Pressor and renal regional hemodynamic effects of urotensin II in neonatal pigs

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Abstract

Renal expression of the peptide hormone urotensin II (UII) and its receptor (UTR) are dependent on kidney maturation and anatomical regions. However, renal regional hemodynamic effects of UII in neonates are unclear. Here, we investigated regional hemodynamic responses to acute intrarenal arterial administration of UII in newborn pigs. Western immunoblotting and immunofluorescence confirmed UTR expression and membrane localization in newborn pig renal afferent arterioles and afferent arteriolar smooth muscle cells respectively. Intrarenal arterial bolus injections of human UII (hUII; 1–100 ng/kg) resulted in a dose-dependent decrease in total renal blood flow (RBF) and an increase in mean arterial pressure (MAP) and renal vascular resistance (RVR) in newborn pigs. Moreover, hUII dose dependently reduced cortical blood flow (CBF) but increased medullary blood flow (MBF) in the piglets. hUII-induced MAP elevation and hemodynamic changes were inhibited by urantide, a UTR antagonist, but not losartan, a type 1 angiotensin II receptor antagonist. U-73122, a phospholipase C (PLC) inhibitor, and 2-aminoethoxydiphenyl borate, an inositol 1,4,5 trisphosphate (IP$_3$) receptor antagonist, attenuated hUII-induced MAP and RVR elevations, RBF and CBF reductions, but not MBF increase. These findings indicate that intrarenal arterial administration of hUII elevates blood pressure and induces region-selective renal hemodynamic changes in newborn pigs. Our data also suggest that the PLC/IP$_3$ signaling pathway contributes to hUII-induced alterations in MAP, RBF, RVR, and CBF but not MBF in newborn pigs.

Key Words
- Urotensin II
- neonates
- renal hemodynamics
- mean arterial pressure
- renal arterioles

Introduction

The kidney is a major source of vasoactive mediators, including angiotensin II (AngII), prostaglandins (PGs), endothelin-1 (ET-1), bradykinin, thromboxane, and urotensin II (UII). These compounds can act in an endocrine, autocrine, or paracrine fashion to modulate cardiovascular and renal homeostasis. Thus, alterations in expression, function, and regulation of these mediators and their receptors underlie pathophysiological mechanisms of a variety of cardiovascular and renal diseases (Douglas et al. 2004, Ponnuchamy & Khalil 2009).

UII, a cyclic peptide hormone, has been described as the most potent vasoconstrictor, constricting human coronary, mammary and radial arteries, rat aorta and pulmonary arteries, pig and monkey coronary arteries, and monkey basilar, renal, and mesenteric arteries with potency up to 109-fold more than ET-1 (Ames et al. 1999, 2001).
Douglas et al. 2000, MacLean et al. 2000, Maguire et al. 2000). However, other studies have shown that UII can also cause vasodilation. For example, UII-induced endothelium-dependent vasodilation in rat-isolated coronary and mesenteric arteries precontracted with 5-HT and methoxamine respectively (Bottrill et al. 2000). UII also dilated ET-1-precontracted human pulmonary and abdominal resistance-sized arteries and phenylephrine-precontracted rat renal arteries (Stirrat et al. 2001, Zhang et al. 2003). These studies suggest that UII-induced regulation of vascular tone may depend on species and blood vessel anatomical origin or size (Douglas et al. 2000, Ashton 2006, Zoccali & Mallamaci 2008).

Alterations in UII and UII receptor (UTR) expression and plasma and urinary UII concentrations have been associated with renal dysfunctions, including hypertensive renal disease, diabetic nephropathy, glomerulonephritis, and renal tubular diseases (Matsushita et al. 2001, Douglas et al. 2004, Langham et al. 2004, Ashton 2006, Balat et al. 2007, Ross et al. 2010). However, the physiological role of UII in the kidney remains controversial with studies reporting attenuation and elevation of important renal parameters, including total renal blood flow (RBF), glomerular filtration rate (GFR), urine flow rate (UV), and urinary sodium (UnaV) and potassium (UkV) excretion (Zhang et al. 2003, Ovcharenko et al. 2006, Song et al. 2006, Abdel-Razik et al. 2008a,b, Shi et al. 2008).

A recent study demonstrated that UII plasma level was increased after birth in lambs (Simpson et al. 2010). An examination of the ontogenetic pattern of UII and UTR expression in rat kidney has also revealed regional variability in UII and UTR expression from embryonic day 19 to 4 weeks after birth (Forty & Ashton 2012). However, unlike in adult, rat UII (rUII) did not alter renal hemodynamics in young (4-week-old) rats (Forty & Ashton 2012).

The physiological characteristics of newborn renal circulation differ significantly from those of young and old adults (Hook & Bailie 1979, Toth-Heyn et al. 2000). Newborns are unable to concentrate urine and exhibit higher renal vascular resistance (RVR) and lower RBF and GFR compared with adults (Hook & Bailie 1979, Toth-Heyn et al. 2000). Accumulating evidence also suggests that alterations in the physiological functions of endogenous vasoactive mediators contribute to maturation-dependent changes in renal hemodynamics (Hook & Bailie 1979, Toth-Heyn et al. 2000). Here, we examined two important, yet largely unexplored, renal actions of UII. First, we studied the effects of UII on neonatal renal circulation. Second, we tested the hypothesis that UII differentially regulate neonatal renal perfusion.

### Materials and methods

#### Animals

All animal protocols were reviewed and approved by the Animal Care and Use Committee of the University of Tennessee Health Science Center (UTHSC). Newborn pigs (1–3 day old; 1.5–2 kg) were purchased from Nichols Hog Farm (Olive Branch, MS, USA) and maintained at the UTHSC Comparative Medicine Department animal core facility. Animals were used within the first week of life.

#### Renal vascular smooth muscle cell isolation

Afferent arterioles were manually dissected from decapsulated newborn pig kidneys using a Zeiss SteREO Discovery.V12 stereomicroscope (Carl Zeiss, Thornwood, NY, USA). Smooth muscle cells were dissociated from the microvessels using a HEPES-buffered isolation solution containing (in mM) 55 NaCl, 80 sodium glutamate, 5.6 KCl, 2 MgCl$_2$, 10 HEPES, and 10 glucose (pH 7.3). Afferent arterioles were placed into isolation solution containing 1 mg/ml papain, 500 µg/ml elastase, 1 mg/ml dithioerythritol, and 1 mg/ml BSA for ~45 min at 37 °C. Arterioles were then washed and incubated in isolation solution containing 1 mg/ml collagenase F and H (2:1), 100 µM CaCl$_2$ and 1 mg/ml BSA for ~20 min at 37 °C. Digested vessels were subsequently washed in isolation solution and triturated using a fire-polished glass Pasteur pipette to yield single smooth muscle cells.

#### Immunofluorescence

Renal vascular smooth muscle cells were allowed to adhere to collagen-coated coverslips. The cells were then fixed with 4% paraformaldehyde in PBS for ~15 min and permeabilized with 0.2% Triton X-100 for ~20 min at room temperature. Following a 1 h incubation in PBS containing 5% BSA to block nonspecific immunoreactive sites, cells were treated overnight at 4 °C with rabbit polyclonal anti-UTR (Alomone Lab, Jerusalem, Israel) or normal rabbit IgG (GenScript, Piscataway, NJ, USA). After a wash, cells were incubated for 1 h at room temperature with Alexa 555-conjugated Donkey Anti-Rabbit (Life Technologies). Following wash and mount, fluorescence images were acquired using a Zeiss LSM Pascal laser-scanning confocal microscope.

#### Western immunoblotting

Afferent arterioles were homogenized in ice-cold RIPA buffer (Thermo Scientific, Rockford, IL, USA) followed by
centrifugation to extract protein. Protein lysates were mixed with SDS sample buffer containing 5% β-mercaptoethanol and boiled at ~100 °C for 5 min. Protein samples were thereafter resolved on a 7% NuPAGE Tris-Acetate Gel (Life Technologies) and then transferred to polyvinylidene difluoride membranes using a Pierce Fast Semi-Dry Blotter (Thermo Scientific). Nonspecific binding sites on the membranes were blocked by Tris buffered saline supplemented with 0.1% Tween 20 (TBS-T) and 5% nonfat milk (Research Products International Corp., Mount Prospect, IL, USA) for 1 h at room temperature. The membranes were probed with rabbit polyclonal anti-UTR (GPR14; Alpha Diagnostic International, Inc., San Antonio, TX, USA) overnight at 4 °C. After washing with TBS-T, the membranes were incubated in a HRP-conjugated secondary antibody for ~1 h at room temperature. Membranes were then processed with a Pierce Chemiluminescence kit (Thermo Scientific) and immunoreactive proteins were visualized on a Kodak Imaging system (Carestream Molecular Imaging, Rochester, NY, USA).

Renal hemodynamic measurements

Newborn pigs were anesthetized with ketamine hydrochloride (20 mg/kg) and xylazine (2.2 mg/kg) intramuscularly and maintained on α-chloralose (50 mg/kg, i.v.) at 37 °C. The animals were intubated via tracheostomy and mechanically ventilated. Animals were continually monitored during experiments for anesthesia depth and re-dosed if necessary. The right femoral artery was catheterized for continuous measurement of mean arterial pressure (MAP) using a MLT1199 physiological pressure transducer (ADInstrument, Colorado Spring, CO, USA). Another catheter was inserted in the left femoral artery and advanced through the abdominal aorta until its tip was positioned at the junction of the aorta and left renal artery for intrarenal administration of drugs. The position of the catheter tip was confirmed at the end of each experiment. A femoral vein was also catheterized for anesthetic and fluid administration. Arterial blood partial pressures of CO₂ and O₂, pH, and hematocrit were measured periodically with a Blood Gas Analyzer (Instrumentation Laboratory, Lexington, MA, USA). Ventilation was adjusted to maintain PCO₂, PO₂, and pH at physiological ~30 mmHg, >85 mmHg, and 7.4 respectively.

The left kidneys of newborn pigs were exposed retroperitoneally through flank incisions to allow clear access to the renal pedicles. To determine total RBF, a left renal artery was carefully separated from the vein and connective tissues. A flow probe (Transonic Systems, Inc., Ithaca, NY, USA) was then placed around the main renal artery and connected to a T206 dual channel small animal flow meter (Transonic Systems). Regional RBF in the pigs was measured using laser Doppler flowmetry (LDF). A laser Doppler probe (PF 407) connected to a holder (Perimed, Jarfalla, Sweden) was placed on the kidney surface to measure cortical blood flow (CBF). Medullary blood flow (MBF) was measured using a needle probe (Perimed) inserted into the kidney to a depth of ~8 mm. The laser Doppler probes were connected to a flow meter (Periflux 4001, Perimed). After each experiment, the position of the intrarenal probe tip was confirmed by dissecting the kidneys. CBF and MBF measurements were obtained as laser Doppler perfusion unit (PU). Laser Doppler probes were calibrated with a standard calibration device using a motility standard (Perimed) such that 1 PU corresponds to an analog output of 10 mV. Data were acquired and analyzed using a PowerLab data acquisition system and LabChart software (ADInstrument).

Figure 1
Newborn pig renal afferent arterioles and arteriolar smooth muscle cells express UTR protein. (A) Western immunoblotting detected unglycosylated (~40 kDa) and glycosylated (~60 kDa) forms of UTR in newborn pig renal afferent arterioles. (B) Immunofluorescence staining demonstrating plasma membrane localization of UTR in smooth muscle cells isolated from newborn pig renal afferent arterioles. Negative controls prepared using normal rabbit (Rb) IgG did not show fluorescence. Bar = 10 μm.
Chemicals

Unless otherwise stated, all reagents were purchased from Sigma Chemical. Human UII (hUII), urantide, U-73122, 2-APB, and ketamine/xylazine were purchased from California Peptide, Inc. (Napa, CA, USA), Peptide International (Louisville, KY, USA), Santa Cruz (Santa Cruz Biotechnology, Inc.), Cayman Chemicals (Ann Arbor, MI, USA), and Butler Schein Animal Health Supply (North Dublin, OH, USA) respectively.

Statistical analysis

All data are expressed as mean ± S.E.M. Statistical significance was determined using Student’s t-tests for paired or unpaired data and ANOVA with Student–Newman–Keuls for multiple comparisons tests. A P value <0.05 was considered significant.

Results

Newborn pig renal afferent arterioles and arteriolar smooth muscle cells express UTR protein

Western blot analysis of neonatal afferent arterioles using a polyclonal rabbit anti-UTR antibody detected 40 and 60 kDa immunoreactive bands corresponding to the approximate molecular weights of unglycosylated and glycosylated UTR respectively (Fig. 1A). Immunofluorescence staining of smooth muscle cells isolated from neonatal pig afferent arterioles also revealed membrane localization of UTR in cells probed with rabbit anti-UTR antibody but not normal rabbit IgG (Fig. 1B). These data confirm that UTR protein is expressed in neonatal renal afferent arterioles and localized in the plasma membrane of afferent arteriolar smooth muscle cells.

hUII elevates MAP and RVR but attenuates RBF in newborn pigs

Basal MAP, RBF, and RVR in anesthetized newborn pigs are shown in Table 1. Cumulative intrarenal arterial bolus inhibited these effects. hUII and urantide were co-administered to the piglets 10 min after intrarenal arterial bolus injection of urantide. *P<0.05 when compared with basal values; *P<0.05 when compared with hUII-induced responses.

Table 1 Basal values of MAP and renal hemodynamic parameters measured in newborn pigs

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Mean ± S.E.M.</th>
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<tbody>
<tr>
<td>MAP (mmHg)</td>
<td>80.2 ± 2.5</td>
<td>26</td>
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<tr>
<td>RBF (ml/min)</td>
<td>12.0 ± 0.8</td>
<td>22</td>
</tr>
<tr>
<td>RVR (mmHg/ml per min)</td>
<td>7.1 ± 0.4</td>
<td>22</td>
</tr>
<tr>
<td>CBF (LDF PU)</td>
<td>*312.8 ± 12.2</td>
<td>26</td>
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<tr>
<td>MBF (LDF PU)</td>
<td>140.4 ± 10.6</td>
<td>26</td>
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*P<0.05, compared with MBF.

Figure 2

hUII elevates MAP and RVR but attenuates RBF in newborn pigs. (A and B) Exemplar tracings illustrating the effects of hUII on newborn pig MAP and RBF respectively. (C, D and E) Mean (± S.E.M.) data showing that hUII dose dependently increased MAP (n=6), attenuated RBF (n=4), and elevated RVR (n=4) in newborn pigs and that urantide (20 μg/kg)
injection of hUII dose dependently increased MAP in newborn pigs (Fig. 2A and C). To determine the effect of hUII on newborn pig RVR, we first measured RBF. As illustrated in Fig. 2B and D, hUII dose dependently reduced RBF in newborn pigs. RVR, calculated as a ratio of MAP:RBF was increased by hUII in the piglets (Fig. 2E). These findings indicate that intrarenal arterial administration of hUII elevates MAP and RVR and reduces RBF in newborn pigs.

hUII reduces cortical but increases medullary perfusion in newborn pigs

To examine whether there is a regional variation in hUII-induced renal hemodynamic changes in newborn pigs, we measured blood perfusion in cortical and medullary regions of newborn pig kidneys. Basal CBF was ~55% more than MBF in newborn pigs (Table 1). hUII dose dependently decreased CBF in newborn pigs (Fig. 3A and C). In contrast, hUII elevated MBF in the pigs (Fig. 3B and D). These data indicate that hUII induces region-selective regulation of renal perfusion in newborn pigs.

UTR mediates renal hemodynamic effects of hUII in newborn pigs

To test the hypothesis that UTR mediates renal hemodynamic effects of hUII in newborn pigs, we studied the effect of urantide, a selective UTR antagonist on hUII-induced renal hemodynamic changes in piglets. Urantide did not alter RBF, RVR, CBF, or MBF in newborn pigs (Table 2). However, urantide significantly reduced MAP by ~4 mmHg in the piglets (Table 2). Furthermore, urantide attenuated hUII-induced pressor effect and renal hemodynamic changes in newborn pigs (Figs 2 and 3). These findings suggest that inhibition of UTR reduces MAP in newborn pigs. Our data also indicate that UTR mediates hUII-induced pressor effect and renal hemodynamic changes in the piglets.

Type 1 AngII receptors are not involved in hUII-induced pressor effect and renal hemodynamic changes in newborn pigs

Next, we investigated whether AngII system contributes to renal hemodynamic responses of newborn pigs to hUII. Losartan, a selective type 1 AngII (AT1) receptor antagonist, did not alter basal RBF, RVR, CBF, or MBF but significantly reduced MAP by ~6 mmHg in newborn pigs (Table 2). Losartan essentially abolished AngII-induced pressor effect and renal hemodynamic changes in newborn pigs (Table 3). In contrast, losartan did not alter pressor effect and renal hemodynamic responses to hUII in the piglets (Fig. 4A, B, C, D and E). Collectively, our data indicate that hUII-induced pressor effect and renal hemodynamic changes in newborn pigs are independent of AT1 receptors.

Activation of the phospholipase C/inositol 1,4,5 trisphosphate pathway contributes to hUII-induced pressor effect and renal hemodynamic changes in newborn pigs

To test the hypothesis that phospholipase C (PLC)/inositol 1,4,5 trisphosphate (IP3)-mediated renal vasoconstriction contributes to hUII-induced pressor effect and renal hemodynamic changes in newborn pigs, we studied the effect of U-73122, a PLC inhibitor and 2-APB, an IP3 receptor (IP3R) antagonist on hUII-induced MAP elevation.
UII activity is mediated by UTR, a G protein-coupled receptor (GPCR; Ames et al. 1999, Liu et al. 1999). Both unglycosylated (~42 kDa) and glycosylated (~60 kDa) forms of UTR protein have been identified in human adrenocortical tissues and COS-7 cells overexpressing UTR (Boucard et al. 2003, Giuliani et al. 2009). Ventricular samples from rats express the unglycosylated form of UTR, whereas the glycosylated form was the only immunoreactive UTR protein identified in rat kidneys, coronary arteries, and cultured coronary artery smooth muscle cells (Gong et al. 2004, Abdel-Razik et al. 2008b, Dominguez-Rodriguez et al. 2012). In this study, we found that newborn pig renal afferent arterioles express both unglycosylated and glycosylated forms of UTR. Thus, the expression pattern of UTR protein isoforms may vary between species and tissues. Our findings also confirm that similar to previous reports in adult rats (Ashton 2006, Song et al. 2006), UTR are expressed in neonatal pig renal vessels.

Previous studies examining renal actions of UII in mature animals have yielded inconsistent findings. For example, intrarenal arterial infusion of hUII did not alter MAP but increased RBF, GFR, UV, and UNaV in rats (Zhang et al. 2006). In contrast, i.v. injection of hUII reduced blood pressure, RBF, GFR, UV, UNaV, and UNV in rats (Song et al. 2006). I.v. injections of hUII also decreased blood pressure, RVR, and GFR but did not alter RBF and UNaV in rats (Ovcharenko et al. 2006). These conflicting reports have been attributed to several factors, including route of administration, dosage, and source of UII (Ashton 2006, 2008).
Zoccali & Mallamaci 2008, Ross et al. 2010, Tsoukas et al. 2011). However, the cyclic hexapeptide sequence in uUI carboxyl-terminus, which is responsible for its biological activity, is highly conserved between species (Itoh et al. 1987, Conlon et al. 1990, Coulouarn et al. 1998, Douglas et al. 2004). Hence, it is unlikely that the physiological function of UUI is dependent on its source (Douglas et al. 2000).

Data presented here indicate that intrarenal arterial administration of hUII elevates RVR and MAP in newborn pigs. However, a previous study showed that intrarenal arterial infusion of hUII did not alter MAP in adult rats (Zhang et al. 2003). These conflicting findings could be due to species differences and/or animal age. Urantide induced a small but significant decrease in MAP in newborn pigs, suggesting that endogenous UII regulates blood pressure in the piglets. However, urantide did not alter basal RBF, RVR, and regional perfusion in the piglets. These results raise the possibility that the hypotensive effect of urantide may be due to its systemic but not localized renal vasoregulatory actions. Neurons of the brain and spinal cord express UII (Coulouarn et al. 1998, Douglas et al. 2004, Ross et al. 2010). Studies have also shown that UII can modulate cardiovascular homeostasis via central neural mechanisms (Gibson et al. 1986, Lu et al. 2002, Lin et al. 2003). However, the expression and function of UII system in renal plexus have not been characterized. Hence, future studies are needed to explore whether hUII alters neonatal blood pressure and renal hemodynamics via regulation of renal sympathetic nerve activity.

Renal perfusion exhibits considerable regional heterogeneity. Changes in renal CBF are not always replicated in the medullary or papillary regions because medullary hemodynamics can be regulated independently of renal CBF (Pallone et al. 1990, Navar et al. 1996, Mattson 2003). Vasoactive agents, including AngII, dopamine, arginine vasopressin, atrial natriuretic peptide, and bradykinin have been demonstrated to induce renal region-specific hemodynamic changes (Takezawa et al. 1987, Pallone et al. 1990, Nobes et al. 1991, Mattson & Cowley 1993, Heyman et al. 1995, Badzynska et al. 2002, Igbe et al. 2012). However, the effect of UII on regional RBF was unclear. In this study, we show that hUII-induced activation of UTR dose dependently reduced CBF but elevated MBF in newborn pigs, suggesting that UII differentially regulates neonatal renal perfusion. Although hUII-induced elevation in MBF in the pigs occurred at the highest dose tested, it is likely that the medullary actions of UII result in localized vasodilation. Endogenous mediators of renal vasodilation, including nitric oxide (NO) and vasodilatory PGs, exert larger vasoregulatory effects.
effects in renal medulla than the cortex (Navar et al. 1996). UII stimulates NO, prostacyclin, and PGE2 release in rat coronary arteries (Ishihata et al. 2005). UII also elevated NO production in rat renal artery endothelium (Zhang et al. 2009). Moreover, the physiological activity of NO and vasodilatory PGs are elevated in fetal and newborn kidneys compared with adults (Hook & Bailie 1979, Toth-Heyn et al. 2000). Conceivably, hUII elevates MBF in newborn pigs via localized generation of NO and/or vasodilatory PGs in the medulla. However, these possibilities require further investigations.

Studies have suggested that a functional cross talk exists between UII and AngII systems. UII-induced elevation in myocardial distensibility and negative ionotropic and lusitropic effects in rabbits were inhibited by losartan (Fontes-Sousa et al. 2009). Furthermore, AngII-induced vasoconstriction was elevated in hUII-pretreated rat aortic segments (Wang et al. 2007). Data here show that at the dose that essentially abolished AngII-induced hypertension and renal hemodynamic changes, losartan did not alter pressor and renal hemodynamic responses of newborn pigs to hUII, indicating that renal actions of UII in newborn pigs are independent of AT1 receptors.

In vascular smooth muscle cells, activation of PLC-coupled GPCRs by a wide variety of vasoconstrictors stimulates phosphoinositide hydrolysis, resulting in generation of IP3 (Berridge 1993). IP3 binds to sarcoplasmic reticulum (SR)-localized IP3Rs leading to Ca2+ release from SR Ca2+ store, an elevation in intracellular Ca2+, and vasoconstriction (Berridge 1993, Sanders 2001). In cerebral and mesenteric arteries, IP3-mediated vasoconstriction can also occur independently of SR Ca2+ release and due to a molecular and functional coupling between smooth muscle cell type 1 IP3R (IP3R1) and canonical transient receptor potential 3 (TRPC3) channels (Xie et al. 2008, Adebiyi et al. 2010, 2012). However, it is unclear whether a coupling exists between IP3R1 and TRPC3 channels in renal vascular smooth muscle cells. hUII constricts rabbit aortic smooth muscle by stimulating PLC-dependent elevation in inositol phosphates, suggesting that PLC/IP3-mediated intracellular Ca2+ elevation contributes to hUII-induced vasoconstriction (Saetrum et al. 2000). Here, U-73122, a PLC inhibitor, and 2-APB, an IP3R antagonist, attenuated hUII-induced MAP and RVR elevations, RBF and CBF reductions, but not MBF increase. These data suggest that PLC/IP3-mediated renal vasoconstriction is involved in hUII-induced neonatal renal hemodynamic changes. Of note, 2-APB can also block store-operated Ca2+ (SOC) channels (Dobrydneva & Blackmore 2001, Bootman et al. 2002). Given that UII also induces vasoconstriction by activating store-operated Ca2+ entry (Domínguez-Rodriguez et al. 2012), SOC channel blockade may contribute to the inhibitory effect of 2-APB on hUII-induced neonatal renal hemodynamic changes.

In summary, data from our study indicate that UII regulates renal hemodynamics in neonatal pigs. Intrarenal arterial bolus administration of hUII elevates RVR and MAP but decreases total RBF in newborn pigs. In addition, hUII reduces CBF but increases MBF in the piglets. We also propose that hUII-induced activation of renal vascular UTR stimulates PLC/IP3-mediated vasoconstriction, leading to an increase in RVR and MAP and a decrease in RBF and CBF in newborn pigs.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding
This work was supported by a start-up fund from UTHSC to A Adebiyi.

Acknowledgements
The authors thank Alex Fedinec for technical advice with animal ventilation and Dr Charles Leffler for critical reading of the manuscript.
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Received in final form 1 April 2013
Accepted 3 April 2013
Accepted Preprint published online 3 April 2013