BBT improves glucose homeostasis by ameliorating β-cell dysfunction in type 2 diabetic mice

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Abstract

Impaired glucose-stimulated insulin secretion (GSIS) and increasing β-cell death are two typical dysfunctions of pancreatic β-cells in individuals that are destined to develop type 2 diabetes, and improvement of β-cell function through GSIS enhancement and/or inhibition of β-cell death is a promising strategy for anti-diabetic therapy. In this study, we discovered that the small molecule, N-(2-benzoylphenyl)-5-bromo-2-thiophenecarboxamide (BBT), was effective in both potentiating GSIS and protecting β-cells from cytokine- or streptozotocin (STZ)-induced cell death. Results of further studies revealed that cAMP/PKA and long-lasting (L-type) voltage-dependent Ca2+ channel/CaMK2 pathways were involved in the action of BBT against GSIS, and that the cAMP/PKA pathway was essential for the protective action of BBT on β-cells. An assay using the model of type 2 diabetic mice induced by high-fat diet combined with STZ (STZ/HFD) demonstrated that BBT administration efficiently restored β-cell functions as indicated by the increased plasma insulin level and decrease in the β-cell loss induced by STZ/HFD. Moreover, the results indicated that BBT treatment decreased fasting blood glucose and HbA1c and improved oral glucose tolerance further highlighting the potential of BBT in anti-hyperglycemia research.

Key Words
- β-cell dysfunction
- glucose-stimulated insulin secretion (GSIS)
- β-cell death
- glucose homeostasis

Introduction

Type 2 diabetes is a global epidemic chronic disease that is characterized by resistance in responding to the normal actions of insulin and progressive dysfunction of pancreatic β-cells in secreting sufficient amounts of insulin for normal metabolic demand (Leahy 1990). Pathologically, dysfunction of β-cells is characterized by the loss of glucose-stimulated insulin secretion (GSIS) and β-cell death that leads to a reduction in insulin content and β-cell mass. Thus, amelioration of β-cell dysfunction by activating GSIS or/and blocking β-cell death has been determined to be one of the promising approaches in delaying or preventing the onset of type 2 diabetes (Rhodes 2005, Prentki & Nolan 2006, Seino et al. 2011, Rachdi et al. 2014).

The mechanism of GSIS is well elucidated. It is suggested that glucose is transported into β-cells by GLUT2 and metabolized through the tricarboxylic acid cycle and mitochondrial electron transport chain, thus inducing the increase in the ratio of ATP:ADP (Brownlee 2005). The elevated ATP:ADP ratio stimulates the closure...
of K<sub>ATP</sub> channels, leading to membrane depolarization. Long-lasting (L-type) voltage-dependent Ca<sup>2+</sup> channel (L-VDCC) is then opened and the extracellular Ca<sup>2+</sup> enters into cells, finally triggering insulin release (Suzuki et al. 1999). In addition, extracellular Ca<sup>2+</sup> influx through L-VDCC also induces Ca<sup>2+</sup> release from endoplasmic reticulum. Elevation of intracellular Ca<sup>2+</sup> activates Ca<sup>2+</sup>/calmodulin to phosphorylate diverse substrates involved in insulin secretion, including Ca<sup>2+</sup>/CaM protein kinase 2 (CaMK2), Ca<sup>2+</sup>/CaM kinase (CaMKK), protein kinase C (PKC), and ERK1/2 (Easom 1999). Besides intracellular Ca<sup>2+</sup>, intracellular cAMP also plays an important role in insulin secretion (Cognard et al. 2013). Elevation of intracellular cAMP directly activates cAMP-dependent PKA, thereby potentiating GSIS (Kaihara et al. 2013). In the clinical treatment of type 2 diabetic patients, sulfonylureas are the most widely used drugs to enhance insulin secretion, although they may cause hypoglycemia because of their long duration and glucose-independent effect on insulin secretion (Meeece 2007, Seino 1999). Recently, GLP1 analogs and DPP4 inhibitors, such as exendin-4 and vildagliptin, have been demonstrated to have glucose-dependent effects in stimulating GSIS (Ahren 2008, Chia & Egan 2008, Doupis & Veves 2008, Girgis & Champion 2011).

Apart from GSIS dysfunction, reduced β-cell mass is also strongly associated with the development of type 2 diabetes (Butler et al. 2003). Chronic hyperglycemia induces high levels of reactive oxygen species and interleukin 1B (IL1B), causing β-cell death (Maedler et al. 2002, Kajimoto & Kaneto 2004), and abnormal accumulation of lipids in β cells by hyperlipidemia can also induce β-cell death (Wrede et al. 2002). It has been reported that the cAMP-mediated cytoprotection against apoptosis involves PKA, MAPK, and PI3K pathways (Webster & Anwer 1998), and GLP1 demonstrates anti-apoptotic activity by activating these pathways in pancreatic cells (Hui et al. 2003). In addition, rosiglitazone (Han et al. 2010), GLP1, and its analogues (Li et al. 2005) could also inhibit the glucolipotoxicity-induced β-cell death.

Therefore, based on the above analyses, it is believed that a reagent able to both activate GSIS and protect against β-cell death may demonstrate potent anti-diabetic potential. Accordingly, we performed the screening against the Specs commercial compound library and the small molecule, N-(2-benzoylphenyl)-5-bromo-2-thiophencarboxamide (BBT, Fig. 1A), was identified. BBT could not only potentiates GSIS but also protect β-cells from streptozotocin (STZ)- or cytokine-induced β-cell death, and effectively ameliorate glucose homeostasis in STZ/high-fat diet (HFD)-induced type 2 diabetic mice. Further study of the mechanism revealed that cAMP/PKA pathways are responsible for functions of BBT in GSIS stimulation and β-cell protection. Our current work has highlighted the potential of BBT in the treatment of hyperglycemia.

**Materials and methods**

**Materials and reagents**

GF109203X (GFX), H89, MDL-12,330A, EGTA, repaglinide, STO-609, KN93, and nifedipine were purchased from Sigma–Aldrich. BBT was obtained from a commercial compound library (Specs, Zoetermeer, the Netherlands).

**Cell culture**

INS-832/13 cells were kindly provided by Professor Yong Liu (Institute for Nutritional Sciences, SIBS, Chinese Academy of Sciences) and grown in RPMI 1640 medium supplemented with 10% FBS, 5.6 mM glucose, 25 mM HEPES, 2 mM l-glutamine, 50 mM β-mercaptoethanol, 100 U/ml penicillin, and 100 mg/ml streptomycin at 37 °C in a humidified atmosphere containing 95% air and 5% CO<sub>2</sub>. Culture medium, FBS, and supplements were purchased from Invitrogen. Pancreatic islets from C57BL6 mice (6 weeks) were isolated using the liberase digestion method (Yao et al. 2013). Briefly, pancreas was digested with liberase enzyme, and the digest was fractionated by gradient centrifugation preformed in 13, 21, 23, and 25% ficoll. The islet fractions were then harvested between 13 and 21% gradients into Hank’s balanced salt solution (Sigma), and finally individual mouse islets were hand-picked with a pipette under a microscope. The islets were cultured in RPMI 1640 supplemented with 10% FBS at 37 °C in an environment of 5% CO<sub>2</sub> overnight.

**3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide and 5-Bromo-2'-deoxyuridine assays**

INS-832/13 cells were seeded at a density of 2×10<sup>4</sup> cells/well in 48-well plates and allowed to attach overnight, they were then treated with BBT and STZ (0.4 mM) or cytokines (10 ng/ml tumor necrosis factor alpha (TNFα) and 5 ng/ml IL1B) for 24 h, and incubated with 0.5 mg/ml of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) for 4 h to test cell viability. The formazan crystals formed were dissolved in DMSO and analyzed using a Benchmark Plus microplate reader at 570 nm.
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Intracellular Ca^{2+} measurement

INS-832/13 cells were plated into 96-well plates at a density of 1×10^4 cell/well and incubated overnight. After 1 h starvation in glucose-free KRB buffer, cells were loaded with 80 μl of the Ca^{2+} dye (Fluo-4 AM) and then incubated at 37 °C for 40 min. Intracellular Ca^{2+} influx was analyzed by FlexStation II384 (Molecular Devices, Sunnyvale, CA, USA) with an excitation wavelength of 485 nm and an emission wavelength of 525 nm. After the baseline fluorescence signal was measured for the first 16 s at 2.8 mM glucose, the test compound prepared as a 3× stock in Hank’s balanced salt solution buffer and 16.8 mM glucose was added to the plate through an automated pipette (50 μl/well). The relative fluorescence signals were measured at 1.6 s intervals for 60–120 s.

Luciferase activity assay

Luciferase report plasmid (CRE-Luc) and Renilla luciferase plasmid were kindly provided by Professor Yong Liu (Institute for Nutritional Sciences, SIBS, Chinese Academy of Sciences). HEK293T or INS-832/13 cells were transfected

Figure 1

BBT-potentiated GSIS and protected against β-cell death. (A) Chemical structure of BBT. (B) After 2-h incubation with glucose-free KRB buffer, INS-832/13 cells were stimulated with 2.8, 5.6, 11.2, or 16.8 mM glucose, respectively, in the presence or absence of BBT (20 μM) for 2 h, and insulin secretion was determined by ELISA assay. (C) After 2 h incubation with glucose-free KRB buffer, primary islets were stimulated with 2.8 or 16.8 mM glucose in the presence of BBT (20 μM) for 2 h, and then insulin secretion was determined. (D) STZ (0.4 mM) or (F) cytokines (10 ng/ml TNFα and 5 ng/ml IL1β) were used to induced INS-832/13 cell death, BBT (5, 10, and 20 μM) co-incubated with STZ or cytokines for 24 h, and cell viability was determined by MTT assay. (G) INS-832/13 cells were incubated with BBT (10 and 20 μM) for 24 h, and then cell proliferation was detected by 5-Bromo-2′-deoxyuridine (BrdU) assay. INS-832/13 cells were incubated in the glucose-free KRB buffer containing (H) STZ or (I) cytokines (10 ng/ml TNFα and 5 ng/ml IL1β) in the presence of the indicated compounds for 2 h, and then stimulated with 16.8 mM glucose KRB buffer in the presence of BBT (20 μM) for 2 h, and insulin secretion was determined. All data were obtained from three independent experiments and are presented as means ± S.E.M. (*P<0.05, **P<0.01, and ***P<0.001).
with cAMP-responsive luciferase reporter plasmid (CRE-Luc) plus Renilla luciferase plasmid for 5 h, and then incubated with the indicated concentrations of compounds for another 18 h. Finally, the cells were lysed and luciferase activities measured using the Dual Luciferase Assay System Kit (Promega). The results were presented as the fold-change of activation compared with the untreated cells after normalization using Renilla luciferase values. Each experiment was repeated at least three replicates.

**GSIS assay**

INS-832/13 cells were seeded in 24-well plates and incubated in 5% CO₂ for 48 h. Briefly, cells were pre-incubated with Krebs–Ringer bicarbonate HEPES buffer (KRB buffer: 115 mM NaCl, 5 mM KCl, 1 mM MgCl₂, 24 mM NaHCO₃, 2.5 mM CaCl₂, and 10 mM HEPES) supplemented with 0.5% BSA for 2 h, followed by incubation in KRB buffer containing 2.8, 5.6, 11.2, or 16.8 mM glucose with indicated compounds for 2 h. The supernatant of the cells was collected for the determination of insulin secretion. Insulin concentration was measured using the Rat/Mouse Insulin ELISA Kit (Millipore, Billerica, MA, USA).

**Western blotting analysis**

MABs against ERK1/2 and p-ERK1/2 were purchased from Cell Signaling Technology. For western blot analysis, cell lysate was separated by SDS–PAGE and transferred to PVDF membrane (Amersham Biosciences). After incubation with corresponding antibodies overnight, the membranes were visualized using the Dura detection system (Thermo Scientific, Waltham, MA, USA). For immunohistochemistry analysis, the pancreas was removed and fixed in 10% paraformaldehyde solution. The tissues were subsequently embedded in paraffin and cut into sections with a thickness of 5 μm using a microtome. The sections were incubated with insulin antibody for 1 h and then biotinylated rabbit anti-goat IgG for 30 min at room temperature, and labeled using a modified avidin–biotin complex immunoperoxidase staining procedure. Positive staining was visualized using DAB peroxidase substrate solution for 5–10 min, and tissues were counterstained with hematoxylin.

**Intracellular cAMP assay**

Intracellular cAMP was detected by using the ELISA Assay Kit (R&D, Minneapolis, MN, USA). INS-832/13 cells were washed three times in cold PBS (137 mM NaCl, 2.7 mM KCl, 10 mM Na₂HPO₄, 2 mM KH₂PO₄, and pH 7.2), and then resuspended in lysis buffer to a concentration of 10⁷ cells/ml. The cells were frozen at −20 °C and thawed at room temperature for three cycles, and then centrifuged at 600 g for 10 min at 4 °C. The supernatant was analyzed according to the manufacturer’s protocol. cAMP content was calculated by the standard curve.

**Type 2 diabetic animal model and BBT administration**

All animals were received humane care, and the animal-related protocols were approved by the Institutional Animal Care and Use Committees at Shanghai Institute of Materia Medica, CAS. The STZ/HFD-induced type 2 diabetic mice model was constructed mainly based on the published approaches (Wang et al. 2000, Wu et al. 2005, Arulmozhi et al. 2008). Six-week-old male mice were fed with HFD (consisting of 58% fat, 17% carbohydrate, and 25% protein) for 4 weeks and then administered STZ at 25 mg/kg in 0.1 M citrate-buffered saline (pH 4.5) by intraperitoneal (i.p.) injection after 8 h fasting for 5 consecutive days. Fasting plasma glucose was measured in STZ-injected mice after 3 days to select the diabetic mice for use. Diabetic mice were subdivided into two groups: the control group (n=9) and the BBT treatment group (50 mg/kg per day) (n=9). BBT was administrated by i.p. injection daily for 4 weeks. Fasting plasma glucose was measured weekly. At the termination of the study, mice were killed and tissues were analyzed. In addition, the mice administrated HFD or normal chow diet were used to evaluate the effects of BBT on β-cell mass. Six-week-old male mice were fed with HFD similarly to the HFD/STZ model mice or with normal chow for 6 weeks and then i.p. injected daily with BBT for 4 weeks.

**Oral glucose tolerance test**  The oral glucose tolerance test (OGTT) was carried out on the STZ/HFD-induced diabetic mice fasted overnight. 1.5 g/kg glucose was administered orally with a gavage needle. Glucose levels were measured from tail blood at 0, 15, 30, 60, 90, and 120 min using the ACCU-CHEK Active blood sugar system (Roche). Meanwhile, the insulin release during OGTT was also measured, blood samples were obtained from tail veins and serum insulin concentration was determined using the ELISA Kit (Millipore).

**Insulin tolerance test**  The insulin tolerance test (ITT) was carried out on the STZ/HFD-induced diabetic mice fasted overnight. 1 U/kg insulin was administered by peritoneal injection. Glucose levels were measured from...
Statistical analysis

Data are shown as mean ± S.E.M. Student’s t-test was performed for comparison of two groups and one-way ANOVA for comparison of more than two groups using GraphPad Prism 5.0 Software (La Jolla, CA, USA) with values of \( P<0.05 \) being statistically significant. Significant differences are indicated by *\( P<0.05 \), **\( P<0.01 \), and ***\( P<0.001 \).

Results

BBT improved pancreatic β-cell dysfunction

INS-832/13 cells which present high response to changes in glucose concentration (Hohmeier et al. 2000) were used to screen the active compounds from the Specs commercial library with features of potentiating GSIS and protecting β-cells. After screening approximately 8000 compounds, BBT was finally determined to have the most potent dual-function among 12 candidates.

BBT-potentiated GSIS As indicated in Fig. 1B, BBT-stimulated insulin secretion in a glucose-dependent manner. The results shown in Fig. 1C further confirmed that BBT dose-dependently potentiated GSIS in the cells with a high concentration of glucose (16.8 mM). In addition, we also isolated the primary islets and detected the effects of BBT on GSIS. As indicated in Fig. 1D, BBT enhanced GSIS in primary islets. These results indicated that BBT affected GSIS per se.

BBT protected against β-cell death In the investigation of BBT protection against β-cell death, the compound was assayed for its activity in protecting against the STZ- or cytokine-induced INS-832/13 cell death. As shown in Fig. 1E and F, treatment with STZ (0.4 mM) or cytokines (10 ng/ml TNFα and 5 ng/ml IL1B) induced β-cell death, and BBT effectively reversed this effect in a dose-dependent manner. In addition, BBT itself had no effects on β-cell proliferation according to the BrdU assay (Fig. 1G).

BBT improved the STZ- or cytokine-induced GSIS defect In addition, by considering that STZ or cytokines can induce β-cell secretory defects (Delaney et al. 1995, Andersson et al. 2001), we also examined whether BBT was effective in improving β-cell secretory defects induced by treatment with STZ or cytokines. As shown in Fig. 1H and I, BBT could expectedly reverse the STZ- or cytokine-induced GSIS defect.

Accordingly, all the above results indicated that BBT improved pancreatic β-cell dysfunction by both potentiating GSIS and protecting β-cells from the STZ- or cytokines-induced cell death.

ERK1/2 regulation is involved in the BBT-potentiated GSIS

Next, we tried to investigate the underlying mechanism of BBT in the improvement of pancreatic β-cell dysfunction. It has been reported that ERK phosphorylation, as one of the MAPK signaling pathways, functions potently in cell survival and apoptosis (Xia et al. 1995), and that the ERK pathway is linked closely to β-cell apoptosis and insulin secretion (Wijesekara et al. 2010, Yeo et al. 2012). In this study, based on the effects of BBT in both potentiating GSIS and protecting β-cells, we thus wondered whether ERK regulation was also involved in such BBT-induced events. Accordingly, we carried out related assays in INS-832/13 cells.

BBT-stimulated ERK1/2 phosphorylation In the investigation of the effect of glucose on the BBT-activated ERK1/2 phosphorylation, the cells were incubated for 2 h in glucose-free KRB buffer, followed by incubation with various concentrations of glucose in the presence of BBT (20 μM) for 2 h. As indicated in Fig. 2A, 2.8 mM glucose failed to activate ERK1/2 phosphorylation, and BBT had no effects on ERK1/2 activation with 2.8 mM glucose, while BBT induced a sustained increase in ERK1/2 phosphorylation in the presence of 5.6, 11.2, and 16.8 mM glucose. In addition, as shown in Fig. 2B, BBT, as expected, stimulated ERK1/2 phosphorylation in the cells with 16.8 mM glucose in a dose-dependent manner.

Therefore, these results indicated that BBT stimulated ERK1/2 phosphorylation in both glucose- and dose-dependent manners, which is consistent with its regulation of GSIS (Fig. 1B and C).

The ERK1/2 pathway is involved in the BBT-potentiated GSIS To further examine the association of ERK1/2 regulation with the BBT-potentiated GSIS, we carried out assays in the cells, in which U0126 (a specific MEK/ERK1/2 inhibitor (Duncia et al. 1998)) was used. As shown in Fig. 2C and D, U0126 treatment with the cells in 16.8 mM glucose dose-dependently decreased the BBT-potentiated ERK1/2 phosphorylation, and
BBT-potentiated GSIS in INS-832/13 cells. The potential association of the cAMP/PKA pathway with the secretion (Renstrom et al. 2012) has been reported, besides the cAMP/PKA pathway, Ca$^{2+}$-influx-related signaling also functions potently in insulin secretion (Rustenbeck et al. 1997). Thus, we performed an assay for inspecting the potential involvement of PKA regulation in the BBT-potentiated GSIS and ERK1/2 phosphorylation, where INS-832/13 cells were pre-incubated with H89 (a specific PKA inhibitor (Chijiwa et al. 1990)) in glucose-free KRB buffer for 2 h, followed by stimulation with 16.8 mM glucose in the presence of BBT. As shown in Fig. 3D and E, H89 treatment potently decreased the BBT-potentiated GSIS and ERK1/2 phosphorylation.

Therefore, all the results indicated that the cAMP/PKA pathway is involved in the BBT-potentiated GSIS and ERK1/2 phosphorylation.

L-VDCC/CaMK2 is involved in the BBT-potentiated GSIS

Ca$^{2+}$ is essential for the BBT-potentiated GSIS

As demonstrated in Fig. 3A, BBT (20 µM) increased intracellular cAMP with 16.8 mM glucose. Accordingly, we determined whether cAMP was essential for the BBT-potentiated GSIS. In the assay, MDL-12,330A (an adenylyl cyclase (AC)-specific inhibitor (Seifert et al. 2012)) was used. As shown in Fig. 3B and C, MDL-12,330A (20 µM) treatment of cells in 16.8 mM glucose strongly decreased the BBT-potentiated GSIS and ERK1/2 phosphorylation. These results thus indicated that the cAMP pathway was involved in the BBT-potentiated GSIS and ERK1/2 activation.

Generally, the effects of cAMP in regulated exocytosis are considered to be mediated by the activation of cAMP-dependent PKA, a main cAMP target (Renstrom et al. 1997). Thus, we performed an assay for inspecting the potential involvement of PKA regulation in the BBT-potentiated GSIS and ERK1/2 phosphorylation, where INS-832/13 cells were pre-incubated with H89 (a specific PKA inhibitor (Chijiwa et al. 1990)) in glucose-free KRB buffer for 2 h, followed by stimulation with 16.8 mM glucose in the presence of BBT. As shown in Fig. 3D and E, H89 treatment potently decreased the BBT-potentiated GSIS and ERK1/2 phosphorylation.

Therefore, all the results indicated that the cAMP/PKA pathway is involved in the BBT-potentiated GSIS and ERK1/2 phosphorylation.
The results indicated that Ca\textsuperscript{2+} secretion, but BBT could reverse the thapsigargin-(1 or 10 \textmu M)-treatment totally inhibited the BBT-stimulated insulin secretion. As indicated in Fig. 4B, EGTA (0.5 mM) was applied (Quevedo et al. 1999). As demonstrated previously, endoplasmic reticulum Ca\textsuperscript{2+} release (Lytton et al. 1991) were applied. As indicated in Fig. 4B, EGTA (0.5 mM) treatment totally inhibited the BBT-stimulated insulin secretion, but BBT could reverse the thapsigargin-(1 or 10 \textmu M)-treatment-stimulated decrease in insulin secretion. The results indicated that Ca\textsuperscript{2+} was essential for the BBT-potentiated GSIS and that endoplasmic reticulum Ca\textsuperscript{2+} release was not the main factor for the BBT-potentiated GSIS.

Extracellular Ca\textsuperscript{2+} influx is mainly responsible for the BBT-potentiated GSIS As demonstrated previously, endoplasmic reticulum Ca\textsuperscript{2+} release was not the main factor in the BBT-potentiated GSIS. Therefore, we investigated the relationship between extracellular Ca\textsuperscript{2+} influx and BBT-potentiated GSIS. A specific L-VDCC blocker (nifedipine) was applied (Quevedo et al. 1998). As shown in Fig. 4C, nifedipine dose-dependently reversed the BBT-potentiated GSIS. This result thus indicated that L-VDCC was involved in the BBT-potentiated GSIS.

The K\textsubscript{ATP} channel is not involved in BBT-potentiated GSIS Given that the K\textsubscript{ATP} channel plays an important role in the glucose-induced Ca\textsuperscript{2+} influx (Shibasaki et al. 2004), we investigated the potential association of the K\textsubscript{ATP} channel with the BBT-potentiated GSIS. As shown in Fig. 4D, repaglinide (a specific K\textsubscript{ATP} channel blocker (Stephan et al. 2006)) alone significantly increased insulin secretion in the cells with 16.8 mM glucose. In view of the fact that repaglinide at 10 nM could inhibit the K\textsubscript{ATP} channel by almost 95% in pancreatic cells (Stephan et al. 2006), high concentrations of repaglinide at 1 or 10 \textmu M (Fig. 4D) were thus suggested to be obviously enough to totally blockade the K\textsubscript{ATP} channel as indicated by the result that the repaglinide-stimulated insulin secretion reached a peak at 1 and 10 \textmu M (Fig. 4D). Therefore, the finding that BBT could still enhance insulin secretion in the presence of repaglinide implied that BBT was not a direct blocker of the K\textsubscript{ATP} channel.

Next, diazoxide (an opener of the K\textsubscript{ATP} channel (Ritzel et al. 2004)) was thus used in an attempt to elucidate the effect of BBT downstream or upstream of the K\textsubscript{ATP} channel. As also shown in Fig. 4D, although diazoxide decreased the BBT-potentiated GSIS, BBT could still reverse the effect of diazoxide on GSIS, this result combined with above information from the experiments using the K\textsubscript{ATP} channel blocker indicated that the K\textsubscript{ATP} channel might not be involved in BBT-potentiated GSIS. This suggestion could be also supported by the assay examining the involvement of ERK1/2, where BBT obviously stimulated the diazoxide/repaglinide-regulated ERK1/2 phosphorylation (Fig. 4E), similar to the cases for diazoxide and repaglinide in the regulation of GSIS.

L-VDCC/CaMKII pathway is involved in the BBT-potentiated GSIS Given that L-VDCC is involved in

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Figure 3

cAMP/PKA pathway participated in the BBT-potentiated GSIS. (A) INS-832/13 cells were pre-incubated in glucose-free KRB buffer for 2 h, and then stimulated with 2.8 or 16.8 mM glucose in the presence or absence of BBT (20 \textmu M) for 2 h, and the cells were collected for determining intracellular cAMP levels. (B) INS-832/13 cells were pre-incubated in glucose-free KRB buffer with MDL-12,330A (20 \textmu M) for 2 h, and then stimulated with 2.8 or 16.8 mM glucose for 2 h in the presence of BBT (20 \textmu M) or BBT (20 \textmu M) with MDL-12,330A (20 \textmu M), the insulin concentration was determined by ELISA assay. (C) The cell lysate from (B) was analyzed by western blotting. (D) INS-832/13 cells were incubated in glucose-free KRB buffer with or without H89 (5, 10, and 20 \textmu M) for 2 h, and stimulated with 2.8 or 16.8 mM glucose for 2 h in the presence of BBT (20 \textmu M) or BBT (20 \textmu M) with H89 (5, 10, and 20 \textmu M). Insulin concentration was determined by ELISA assay. (E) The cell lysate in (D) was analyzed by western blotting. All data were obtained from three independent experiments and are presented as means ± S.E.M. (*P < 0.05, **P < 0.01, and ***P < 0.001).
Figure 4

L-VDCC/CaMK2 activation is involved in the BBT-potentiated GSIS. (A) Intracellular Ca\(^{2+}\) level in INS-832/13 cells was monitored using Fluo-4 AM fluorescence dye. The cells were incubated with 2.8 mM glucose for 16 s, followed by stimulation with 16.8 mM glucose in the presence of BBT (10 and 20 \(\mu\)M), and fluorescent signals were detected. (B) INS-832/13 cells were incubated in glucose-free KRB buffer with or without EGTA (0.5 mM) and thapsigargin (1 \(\mu\)M) for 2 h, and then incubated in 2.8 or 16.8 mM glucose in the presence or absence of BBT (20 \(\mu\)M), EGTA (0.5 mM), BBT with EGTA, thapsigargin (1 \(\mu\)M), BBT with thapsigargin (1 \(\mu\)M), thapsigargin (10 \(\mu\)M) or BBT with thapsigargin (10 \(\mu\)M) for 2 h, and insulin concentrations were determined by ELISA assay. (C) INS-832/13 cells were pre-incubated in glucose-free KRB buffer with or without nifedipine (5, 10, and 20 \(\mu\)M) for 2 h, and then stimulated with 2.8 or 16.8 mM glucose KRB buffer for 2 h in the presence of BBT (20 \(\mu\)M) or BBT with nifedipine (5, 10, and 20 \(\mu\)M), and for 2 h, insulin concentrations were determined by ELISA assay.

The BBT-potentiated GSIS, the potential action of BBT against the L-VDCC downstream Ca\(^{2+}\)-dependent protein kinases, including PKC, CaMK2, and CaMKK, were investigated in INS-832/13 cells.

In the assay, the cells were separately incubated with GFX (a PKC inhibitor (Yokota et al. 2003)), KN93 (a CaMK2 inhibitor (Sumi et al. 1991)), or STO-609 (a CaMKK inhibitor (Tokumitsu et al. 2003)) for 2 h in glucose-free KRB buffer, followed by stimulation with 16.8 mM glucose in the presence or absence of BBT for another 2 h. As shown in Fig. 4F, only KN93 dose-dependently blocked the BBT-potentiated GSIS, implying that CaMK2 activation might contribute to the BBT-potentiated GSIS. Moreover, INS-832/13 cells pretreated with STO-609 or GFX displayed greater increases in insulin secretion in the presence of BBT, this might be the result of the fact that extracellular the Ca\(^{2+}\) influx induced by BBT might activate its downstream kinases including PKC, CaMK2, and CaMKK, and inhibition of PKC or CaMKK activation could better focus the extracellular Ca\(^{2+}\) influx induced by BBT into the activation of CaMK2 and increase the insulin secretion. In addition, the results indicating that both
nifedipine and KN93 dose-dependently decreased the BBT-potentiated ERK1/2 phosphorylation further confirmed that L-VDCC and its downstream kinase CaMK2 were involved in the BBT-potentiated GSIS (Fig. 4G). Therefore, all results indicated that the L-VDCC/CaMK2 pathway is involved in the BBT-potentiated GSIS.

cAMP/PKA as an upstream pathway of L-VDCC/CaMK2 is responsible for the BBT-enhanced Ca\(^{2+}\) influx

As BBT has been determined to activate intracellular cAMP and Ca\(^{2+}\) influx, we next investigated the association of cAMP with the BBT-enhanced intracellular Ca\(^{2+}\) in INS-832/13 cells. In the assay, the cells were incubated with MDL-12,330A (20 \(\mu\)M) or H89 (20 \(\mu\)M) in glucose-free KRB buffer for 2 h, and then stimulated with 16.8 mM glucose in the presence or absence of BBT (20 \(\mu\)M). As shown in Fig. 5A, either MDL-12,330A or H89 decreased the enhancement of the intracellular level of Ca\(^{2+}\) induced by BBT, which indicated that cAMP/PKA was involved in the BBT-activated Ca\(^{2+}\) influx. It is noted that the effect of BBT in enhancing the high-glucose-(16.8 mM)-induced intracellular Ca\(^{2+}\) level is not so obvious compared with the result shown in Fig. 4A, this might be caused by the

![Figure 5](https://example.com/figure5.png)

**Figure 5**
cAMP/PKA as an upstream pathway of L-VDCC/CaMK2 is responsible for the BBT-enhanced Ca\(^{2+}\) influx. (A) Intracellular Ca\(^{2+}\) levels were monitored using Fluo-4 AM fluorescence dye in INS-832/13 cells. The cells were pre-incubated in glucose-free KRB buffer for 2 h and then stimulated with 2.8 mM glucose for 16 s, followed by stimulation with 16.8 mM glucose in the presence of BBT (20 \(\mu\)M), BBT with H89 (20 \(\mu\)M), or BBT with MDL-12,330A (20 \(\mu\)M), the fluorescent signal was detected. (B) INS-832/13 cells were incubated in glucose-free KRB buffer with or without nifedipine (20 \(\mu\)M) or KN93 (20 \(\mu\)M) for 2 h, and stimulated with 16.8 mM glucose for 2 h with BBT (20 \(\mu\)M), nifedipine (20 \(\mu\)M), BBT with nifedipine (20 \(\mu\)M), KN93 (20 \(\mu\)M), or BBT (20 \(\mu\)M) with KN93 (20 \(\mu\)M), the cells were collected to determine intracellular cAMP levels. (C) PDEs activity was determined by enzyme activity assay, and then the effects of BBT (20 \(\mu\)M) and 3-isobutyl-1-methylxanthine (IBMX) (50 \(\mu\)M) were detected. (D) 293T cells or (E) INS-832/13 cells were transfected with cAMP-responsive luciferase reporter plasmid (CRE-Luc, 300 ng/well) in 24-well plates and then the cells were treated with FSK (10 \(\mu\)M), exendin-4 (1 \(\mu\)M), or BBT (20 \(\mu\)M) for 18 h, and luciferase activity was determined. All data were obtained from three independent experiments and are presented as means±S.E.M. (**P<0.05, ***P<0.01, and ****P<0.001).
change in absolute live-cell numbers in different batches of experiments. Next, the role of L-VDCC or CaMK2 activity in the increase in intracellular cAMP due to BBT was examined. As shown in Fig. 5B, neither nifedipine nor KN93 affected cell viability. Therefore, these results indicated that cAMP/PKA as an upstream pathway of L-VDCC/CaMK2 is responsible for the increase in Ca\(^{2+}\) influx induced by BBT. Given that cAMP is regulated by PDEs, AC, and GPCRs (Szkudelski 2001), we also performed the relevant assays in an attempt to determine the potential targets of BBT. We found that IBMX as an inhibitor of PDEs decreased the activity of PDEs and that BBT had no effects on the activity of PDEs, which indicated that BBT was not an inhibitor of PDEs (Fig. 5C). Considering the similarity of AC catalytic domains to each other and that activator of AC could activate all types of AC, such as forskolin (FSK; Szkudelski 2001), cAMP-responsive luciferase reporter plasmid (CRE-Luc) was next used to study whether BBT targeted AC or GPCRs. In the assay, CRE-Luc plasmid was transfected into 293T-cells (Fig. 5D) or INS-832/13 cells (Fig. 5E). It was found that FSK (10 \(\mu\)M) enhanced the luciferase activity in both 293T and INS-832/13 cells, while BBT (20 \(\mu\)M) only enhanced the luciferase activity in INS-832/13 cells similarly to the GLP1 receptor agonist, exendin-4, which indicated that BBT enhanced intracellular cAMP through GPCR on \(\beta\)-cells. On the basis of its effects on cAMP, we speculated that BBT might act as a GPCR agonist coupled to Gs. Furthermore, we investigated whether BBT could active GLP1 receptor, GPR35 and GPR55, but,

![Figure 6](image-url)

**Figure 6**

The cAMP/PKA pathway is involved in BBT-induced \(\beta\)-cell protection. INS-832/13 cells were incubated with (A) STZ (0.4 mM) or (B) cytokines (10 ng/ml TNF\(\alpha\) and 5 ng/ml IL1B) in the presence of the indicated compounds for 24 h, and then cell viability was evaluated by MTT assay. (C) INS-832/13 cells were incubated with the kinase inhibitors (KN93, H89, or U0126), and then cell viability was evaluated by MTT assay. (D) INS-832/13 cells were incubated with STZ (0.4 mM) or cytokines (10 ng/ml TNF\(\alpha\) and 5 ng/ml IL1B) in the presence or absence of BBT (20 \(\mu\)M) for 24 h, and then mRNA level of CHOP was determined. (E) Protein level of CHOP was determined by western blotting. All data were obtained from three independent experiments and are presented as means \(\pm\) S.E.M. (**\(P<0.01\), and ***\(P<0.001\)).
Figure 7
BBT ameliorated glucose homeostasis in the STZ/HFD-induced type 2 diabetic mice. (A) Fasting serum insulin levels were detected in STZ/HFD-induced type 2 diabetic mice after BBT (50 mg/kg per day) treatment (n = 9). (B) Morphology (HE staining) and (C) insulin immunohistochemistry of pancreatic β-cells in chow-diet-fed, HFD-fed, or HFD-fed/STZ-induced mice were examined. (D) Quantification of insulin-positive islets in (C). (E) Fasting serum glucose level was detected in mice after BBT (50 mg/kg per day) or rosiglitazone (5 mg/kg) treatment (n = 9). (F) Serum HbA1c level in mice after BBT (50 mg/kg per day) treatment was determined. (G) OGTT was performed in mice after BBT (50 mg/kg per day) treatment (n = 9). (H) AUC result for OGTT in (G). (I) ITT was performed in mice after BBT (50 mg/kg per day) treatment (n = 9). (J) AUC result for ITT in (I). (K) Serum insulin concentration was determined during OGTT. All data are presented as means ± S.E.M. (*P < 0.05, **P < 0.01, and ***P < 0.001).
disappointingly, could not directly activate the above GPCRs responsible for insulin secretion (data not shown). Further research is ongoing.

The cAMP/PKA pathway is involved in the protection of BBT against β-cell

Given that cAMP/PKA, L-VDCC/CaMK2, and ERK1/2 pathways have been determined to be involved in the BBT-potentiated GSIS, we next inspected the potential mechanism involving BBT in the protection of β-cells. In the assay, INS-832/13 cells were incubated with KN93 (H89 or U0126) and STZ or cytokines in the presence or absence of BBT, and β-cell viability was then detected. As shown in Fig. 6A and B, in either of the two models with STZ- or cytokine-induced β-cell death, KN93 treatment did not affect the protective action of BBT on β-cells, but H89 and U0126 efficiently deprived the effect of BBT on β-cell protection. Notably, BBT itself has no effects on cell viability and none of KN93, H89, and U0126 exhibited any effects on cell viability by themselves (Fig. 6C). Therefore, the influences of these kinase inhibitors on BBT thereby implied that cAMP/PKA and ERK1/2 not the L-VDCC/CaMK2 pathway was involved in the protective effect of BBT against β-cell death.

Given the possibility that upregulation of ERK may suppress CHOP expression thereby protecting β-cells against apoptosis, we also carried out the relevant assays. As demonstrated in Fig. 6D and E, BBT had no effects on the mRNA or protein level of CHOP, which indicated that CHOP was not involved in BBT events.

Therefore, all results indicated that the cAMP/PKA/ERK1/2 pathway is involved in the protective action of BBT on β-cells.

BBT ameliorated glucose homeostasis in STZ/HFD-induced type 2 diabetic mice

Next, we explored the potential effect of BBT on glucose homeostasis in STZ/HFD-induced type 2 diabetic mice. The type 2 diabetic model in our study was also at first confirmed using rosiglitazone, the generally accepted reagent for type 2 diabetes but not type 1 diabetes (Gastaldelli 2011). The finding that rosiglitazone (5 mg/kg) significantly decreased the fasting glucose level of STZ/HFD-induced diabetic mice (Fig. 7E) clearly confirmed the type 2 diabetic features of the model. As shown in Fig. 7E and F, chronic administration of BBT for 4 weeks efficiently reduced the fasting serum glucose and HbA1c levels in the mice. In addition, the results of the OGTT assay indicated that BBT treatment was effective in improving the glucose tolerance in HFD/STZ-induced type 2 diabetic mice (Fig. 7G and H), and that BBT had no effects on the ITT assay (Fig. 7I and J). Furthermore, BBT treatment effectively increased the serum insulin concentration during the OGTT (Fig. 7K), which thus strengthened support for the insulinoicotropic effect of BBT in vivo.

Discussion

Type 2 diabetes is a chronic metabolic disease, which has brought much burden to societies (Zimmet et al. 2001). Although the underlying pathological basis of type 2 diabetes is not clear, insulin resistance and β-cell dysfunction contribute much to this heterogeneous disease (Gastaldelli 2011). Recently, amelioration of β-cell dysfunction by activation of GSIS and/or prevention of β-cell death have been determined to be promising approaches in delaying or preventing the onset of type 2 diabetes (Rhodes 2005, Prentki & Nolan 2006, Seino et al. 2011, Rachdi et al. 2014). In this work, we found that the
thiophenecarboxamide derivative BBT was effective in both enhancing GSIS and alleviating β-cell death, and the results of the in vivo assay further demonstrated its capabilities in the improvement of glucose homeostasis in type 2 diabetic mice. These results thus highlighted the potential of BBT in anti-diabetic research.

ERK1/2 has been implicated in many cellular events, including proliferation, differentiation, survival, and secretion (Volmat & Pouyssegur 2001). In β-cells, the glucose-activated ERK1/2 regulated insulin gene transcription and insulin secretion (Benes et al. 1999). Although the role of ERK1/2 in the progress of insulin secretion was controversial (Khoo & Cobb 1997, Longuet et al. 2005), ERK1/2 was activated by insulinotropic agents. Interestingly, sulfonylureas and FSK activated ERK1/2 phosphorylation independently from glucose (Khoo & Cobb 1997, Arnette et al. 2003), which was different from the cases for GLP1 and its analogs in glucose-dependently stimulating ERK1/2 phosphorylation. In our current work, BBT-potentiated GSIS and the glucose-induced ERK1/2 phosphorylation, demonstrating the importance of glucose concentration for BBT function. Therefore, our results further indicated that BBT may get rid of the hypoglycemic effect under normal conditions, which is a serious clinical side effect for many insulinotrophic drugs with glucose-independent stimulatory effects on insulin secretion (de Heer & Holst 2007).

cAMP/PKA and intracellular Ca\(^{2+}\) have both been reported to mediate ERK1/2 phosphorylation and GSIS in β-cells (Wang et al. 1993, Briaud et al. 2003), and enhancement of cytoplasmic Ca\(^{2+}\) led to cAMP production and subsequent PKA activation (Briaud et al. 2003), while elevation of cAMP level also regulated the entry of Ca\(^{2+}\) into β-cells through L-VDCC to induce insulin secretion (Wang et al. 1993). GLP1 activated ERK1/2 and GSIS through activation of PKA and CaMK2 (Gomez et al. 2002). Herein, BBT enhanced the intracellular cAMP level, and then induced Ca\(^{2+}\) influx through L-VDCC and CaMK2 activation, and finally promoted ERK1/2 phosphorylation and insulin secretion (Fig. 8).

In pancreatic β-cells, cAMP/PKA activation has been reported to reduce apoptosis and stimulate cell survival (Huì et al. 2003), and the finding that BBT protected β-cells through activation of cAMP/PKA was consistent with the published result. Notably, the role of CaMK2 in β-cell protection was not clear, our results indicated that CaMK2 was not involved in the BBT-mediated β-cell protection although CaMK2 was activated by BBT. Furthermore, the finding that BBT could reverse the STZ- or cytokine-induced insulin secretion deficiency indicated that BBT may have potential in improving insulin secretory deficiency (Leahy et al. 1992).

As has been reported, thiophenecarboxamide-type compounds have effects on many pharmacological events, such as anti-inflammation (Baxter et al. 2004), cell cycle regulation (Janetka et al. 2008), antifungal actions, DNA damage inhibition (Shinkwin et al. 1999), and inhibition of Ca\(^{2+}\)-release-activated calcium channels (Yonetoku et al. 2006), and no reports have been published on their anti-diabetic effects. Therefore, our current report that the thiophenecarboxamide derivative BBT efficiently improved β-cell dysfunction is expected to have expanded the pharmacological understanding of thiophenecarboxamide compounds.

In conclusion, the small-molecule thiophenecarboxamide derivative BBT was discovered to potentiate GSIS, protect β-cells, and effectively improve glucose homeostasis in type 2 diabetic mice. Both cAMP/PKA/CaMK2 and ERK1/2 pathways were involved in the BBT-promoted GSIS, and the cAMP/PKA pathway was essential for the protective effect of BBT on β-cells. Our current work has highlighted the potential of BBT in anti-hyperglycemia research.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding
This work was supported by the National Natural Science Foundation of China (grant numbers 81373461, 81473141, 91413102, 81220108025, and 91213306).


**Received in final form 24 December 2014**

**Accepted 8 January 2015**

**Accepted Preprint published online 8 January 2015**