Pro-TRH and pro-CRF expression in paraventricular nucleus of small litter-reared fasted adult rats

F Aréchiga-Ceballos1,2, E Alvarez-Salas1, G Matamoros-Trejo1, M I Amaya1, C García-Luna1 and P de Gortari1

1Neurofisiología Molecular, Dirección de Investigaciones en Neurociencias, Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz (INPRFM), Calzada México-Xochimilco 101, Col. San Lorenzo Huipulco, C.P. 14370, México, Distrito Federal, México
2Escuela de Dietética y Nutrición, ISSSTE, Callejón Vía San Fernando #12, México, Distrito Federal, México

Correspondence should be addressed to P de Gortari
Email gortari@imp.edu.mx

Abstract

Neuroendocrine axes adapt to nutrient availability. During fasting, the function of the hypothalamus–pituitary–thyroid axis (HPT) is reduced, whereas that of the hypothalamus–pituitary–adrenal axis (HPA) is increased. Overfeeding-induced hyperleptinemia during lactation may alter the regulatory set point of neuroendocrine axes and their adaptability to fasting in adulthood. Hyperleptinemia is developed in rodents by litter size reduction during lactation; adult rats from small litters become overweight, but their paraventricular nucleus (PVN) TRH synthesis is unchanged. It is unclear whether peptide expression still responds to nutrient availability. PVN corticotropin-releasing factor (CRF) expression has not been evaluated in this model. We analyzed adaptability of HPT and HPA axes to fasting-induced low leptin levels of reduced-litter adult rats. Offspring litters were reduced to 2–3/dam (early-overfed) or maintained at 8/dam (controls, C). At 10 weeks old, a subset of animals from each group was fasted for 48 h and leptin, corticosterone, and thyroid hormones serum levels were analyzed. In brain, expressions of leptin receptor, NPY and SOCS3, were evaluated in arcuate nucleus, and those of proTRH and proCRF in PVN by real-time PCR. ProTRH expression in anterior and medial PVN subcompartments was assayed by in situ hybridization. Early-overfed adults developed hyperphagia and excessive weight, together with decreased proTRH expression in anterior PVN, supporting the anorexigenic effects of TRH. Early-overfed rats presented low PVN proTRH synthesis, whereas fasting did not induce a further reduction. Fasting-induced stress was unable to increase corticosterone levels, contributing to reduced body weight loss in early-overfed rats. We concluded that early overfeeding impaired the adaptability of HPT and HPA axes to excess weight and fasting in adults.

Introduction

Adaptation of the functioning of neuroendocrine axes allows humans and animals to face environmental challenges such as food availability or stressful conditions by saving energy for the display of defensive actions or by reducing their metabolic rate during food deficit. This adaptability may be compromised in adults subjected to stress, malnutrition (Ravelli et al. 1976, Parsons et al. 2001) or overfeeding during prenatal and early post-natal...
periods (Rodrigues et al. 2009), thus enhancing the risk for the development of different metabolic disturbances.

Our aim was to evaluate the responsiveness of neurons in the paraventricular nucleus (PVN) that express the neuropeptides, pro-thyrotropin-releasing hormone (proTRH) and pro-corticotropin-releasing factor (proCRF), which regulate the hypothalamus–pituitary–thyroid (HPT) and hypothalamus–pituitary–adrenal (HPA) axes. This responsiveness was assessed after the known 48 h fasting-induced shift in leptin and corticosterone-circulating levels in early-overfed animals. In particular, we propose the hypothesis that the function of HPT and HPA axes would be compromised in adults by litter size reduction during lactation, when they are subject to 48-h period of fasting.

Nutrient availability is capable of modifying the functioning of HPT and HPA axes. HPT maintains energy homeostasis and regulates thermogenesis and basal metabolic rate by controlling the serum concentration of thyroid hormones (TSH) – triiodothyronine (T3) and thyroxine (T4) – as well as their degrading effects on fuel reservoirs. TSH in turn induces TH production in the thyroid gland (Nillni 2010). The HPA is driven directly by PVN CRF through the stimulation of corticotropin (ACTH) release from the adenohypophysis, which in turn induces corticosterone release from the adrenal cortex.

Fasting and prolonged food restriction decrease T3 serum levels and mPVN proTRH mRNA content despite the negative feedback that TH generally exerts on TRH synthesis (Blake et al. 1991). TRH downregulation is, at least in part, due to low circulating leptin levels induced by negative energy balance, along with high circulating levels of corticosterone. In addition, we evaluated whether proTRH mRNA levels in the anterior PVN (aPVN) changed in a manner correlated with the serum leptin and corticosterone and leptin serum levels of early-overfed adult rats (Lopez et al. 2005, Rodrigues et al. 2009), proTRH mRNA content in the PVN is unchanged (Lopez et al. 2007). Thus, it is difficult to determine whether the HPT axis of these animals is still responding to changes in serum leptin and corticosterone levels and to nutrient availability. ProCRF expression in the PVN of early-overfed adult rats has not been described, whereas corticosterone serum levels are normal (Boullu-Ciocca et al. 2005). Normal values of PVN proTRH expression and corticosterone and leptin serum levels of early-overfed adult rats do not ensure that neuroendocrine axes can respond to a new metabolic challenge as those of control animals do.

In order to induce a metabolic challenge, we subjected early post-natal overfed adult rats to a 48-h period of fasting and evaluated both expression changes of proTRH mRNA content in the hypophysiotropic neurons of the PVN and HPT adaptation to low circulating levels of leptin and high circulating levels of corticosterone. In addition, we evaluated whether proTRH mRNA levels in the anterior PVN (aPVN) changed in a manner correlated with the hyperphagia of animals, since the peptide from this subcompartment has been implicated in feeding regulation (Wittmann et al. 2009a, Alvarez-Salas et al. 2012). To identify the functioning of the transduction pathway of leptin in adults, we also evaluated expression patterns of target genes of leptin signaling in the arcuate nucleus (ARC) and PVN, such as Lepr and Npy, which directly or indirectly affect PVN proTRH mRNA levels during fasting.

We also analyzed changes in PVN proCRF mRNA levels of both animals allowed to feed ad libitum and fasted animals from normal or small litters, since an appropriate degradation of fuel stores by corticosterone ensures enough energy availability for brain and basal functioning of the organism, and we wanted to elucidate whether this process was adequately achieved by overfed animals.

**Materials and methods**

**Animals**

All experiments were approved by the local committee of ethics on animal experimentation and complied with the

---

http://joe.endocrinology-journals.org
DOI: 10.1538/JOE-13-0458
© 2014 Society for Endocrinology
Printed in Great Britain

Published by Bioscientifica Ltd.

Downloaded from Bioscientifica.com at 11/20/2022 07:47:27PM
via free access
Mexican official norm NOM-062-ZOO-1999 relating to experiments with laboratory animals.

Pregnant female Wistar rats (n=10; 300–350 g) (INPRFM animal house) were individually housed at 25 °C and maintained on a 12 h light:12 h darkness cycle, with free access to standard laboratory rat chow (Lab rodent diet no. 5001; PMI Feeds, Brentwood, MO, USA) and tap water. To induce early post-natal over-feeding, on lactation day 3, litter size for six dams was reduced to two to three male pups, while normal litter groups were formed by four dams with eight pups (males and females) per litter.

A set of males from each size of litter (8/litter size group) was killed at weaning. The remaining males were assigned to four groups: i) control (C, n=7) whose litter was normal sized; ii) early overfed from reduced litters (n=5). Both groups were allowed to feed ad libitum; iii) fasted controls (FC, n=8); and iv) fasted overfed (F-overfed, n=5); rats were fasted 48 h before they were killed. Body weight and food intake were measured weekly, starting at weaning and continuing until 10 weeks of age, when they were killed by decapitation. The experiment was repeated twice, thus the final number of animals per group was 18 in average. Trunk blood was collected to measure T3, T4, and corticosterone in adults, animals per group was 18 in average. Trunk blood was collected to measure T3, T4, and corticosterone in adults, the resulting 14-μm coronal slices were adhered to Superfrost/Plus slides (Fisher Scientific, Pittsburgh, PA, USA). Sections through the whole extension of the PVN were collected (aPVN comprised from −1.1 to −1.5 mm from bregma, and mPVN from −1.5 to −2.3 mm from bregma) (Paxinos & Watson 2005). ProTRH expression was analyzed in two different PVN subcompartments because that of the aPVN is implicated in feeding regulation, and that of the mPVN in the regulation of HPT axis function. As there is no available evidence showing that proCRF is expressed in PVN, we did not perform ISH analysis of proCRF expression. The sections were fixed with 4% paraformaldehyde at 4 °C followed by digestion with pronase (24 U/ml) at 20 °C. The slides were air-dried before hybridization.

### Tissue treatment for ISH histochemistry

Brains stored at −20 °C were sectioned using a cryostat (Microm HM 525; Carl Zeiss IMT Corp., Maple Grove, MN, USA). The resulting 14-μm coronal slices were adhered to Superfrost/Plus slides (Fisher Scientific, Pittsburgh, PA, USA). Sections through the whole extension of the PVN were collected (aPVN comprised from −1.1 to −1.5 mm from bregma, and mPVN from −1.5 to −2.3 mm from bregma) (Paxinos & Watson 2005). ProTRH expression was analyzed in two different PVN subcompartments because that of the aPVN is implicated in feeding regulation, and that of the mPVN in the regulation of HPT axis function. As there is no available evidence showing that proCRF is differentially expressed in aPVN or mPVN in feeding and HPA axis regulation, we did not perform ISH analysis of proCRF expression. The sections were fixed with 4% paraformaldehyde at 4 °C followed by digestion with pronase (24 U/ml) at 20 °C. The slides were air-dried before hybridization.

### ISH histochemistry

Oligonucleotides were used as ISH probes. The following sequences directed against proTRH mRNA (GenBank accession no. NM_013046.2) were labeled by 3′-tailing with [35S]dATP (1250 Ci/mmol; PerkinElmer, Waltham, MA, USA) or with Dig-11-dUTP using terminal deoxynucleotidyltransferase (Roche Diagnostics GmbH): i) 5′-TG CCA TGA ATA CTG CTT GTC GTG GCC AGC TCG GCC GGG GTG CTG-3′; ii) 5′-ATC AGA CTC CAT CCA GGG GAA GGA TCG CCT GCC AGG GTG CTG CCG-3′; iii) 5′-AGG...
The ISH protocol has been described elsewhere (de Gortari & Mengod 2010). Specificity of the oligonucleotides was verified by the unique distribution pattern of the proTRH signal similar to that previously described (Segerson et al. 1987). Also, we observed no signal in the PVN when slices were hybridized with sense sequences of the same oligonucleotides and labeled with digoxigenin (Fig. 1). Briefly, hybridizations were performed by incubating slides overnight at 42 °C with buffer containing 50% formamide/4× SSC, 10% dextran sulfate, 1× Denhardt’s solution (0.02% BSA, 0.02% Ficoll 400, 0.02% polyvinylpyrrolidone), 1% sarkosyl, 20 mM phosphate buffer (pH 7.0), 250 μg/ml yeast tRNA, 500 μg/ml denatured salmon sperm DNA, 5 M dithiothreitol, and 10–20×10⁶ c.p.m. radiolabeled oligonucleotide probe per milliliter of hybridization buffer or 4 pmol of dig-labeled sense or antisense probes per milliliter of hybridization buffer. Slides were washed four times at 60 °C with a buffer containing 0.6 M NaCl and 10 mM Tris–HCl (pH 7.5), and those hybridized with radiolabeled probes were subsequently dipped into Kodak NTB autoradiography emulsion (Eastman Kodak). Autoradiograms were developed with Kodak-D19 after a 3-week exposure at 4 °C. Following the washes (0.6 M NaCl, 10 mM Tris–HCl pH 7.5), brain sections hybridized with sense and antisense dig-labeled probes were immersed in a buffer containing 0.1 M Tris–HCl pH 7.5, 1 M NaCl, 2 mM MgCl₂, and 0.5% BSA (Sigma–Aldrich) and then incubated overnight at 4 °C in the same buffer containing alkaline-phosphatase-conjugated anti-digoxigenin-F(ab) fragments (1:5000) (Roche Diagnostics GmbH). The same solution but without the antibody was used to wash the slides three times for 10 min/each, followed by two washes in an alkaline buffer that contained 0.1 M Tris–HCl pH 9.5, 0.1 M NaCl, and 5 mM MgCl₂. Development of alkaline phosphatase activity was achieved by incubating sections with 3.3 mg nitroblue tetrazolium and 1.65 mg bromochloroindolyl phosphate (Roche Diagnostics GmbH) diluted in 10 ml of alkaline buffer. Thorough rinses in the alkaline buffer containing 1 mM EDTA were used to stop the enzymatic reaction. Finally, the slides were dehydrated rapidly in 70 and 100% ethanol, air-dried, and mounted in 50% of glycerol in PBS solution.

**Image analysis**

Signal from the autoradiograms was visualized using an Olympus BH-2 imaging microscope (New York Microscope Co., Inc., Hicksville, NY, USA) under dark-field illumination. Images were captured with a 4× objective using an Evolution MP Media Cybernetics video camera (Media Cybernetics, Inc., Bethesda, MD, USA) and analyzed using Image-Pro Plus 7.0 computer software (Media Cybernetics, Inc.). For each animal, integrated O.D. values (with background correction) were measured in six rostrocaudal slices which spanned both PVN subdivisions of interest.

**Figure 1**

Bright-field illumination photomicrographs of sections from the medial subdivision of hypothalamic PVN after *in situ* hybridization with digoxigenin-labeled antisense (A) and sense (B) probes directed against proTRH mRNA. Note the lack of specific signal in the PVN with the sense probes. III, third ventricle. Scale bar: 100 μm.
Analysis of hormone serum levels

Serum total T3 concentration was determined by ELISA (Alpco Diagnostics Kit, Salem, NH, USA); sensitivity: 20 ng/dl, inter- and intra-assay variability: 4.9 and 5.6% respectively. Serum corticosterone concentration was assessed by RIA (Coat-A-Count Rat Corticosterone 125 TKRC1, Siemens Medical Solutions Diagnostics, Los Angeles, CA, USA); sensitivity 5.7 ng/ml, inter- and intra-assay variation: 8.5 and 6.8% respectively. Circulating leptin concentration was determined colorimetrically with an ELISA kit for rat (Assay Designs, Ann Arbor, MI, USA); sensitivity: 0.672 ng/ml, inter- and intra-assay variability: 7.2 and 5.9% respectively. Total T4 was also determined by ELISA (Alpco Diagnostics Kit); sensitivity: 0.5 μg/dl, inter and intra-assay variability: 3.6 and 3.8% respectively.

Statistical analysis

A Student’s t-test was performed to compare body weight and food intake between control and early-overfed adult groups on each week of the experiment. A two-way ANOVA was performed to analyze differences between animals allowed to feed ad libitum and fasted adult animals. A one-way ANOVA was used for all parameters in weaned pups. Before the performance of variance analyses, we confirmed the Gaussian distribution of the data. A Pearson correlation was performed between PVN TRH mRNA levels and corticosterone and leptin serum concentrations. When the difference was significant (P<0.05), the analysis was followed by Fisher’s post-hoc test.

Results

Body and white adipose tissue weights

Early-overfed weaned animals had a body weight 20% higher than that of control offspring, and with the exception of week 7, it remained increased up to the end of the experiment. Differences between groups were analyzed by Student’s t-test each week (Fig. 2A). Abdominal fat weight was higher in the early-overfed group by 21% (3.8±0.2 g) vs controls (3.16±0.2 g, 100%). After fasting, FC groups showed a decrease by 33% (2.4±0.16 g) vs controls whereas in the F-overfed group abdominal fat was only reduced by 18% (3.1±0.1 g), but still was different from that of the early-overfed group. A two-way ANOVA showed a significant effect of group: F(1,10)=23.23, P<0.001 and diet: F(1,10)=46.216, P<0.0001 but no significant interaction between variables F(1,10)=0.091, P>0.05.

Body weight loss after fasting

F-overfed adult animals lost 11% of their body weight, whereas the FC group lost 14.3% of the body weight compared with their respective controls. The difference was significant when a Student’s t-test was performed: t=2.189, P<0.05.

Food intake

After weaning, early-overfed animals ate more food than controls starting on week 5 (4%) and up to the end of the experiment (11% on average); differences between groups were evaluated by Student’s t-test every week (Fig. 2B).

Serum hormones

Leptin

Weaned early-overfed animals exhibited a 116% increase in serum leptin levels vs those of the control group (C=1.4±0.2; early overfed=3.1±0.3 ng/ml, F(1,18)=20.922, P<0.001), whereas in adults, there was no difference in leptin concentration between
Experimental and control groups. Fasting reduced leptin levels to undetectable values both in FC and F-overfed animals (Table 1). A two-way ANOVA showed a significant effect of diet: $F_{(1,18)} = 18.54, P < 0.001$ and no significant effect of group: $F_{(1,18)} = 0.486, P > 0.05$ or interaction between variables: $F_{(1,18)} = 0.093, P > 0.05$.

**Corticosterone** Control and early-overfed adults did not differ in serum corticosterone content; FC showed an increase of 92% and F-overfed a decrease to 37% vs their respective controls allowed to feed *ad libitum*. A two-way ANOVA showed a significant effect of groups: $F_{(1,32)} = 5.740, P < 0.05$ and interaction between variables: $F_{(1,32)} = 5.791, P < 0.05$ (Table 1).

**Triiodothyronine** Controls and early-overfed adults presented similar T₃ serum content; fasting reduced levels 8.6, $P = 0.0001$, and interaction between variables: $F_{(1,32)} = 12.014, P < 0.01$, diet: $F_{(1,16)} = 11.915, P < 0.01$ and interaction between variables: $F_{(1,16)} = 9.584, P < 0.05$ (Table 1).

**Thyroxine** Early-overfed adults showed a slight reduction of 16% in T₄ circulating levels vs controls. After fasting, FC and F-overfed groups exhibited 32 and 21% lower T₄ than their respective groups allowed to feed *ad libitum*. There was a significant effect of group: $F_{(1,10)} = 8.6, P < 0.01$, diet: $F_{(1,16)} = 69.63, P < 0.0001$, and interaction between variables: $F_{(1,16)} = 4.888, P < 0.05$ (Table 1).

**Corticosterone** Controls and early-overfed adults did not differ in serum corticosterone content; FC showed an increase of 92% and F-overfed a decrease to 37% vs their respective controls allowed to feed *ad libitum*. A two-way ANOVA showed a significant effect of groups: $F_{(1,32)} = 5.740, P < 0.05$ and interaction between variables: $F_{(1,32)} = 5.791, P < 0.05$ (Table 1).

Changes in hypothalamic mRNA peptide expression

**ARC** ObRb expression was higher in ARC for all experimental groups – early-overfed, FC, and F-overfed – compared with controls but no difference was found between experimental groups. ANOVA showed a significant effect of diet: $F_{(1,10)} = 15.09, P < 0.01$ and interaction between variables: $F_{(1,10)} = 20.017, P < 0.01$ (Fig. 3). SOCS3 mRNA content increased in FC and F-overfed vs controls. The highest expression was found in the F-overfed group which was significantly different from the other three groups. ANOVA showed a significant effect of group: $F_{(1,8)} = 10.67, P < 0.05$, diet: $F_{(1,8)} = 25.9, P < 0.001$, and interaction between variables: $F_{(1,8)} = 5.71, P < 0.05$ (Fig. 3). NPY mRNA content increased only in the FC group vs controls. A two-way ANOVA yielded a significant effect of group: $F_{(1,8)} = 12.014, P < 0.01$, diet: $F_{(1,8)} = 11.815, P < 0.01$ and interaction between variables: $F_{(1,8)} = 9.584, P < 0.05$ (Fig. 3).

**Paraventricular nucleus** ObRb and SOCS3 expression did not change in any of the groups. ProTRH mRNA levels decreased in all experimental groups vs controls; reduction of proTRH expression in the early overfed group was greater than the one induced by fasting in the FC group. A two-way ANOVA showed a significant effect of group: $F_{(1,11)} = 37.11, P < 0.0001$, diet: $F_{(1,11)} = 8.45, P < 0.05$ and interaction between variables: $F_{(1,11)} = 20.454, P < 0.001$ (Fig. 3). Correlations between TRH mRNA levels and those of corticosterone and leptin concentrations in C rats resulted significantly negative ($r = -0.917, P < 0.05$) and positive ($r = 0.88, P < 0.05$), respectively, but no correlation was found in early-overfed animals (corr: $r = -0.68, P > 0.05$; leptin: $r = -0.49, P > 0.26$). ProCRF expression was also lower in the three experimental groups when compared with controls; ANOVA showed a significant effect of group: $F_{(1,10)} = 21.012, P < 0.01$, diet: $F_{(1,10)} = 5.313, P < 0.05$, and interaction between variables: $F_{(1,10)} = 5.754, P < 0.05$ (Fig. 3).

**ProTRH expression in PVN subcompartments by ISH**

The early-overfed group exhibited a reduction to 35% in proTRH expression in the aPVN, whereas in FC it decreased to 42% when compared with controls (100%). The F-overfed group did not show any difference vs C. A two-way ANOVA showed a significant interaction between diet and group: $F_{(1,21)} = 9.7, P < 0.01$ (Figs 3 and 4).

In the mPVN, early-overfed animals showed a decrease of 47.8% in proTRH mRNA levels vs controls. Fasting also induced a decrease to ~47% in both FC and F-overfed groups vs controls allowed to feed *ad libitum* (100%).

### Table 1 Serum hormone concentrations. Serum hormone content was evaluated in weaned adult animals allowed to feed *ad libitum* or fasted. The number of animals for T₃ determination was 18/group; for T₄, n=5/group, and for corticosterone and leptin, n=10/group. Values are the mean±SEM and are expressed in ng/ml for leptin and corticosterone; ng/dl for T₃; μg/dl for T₄. Differences were found by two-way ANOVA between data of groups and diets followed by Fisher’s post-hoc test, when differences were significant.

<table>
<thead>
<tr>
<th></th>
<th>C</th>
<th>Overfed</th>
<th>FC</th>
<th>F-overfed</th>
</tr>
</thead>
<tbody>
<tr>
<td>T₃ (ng/dl)</td>
<td>100±4</td>
<td>98±4</td>
<td>75±4.8*$^+$</td>
<td>83±5.7*</td>
</tr>
<tr>
<td>T₄ (μg/dl)</td>
<td>14±0.3</td>
<td>11.9±0.4*</td>
<td>9.6±0.35*</td>
<td>9.3±0.5*</td>
</tr>
<tr>
<td>Leptin (ng/ml)</td>
<td>2±0.3</td>
<td>1.67±0.2</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Corticosterone (ng/ml)</td>
<td>99±20</td>
<td>102±35</td>
<td>192±45*$^+$</td>
<td>37±5*$^+$</td>
</tr>
</tbody>
</table>

C, control; FC, fasted-control; F-overfed, fasted-overfed; ND, not-detectable values. $^+$P<0.05 compared to C; $^*$P<0.05 vs early-overfed.
A two-way ANOVA showed a significant effect of group: \(F_{(1,26)} = 7.324, P < 0.05\), diet: \(F_{(1,26)} = 7.2, P < 0.05\), and interaction between variables: \(F_{(1,26)} = 7.237, P < 0.05\) (Figs 4 and 5).

Discussion

During critical stages of development, alterations in serum hormone concentrations involved in energy balance regulation, as well as high glycemia due to overfeeding or to a diabetic dam, impair the ability of neuroendocrine axes to adapt to metabolic or stressful challenges in adult humans and animals. These early disturbances may put the survival of an organism at risk or increase its susceptibility to developing pathologies such as the metabolic syndrome later in life. The aim of this study was to evaluate the adaptability of HPT and HPA axes to a 48-h period of fasting in adult animals that presented hyperleptinemia and overweight during lactation due to litter size reduction.

Early-overfed adult rats allowed to feed ad libitum exhibited similar changes to those previously observed by others regarding body weight, leptin levels, and mRNA expression of ObRb and NPY in the arcuate nucleus (Lopez et al. 2005, 2007, Rodrigues et al. 2011). In addition, the expected hyperleptinemia and overweight were present in weaned early-overfed animals.

Body weight and food intake in the early-overfed group were higher than those of controls starting at weaning and up to the tenth week of age, as described in Figure 3.

Figure 3
Peptide mRNA content in arcuate and paraventricular nuclei of adult animals, analyzed by quantitative real-time PCR. Values are mRNA expression of each examined amplicon (for ARC: ObRb, SOCS3, NPY; for PVN: ObRb, SOCS3, proTRH and proCRF) after being normalized against a factor consisting in the geometric mean of the expression of two reference genes: Actb and Gapdh, and expressed as the mean \(\pm\) S.E.M. of the % of controls (100%) (\(n=5/\)group). A two-way ANOVA was performed to find differences between groups and diets of adult animals: *\(P < 0.05\) vs C (controls); *\(P < 0.05\) vs FC (fasted-controls); *\(P < 0.05\) vs F-overfed (fasted-overfed).

Figure 4
ProTRH mRNA expression in PVN subcompartments determined by ISH. Changes in proTRH mRNA levels in aPVN and mPVN of control (C), early-overfed, control animals fasted for 48 h (FC) and animals from small litters fasted for 48 h (F-overfed) animals quantified by ISH histochemistry. Rats were killed at 10 weeks of age. Values are expressed as the mean \(\pm\) S.E.M. of the percentage for controls (100%) of integrated O.D. *\(P < 0.05\) vs C; *\(P < 0.05\) vs F-overfed.
previously (Davidowa et al. 2003, Boullu-Ciocca et al. 2005). Increased body weight and white adipose tissue mass of early-overfed adults did not correlate with their circulating leptin levels, which remained similar to those of controls. A plausible explanation for this are the basal corticosterone levels found in early-overfed rats, which were similar to control values. It is commonly observed that circulating glucocorticoid concentrations are higher in overweight humans and animals; glucocorticoids are well-known secretagogues of leptin (Hardie et al. 1996, Russell et al. 1998, Williams et al. 2000). As we discuss below, we found a deregulated adrenal axis in early-overfed adult rats that could contribute to the unchanged circulating leptin levels in this group. Rodrigues et al. (2011) have also reported leptin levels similar to controls in early-overfed groups using the same model.

We found enhanced ObRb and SOCS3 expression in the ARC of early-overfed animals, supporting the resistance of this nucleus to the effects of leptin, as has been suggested by other authors (Rodrigues et al. 2011). SOCS3 is a negative regulator of the leptin signaling cascade; hence its higher expression may impair the positive and negative effects of leptin on the synthesis of anorexigenic and orexigenic peptides respectively. However, NPY expression was not different in early-overfed compared with control adults, as has been described previously (Lopez et al. 2007), which could argue against leptin resistance in the early-overfed animals allowed to feed ad libitum. As this group showed serum leptin levels in the normal range, we needed to challenge the ARC response to fasting-induced hypoletinemia to confirm a blocked leptin signaling. We also observed basal values of glucocorticoids in the early-overfed group; these compounds are known to stimulate NPY synthesis in the ARC (White et al. 1990, Sato et al. 2005, Goto et al. 2006), thus by increasing serum glucocorticoid levels after fasting we could possibly identify the response of NPY-expressing neurons to another peripheral signal of energy status.

TRHergic neurons implicated in hypophysiotrophic functions are only those of the medial parvocellular part of the PVN (Wittmann et al. 2009b). As a consequence, we evaluated proTRH expression in this subcompartment to assay neurons driving the function of the HPT axis. T4 serum content is slightly decreased in the early-overfed group according to Plagemann et al. (1999) and we found that proTRH expression in the mPVN subcompartment was also lower than that in controls. Since early-overfed adult rats were overweight, the HPT axis must be activated, as is observed in obese humans and in some obesity models in rodents (Reinehr & Andler 2002, Perello et al. 2010). However, the early-overfed group exhibited an altered flexibility of the HPT axis to increased adiposity, which might induce, in the long term, deceleration of the metabolic rate, and favor the development of obesity and its complications. These results indicate that central hypothyroidism may be the cause of the slightly low TH serum content found in overweight early-overfed adults.

![Figure 5](http://joe.endocrinology-journals.org/C209/2014/Society-for-Endocrinology/DOI: 10.1530/JOE-13-0458/Printed-in-Great-Britain)

**Figure 5**

Dark-field illumination photomicrographs of representative samples of proTRH mRNA in PVN. Images are low-power magnifications of anterior (1) and medial (2) subdivisions of the PVN of control group (A), early-overfed group (B), control animals fasted for 48 h (FC) group (C) and animals from small litters fasted for 48 h (F-overfed) group (D). Scale bar, 200 μm.
PVN TRHergic cells are innervated by ascending catecholaminergic neurons from the brainstem (Cunningham & Sawchenko 1988, Füzesi et al. 2009). Noradrenaline (NA) is an upregulator of hypophysiotropic proTRH expression (Uribe et al. 1993, Perello et al. 2007). Importantly, brain NA concentrations in early-overfed animals are low (Seidler et al. 1990), and the binding of adenergetic receptors in PVN is reduced in diet-induced obese rats (Levin 1996). This is also indicated by the reduced catecholamine release in caffeine-stimulated adrenal cells of early-overfed adult rats produced by litter reduction, as well as by low expression of the β2-adrenergic receptor evaluated in liver homogenates, which may lead to decreased peripheral sensitivity to catecholamines and lipolysis rate (Conceição et al. 2013). Thus, NA is a candidate inhibitor of TRH synthesis in the mPVN, as well as that of CRF as described below.

TRH-expressing neurons of the aPVN do not project to the median eminence, but to other regions involved in feeding regulation, such as the dorsomedial and ARC hypothalamic nuclei (Wittmann et al. 2009a). In addition, proTRH expression in aPVN is increased in food-avoiding animals (Alvarez-Salas et al. 2012). We found that in this subcompartment proTRH mRNA levels were down-regulated in the early-overfed group, supporting the anorexigenic role of aPVN and, in part, the underlying hyperphagia of the early-overfed group. This is in agreement with early studies showing that PVN destruction leads to hyperphagia and overweight (Leibowitz et al. 1981, Fletcher et al. 1993). Leptin signaling in the PVN of the early-overfed group seemed functional considering the observed normal mRNA levels of ObRb and SOCS3; this is evident in the reduction of food intake of early-overfed adults receiving an i.c.v. injection of leptin (Lozano et al. 2007). Thus, we cannot discard the possibility that proTRH expression reduction in this group was more probably due to ARC feeding-regulatory neuropeptides, whose effects on PVN are altered by early post-natal hyperleptinemia (Davidowa et al. 2003). This hypothesis is likely given that aPVN and mPVN TRHergic neurons are innervated by axons containing α-MSH and AgRP receptors (Legradi & Lechan 1999, Fekete et al. 2000); but this issue needs to be elucidated.

PVN proCRF expression, which has anorexigenic effects (Arase et al. 1988), was decreased in the early-overfed group vs controls. As happened with proTRH from aPVN, this is also in accordance with the hyperphagic state of the early-overfed group. Early-overfed animals showed similar corticosterone serum levels to those of controls, as others have found (Boullu-Ciocca et al. 2005, Rodel et al. 2010, Bulfin et al. 2011). The decreased proCRF expression and normal corticosterone serum levels of the early-overfed group are not congruent with the negative feedback regulation of the HPA axis exerted by glucocorticoid levels. Our results indicate (and were confirmed by the results for the fasted-overfed group) an altered regulation of the adrenal axis in early-overfed adults. This result supports the idea of an impaired afferent catecholaminergic pathway to the PVN in the overweight early-overfed group, because CRF is upregulated by NA (Cole & Sawchenko 2002). Interestingly, fetally fetal undernourished adults rats present high proCRF expression (Núñez et al. 2008), as well as enhanced brain noradrenergic synthesis and release (Soto-Moyano et al. 1998a,b).

Fasting

We evaluated the adaptability of the HPT axis to reduced leptin content induced by a 48-h period of fasting, which decreases body weight and represents a new metabolic challenge. The FC group displayed the expected changes induced by low nutrient availability: decreased leptin and increased serum corticosterone concentrations, whereas F-overfed animals showed reduced leptin levels but not the activation of the adrenal axis, confirming its deregulation in early post-natal overweight adult rats.

In ARC of the FC group, NPY, SOCS3, and ObRb synthesis were elevated, which is in agreement with the low leptin levels observed. These changes along with reduced proTRH expression in aPVN and mPVN were coincident with the expected stimulation of hypothalamic orexigenic and inhibition of anorexigenic peptides as a result of a negative energy balance. This reduced PVN proTRH expression could be due to the combined actions of high NPY release into this nucleus, as proposed by others (Legradi & Lechan 1999, Fekete et al. 2001), and supported here by high NPY synthesis in ARC. As mentioned earlier, it has also been proposed that T₃ local hypothalamic content increases by the action of D₂ as a result of fasting-induced low circulating levels of leptin and high circulating levels of corticosterone (Coppola et al. 2005b). High hypothalamic concentrations inhibit TRH synthesis (Coppola et al. 2005a). This could not be the case however in F-overfed animals because they also presented peripheral hypothyroidism along with downregulation of proTRH in mPVN despite their low leptin and corticosterone serum levels, which may not be elevating local T₃ content (Coppola et al. 2005b). The absence of a correlation between serum corticosterone and leptin levels with PVN proTRH expression in the F-overfed group supports the hypothesis of an impaired response of TRHergic cells to...
those hormones after fasting. These results are evidence of alternative factors capable of decreasing proTRH expression during fasting. We can discard the inhibitory effects of NPY on TRH synthesis, since its expression in ARC was normal in the overfed group. Furthermore, altered actions of NPY on PVN neurons have been described in early-overfed animals (Davidowa et al. 2003). Besides, the lack of an increase in ARC NPY expression in F-overfed rats revealed the impaired leptin signaling of this nucleus in early post-natal-overfed animals.

The low corticosterone content in the F-overfed group was surprising because during fasting its elevation facilitates degradation of lipid deposits and ensures glucose availability for brain and peripheral tissues. Thus, this was evidence of a disturbed HPA response to metabolic challenges in the F-overfed group that puts at risk the survival of animals facing a stressful event. Fasting activates NA afferent projections to PVN CRF-expressing neurons, stimulating CRF synthesis and release, which in turn enhances corticosterone serum levels. As part of the negative feedback, glucocorticoids downregulate PVN proCRF expression, which was evident here in the FC group. However, in F-overfed animals, decreased proCRF mRNA levels cannot be due to circulating levels of corticosterone since these were reduced in this group. The proposed impaired activation of NA connections to the PVN is supported by those changes in fasted early-overfed rats. Low corticosterone serum concentration may be avoiding degradation of energy stores by lowering body weight loss in the F-overfed group.

We showed here that early post-natal overfeeding and hyperleptinemia induced in animals by litter size reduction resulted in a long-lasting impairment in the adaptability of HPT and HPA axes to high adiposity and to a novel metabolic challenge such as fasting. Reduced expression of proTRH and proCRF in the PVN of early-overfed animals may contribute to their hyperphagia. These alterations diminished fuel deposit degradation and weight loss during low energy intake.

This animal model may be useful for understanding the refractoriness of adults with infant obesity or the offspring of diabetic mothers (Dussault et al. 1982, Walker & Courtin 1985, Pracyk et al. 1992, Cetin et al. 2000, Tapanainen et al. 2001, Wilcoxon & Redei 2004, Moura et al. 2008) to reduce weight in response to diet therapy and their propensity to develop obesity and its co-morbidities.

Funding
The study was supported by Consejo Nacional de Ciencia y Tecnología (CONACYT, project No.128316).

Acknowledgements
The authors thank Raúl Cardoso and José Luis Calderón for their help with figures and images, and M C Víridiana Alcántara for quantitative real-time PCR technical support.

References


de Gortari P & Mengod G 2010 Dopamine D1, D2 and mu-opioid receptors are co-expressed with adenyl cyclase 5 and phosphodiesterase 7B mRNAs in striatal rat cells. Brain Research 1310 37–45. (doi:10.1016/j.brainres.2009.11.009)


Nillni EA 2010 Regulation of the hypothalamic thyrotropin releasing hormone (TRH) neuron by neuronal and peripheral inputs. Frontiers in Neuroendocrinology 31 134–156. (doi:10.1016/j.yfrne.2010.01.001)


Received in final form 20 January 2014
Accepted 23 January 2014
Accepted Preprint published online 24 January 2014