein (aP2; 0.1 µg/ml; R&D Systems); PPARγ (1:1000); IKKβ (1:1000); inhibitor of κB (IkB) α (1:1000); phospho-IKKα/β (Ser176/180) (all from Cell Signalling, Danvers, MA, USA); ERK1/2 (0.25 µg/ml; EMD Millipore; serves as a loading control); fatty acid synthase (FAS; 1:1000; BD Biosciences, Mississauga, ON, Canada); glyceraldehyde-3-phosphate dehydrogenase (GAPDH; 1:5000; Abcam, Cambridge, MA, USA; serves as a loading control); or wnt-5A (1 µg/ml; Santa Cruz Biotechnology). The membranes were then incubated with the appropriate HRP-conjugated secondary antibodies. Signal intensity was visualized by chemiluminescence (EMD Millipore) and quantified using the Alphalmager imaging system.
(Alpha Innotec Co., San Leandro, CA, USA). Intensity was expressed as integrated optical density units.

**Statistical analysis**

One- or two-way ANOVA was used to compare multiple means (Instat, version 3.05; GraphPad, San Diego, CA, USA and Microsoft Excel 2003). Student–Newman–Keuls (one-way ANOVA) or Tukey’s (two-way ANOVA) post hoc tests were used to determine significance between samples, with P<0.05 considered as significant.

**Results**

THP-1-MacCM was heat-treated to determine whether this would alter its ability to inhibit the differentiation of primary human preadipocytes into adipocytes. Following the heat treatment, THP-1-MacCM no longer inhibited the accumulation of TG nor the induction of PPARγ and aP2, indicating that the anti-adipogenic factor(s) is heat-sensitive (Fig. 1A, B and C). The size fractionation of THP-1-MacCM at a 3 kDa molecular-weight cut-off demonstrated no anti-adipogenic activity at all in the <3 kDa fraction (90% inhibition of TG accumulation, n = 4), and complete inhibitory activity remaining in the ≥3 kDa fraction (90±2% inhibition of TG accumulation, n = 4, P<0.01). Further fractionation suggested that there was some anti-adipogenic activity in the <50 kDa range, based on a consistent trend of lower levels of TG, PPARγ, and aP2, although this did not reach significance (Fig. 1D, E and F). Together, these data on heat sensitivity and molecular size separation are consistent with the factor being a cytokine.

We investigated whether IL1β at a concentration of 500 pg/ml could activate IKKβ (Fig. 2). This is the IL1β concentration observed in our THP-1-MacCM (Fig. 5B). This value is very stable across the batches of THP-1-MacCM that we have produced, and the same applies to TNFα levels (Molga et al. 2012). IL1β strongly stimulated IKKβ phosphorylation from undetectable basal levels and also resulted in complete IkB degradation in human pre-adipocytes. Both responses were comparable with what is observed with THP-1-MacCM stimulation (Fig. 2).

Having established that IKKβ is responsive to IL1β in the human preadipocyte model, we examined the effect of the IKKβ inhibitor SC-514 on the IL1β-mediated inhibition of adipocyte differentiation (Fig. 3). IL1β at 500 pg/ml blocked adipocyte differentiation significantly, reducing TG accumulation by 73%, FAS levels by 81%, and aP2 levels by 80%. In the presence of SC-514 (100 μM), the inhibition of these adipogenic markers was completely prevented. The effect of IL1β on PPARγ levels was a more modest 38% reduction that did not reach significance.

We conducted a dose–response study of IL1β on human adipocyte differentiation. The inhibition of TG accumulation was not evident at 10 pg/ml, but was noted at 1000 and 10 000 pg/ml (Fig. 4A). A similar effect of IL1β was seen with the inhibition of the adipogenic markers PPARγ and aP2 at 1000 and 10 000 pg/ml, but not at 10 pg/ml (Fig. 4B). These results, together with the data in Fig. 3, demonstrate that the concentration of IL1β in THP-1-MacCM is in the range that is sufficient to inhibit human adipocyte differentiation.

We immunodepleted IL1β from THP-1-MacCM to determine its importance for THP-1-MacCM signaling and for the anti-adipogenic effect. The concentration of IL1β in THP-1-MacCM is in the range of 500 pg/ml, and was reduced effectively following immunodepletion (Fig. 5A). With respect to preadipocyte signal transduction, the IL1β-depleted medium still increased IKKβ phosphorylation, but the previously observed degradation of IkB was attenuated (Fig. 5B). However, despite this alteration in the signaling response, there was no change in the anti-adipogenic action of IL1β-depleted THP-1-MacCM on human adipocyte differentiation, assessed by the accumulation of TG as well as by the levels of aP2 and PPARγ (Fig. 5D and E). The observation that the complete anti-adipogenic effect could occur without complete IkB degradation suggests that NF-κB activation may not be critical, which is consistent with studies that we performed showing that the NF-κB inhibitor SN-50 (50 μg/ml) has no effect on the anti-adipogenic action of THP-1-MacCM (89±5% inhibition of TG accumulation without SN-50 vs 91±3% inhibition with SN-50, n = 3, P = 0.48).

We investigated whether three other macrophage factors that have been implicated in the blockade of adipocyte differentiation might be present in THP-1-MacCM. PDGF and wnt-5a levels were undetectable in THP-1-MacCM (n = 3). TNFα, at 20 pg/ml, the concentration present in THP-1-MacCM, had no significant effect on adipocyte differentiation (5±4% inhibition of TG accumulation, n = 3, P = 0.35). As a positive control, TNFα was inhibitory at a higher concentration of 25 000 pg/ml (95±3% inhibition of TG accumulation, n = 3, P < 0.01).

**Discussion**

Adipose tissue levels of IL1β are elevated in obesity-associated insulin resistance, and are produced principally by macrophages and, to a lesser extent, by adipocytes.
Figure 1
Heat sensitivity and size fractionation of the anti-adipogenic activity of THP-1-MacCM. Human preadipocytes were induced to differentiate for 13–15 days in the control medium (Ctrl) or THP-1-MacCM (MacCM) heat-treated or not (A, B and C), and fractionated or not with a 50 kDa MW cut-off filter (D, E and F). (A and D) Representative photomicrographs from one patient sample are shown. (B and E) TG was extracted, quantified, and normalized to protein content. Data are mean ± S.E.M. of three (B) or five (E) separate patient samples. (C and F) Solubilized cellular proteins were immunoblotted with the indicated antibodies. ERK1/2 serves as a loading control. Immunoblots from one patient sample are shown. Densitometric data are the mean ± S.E.M. of three (C) or five (F) separate patient samples, and are normalized to the loading control (ERK1/2). Statistical significance (two-way ANOVA) is denoted as *P < 0.05 or **P < 0.01, between the indicated pairs.
Figure 2
IL1β stimulates IKKβ phosphorylation and IκB degradation in differentiating human preadipocytes. Preadipocytes were stimulated for 30 min with 500 pg/ml IL1β, control medium (Ctrl), or THP-1-MacCM (MacCM), all in the presence of adipogenic inducers. Solubilized cellular proteins were immunoblotted with the indicated antibodies. Immunoblots from one patient sample are shown. Densitometric data are the mean ± S.E.M. of three separate patient samples, and are normalized to the loading control (IKKβ). Statistical significance (one-way ANOVA) is denoted as *P<0.05 or ***P<0.001, compared with the other conditions.

Figure 3
Anti-adipogenic effect of IL1β in human preadipocytes is reduced by SC-514. Human preadipocytes were induced to differentiate for 13–15 days with 500 pg/ml human recombinant IL1β, in the presence or absence of 100 μM SC-514 or vehicle (DMSO). (A) Representative photomicrographs from one patient sample are shown. (B) TG was extracted, quantified, and normalized to protein content. Data are the mean ± S.E.M. of three separate patient samples. (C) Solubilized cellular proteins were immunoblotted with the indicated antibodies. ERK1/2 serves as a loading control. Immunoblots from one patient sample are shown. Densitometric data are the mean ± S.E.M. of three separate patient samples, and are normalized to the loading control (ERK1/2). Statistical significance (two-way ANOVA) is denoted as *P<0.05 between the indicated pairs.
Interleukin 1β, macrophages, and adipogenesis

There has actually been very little investigation on the effects of IL1β on human preadipocytes. One study has reported an inhibition of the differentiation of subcutaneous abdominal preadipocytes (Simons et al. 2005), but used only one concentration of IL1β that was much higher at 10 000 vs the 500 pg/ml range in our study. Another group, studying orbital human preadipocytes, also used only one dose of IL1β at 10 000 pg/ml to demonstrate its anti-adipogenic effect (Yoon et al. 2011). Measurements of IL1β in human adipose tissue in healthy lean individuals by microdialysis are in the 70 pg/ml range (Clausen et al. 2009), and 30 pg/ml in healthy obese women (Siklova-Vitkova et al. 2009). It is possible that higher concentrations within the tissue are found in the setting of obesity-associated insulin resistance with the accompanying influx of pro-inflammatory macrophages (Fain 2010), but microdialysis data in this setting have not been reported.

To determine the importance of IL1β in THP-1-MacCM, we immunodepleted the cytokine and studied the resulting MacCM to determine the consequences on preadipocyte inflammatory signaling and the adipogenic response. IL1β immunodepletion reduced the IkB degradation response, although there was no significant difference in the extent of IKKβ phosphorylation. This result suggests that IL1β, in particular, contributes to the complete IkB degradation in human preadipocytes induced by THP-1-MacCM, whereas residual factors in THP-1-MacCM remain that are capable of elevating IKKβ phosphorylation.

However, despite this altered signaling response to IL1β-depleted THP-1-MacCM, the anti-adipogenic effect of this medium was not affected. A limitation in drawing this conclusion is the low number of adipose tissue samples obtained from a somewhat heterogeneous patient population, and therefore the possibility that an alteration in the differentiation response was missed. Yet, our data (levels of TG, PPARγ, and aP2) show no trend in that direction, and a power analysis indicates that a change of 25% in the extent of inhibition would have been detected at an acceptable type II error of 0.8.

The weaker extent of IkB degradation suggests that NF-κB activation is not required for this medium in order to inhibit adipocyte differentiation. Our studies showing no effect of SN-50 were consistent with this. BY contrast, IKKβ phosphorylation is critical, since the IKKβ inhibitor SC-514 abrogated the anti-adipogenic action of THP-1-MacCM (Yarmo et al. 2010). It is known that IKKβ phosphorylation does not always lead to IkB degradation and NF-κB activation, and that other

of TG accumulation, as well as the induction of PPARγ, aP2, and FAS. We used human IL1β for our studies, but Lu et al. (2010) did not provide information on the type of IL1β that they used, so it is difficult to comment on whether species specificity is an issue. It may be that differences between mouse 3T3-L1 vs primary human stromal preadipocytes contribute to these contrasting results.

Figure 4
IL1β inhibits human adipogenesis in a dose-dependent manner. Human preadipocytes were induced to differentiate for 13–15 days in the absence or presence of human recombinant IL1β at the indicated doses. (A) TG was extracted, quantified, and normalized to protein content. Data are the mean ± S.E.M. of three separate patient samples. (B) Solubilized cellular proteins were immunoblotted with the indicated antibodies. GAPDH serves as a loading control. Immunoblots from one patient sample are shown. Densitometric data are the mean ± S.E.M. of three separate patient samples, and are normalized to the loading control (GAPDH). Statistical significance (two-way ANOVA) is denoted as **P<0.01 between the indicated pairs.
Figure 5
IL1β is not required for the stimulation of IKKβ phosphorylation and the anti-adipogenic effect of THP-1-MacCM. (A) IL1β protein in the control medium (Ctrl) or THP-1-MacCM (MacCM), immunodepleted with IL1β antibodies (Ab), mouse IgG, or no Ab. Data are the mean ± S.E.M. of four separate experiments. Human preadipocytes were treated for 30 min (B) or 14 days (C, D and E) with the control medium (Ctrl) or THP-1-MacCM (MacCM) immunodepleted with or without the IL1β Ab or mouse IgG, all supplemented with adipogenic inducers. Solubilized cellular proteins were immunoblotted with the indicated antibodies. Immunoblots from one patient sample are shown (B and E), with IKKβ and ERK1/2 serving as loading controls, respectively. Densitometric data are the mean ± S.E.M. of three separate patient samples, and normalized to the loading control (IKKβ for (B) and ERK1/2 for (E)). Representative photomicrographs from one patient sample are shown (C). TG was extracted, quantified, and normalized to protein content. Data are the mean ± S.E.M. of three separate patient samples (D). Statistical significance (two-way ANOVA) is denoted as *P < 0.05 or **P < 0.01, between the indicated pairs.
established IKKβ cellular targets could potentially mediate the anti-adipogenic action, including IRS-1, FOXO3, and 14-3-3b (Chariot 2009). Another group has reported the same response pattern of IKKβ phosphorylation without IκB degradation in human bronchial epithelial cells stimulated with interferon-γ (Tudhope et al. 2007). We have also described this pattern in PDGF-stimulated human preadipocytes (Gagnon et al. 2009). However, our analysis of THP-1-MacCM revealed that there was no detectable PDGF.

TNFα has been investigated as an anti-adipogenic macrophage-secreted factor for human adipocyte differentiation. However, in one study, immunoneutralization of TNFα barely altered the anti-adipogenic effect, although it did result in a major reduction in preadipocyte inflammatory gene expression (Lacasa et al. 2007). Furthermore, the amount of TNFα was not actually quantified in their conditioned medium. At 20 pg/ml, the concentration measured in THP-1-MacCM (Molga et al. 2012), TNFα had no effect on human adipocyte differentiation. As a positive control, TNFα at 25 000 pg/ml was anti-adipogenic.

More recently, wnt-5a was demonstrated to be expressed by human blood monocytes and adipose tissue macrophages (Bilkovski et al. 2011). The evidence of wnt-5a secretion and its importance for inhibiting adipocyte differentiation was based on mouse J774A.1 macrophage cultures and mouse 3T3-L1 cells. It is not known whether wnt-5a would have a similar role in human adipocyte differentiation. We were not able to detect wnt-5a in our human THP-1-MacCM.

Investigations concerning the anti-adipogenic effect of MacCM have used multiple macrophage and preadipocyte cell culture models. The data accumulated are now sufficient to reveal that different anti-adipogenic factors may operate in the various systems. For example, using mouse J774A.1 macrophages and 3T3-L1 cells, an anti-adipogenic factor was found to be heat stable and <3 kDa (Ide et al. 2011), vs the larger, heat-sensitive factor noted here using human THP-1 macrophages and primary human preadipocytes. Future studies should carefully specify the experimental models used as this important question is pursued.

In the in vivo setting, it may be expected that anti-adipogenic effects of macrophages would constrain adipose tissue growth and favor hypertrophy, and thus lead to inflamed, insulin-resistant adipocytes and associated metabolic complications. There will also be additional complexity in that adipose macrophages may exist in a variety of polarized states, including M1 pro-inflammatory, M2 anti-inflammatory, and mixed phenotypes, and they may interact with preadipocytes by direct contact as well as through paracrine signals.

In summary, our results demonstrate that IL1β is able to inhibit human adipocyte differentiation, but at concentrations much higher than reported for 3T3-L1 adipocyte differentiation. However, our immunodepletion strategy shows that it is not a critical component for the anti-adipogenic activity of THP-1-MacCM. Identifying key anti-adipogenic factors in the future will allow them to be evaluated as potential targets to improve adipose tissue remodeling and function.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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