Sub-erythemal ultraviolet radiation reduces metabolic dysfunction in already overweight mice

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Abstract

Exposure to sunlight may limit cardiometabolic risk. In our previous studies, regular exposure to sub-erythemal (non-burning) ultraviolet radiation (UVR) reduced signs of adiposity and cardiometabolic dysfunction in mice fed a high-fat diet. Some of the observed effects were dependent on skin release of nitric oxide after UVR exposure. Here, we examine the effects of sub-erythemal UVR on signs of adiposity and metabolic dysfunction in already overweight mice, comparing the effects of two sunlamps with distinct emitted light spectra. Mice were fed a high-fat diet from 8 weeks of age, with UVR administered twice a week from 14 weeks of age until they were killed at 20 weeks of age. Mice were irradiated with the same dose of UVB radiation (1 kJ/m²) from either FS40 (65% UVB, 35% UVA) or CLEO (4% UVB, 96% UVA) sunlamps, but substantially more UVA from the latter. FS40 UVR (but not CLEO UVR) significantly reduced mouse weights and weight gain, compared to mice fed a high-fat diet (only). These effects were dependent on nitric oxide. Conversely, CLEO UVR (but not FS40 UVR) significantly reduced circulating LDL cholesterol. Both light sources reduced fasting insulin levels, and the extent of hepatic steatosis; the latter was reversed by topical application of cPTIO, suggesting an important role for skin release of nitric oxide in preventing hepatic lipid accumulation. These results suggest that there may be a number of benefits achieved by regular exposure to safe (non-burning) levels of sunlight or UV-containing phototherapy, with effects potentially dependent on the predominance of the wavelengths of UVR administered.

Introduction

Controlling the development of obesity and its comorbidities like metabolic syndrome and type-2 diabetes is now the centerpiece of many government health strategies around the world. Although there is ongoing focus on energy-rich diets and insufficient exercise, there is an underappreciated potential for inadequate exposure to sunlight as an additional lifestyle modifier. There is evidence from human and preclinical
studies that increased exposure to sunlight or ultraviolet radiation (UVR) may prevent the development of obesity and metabolic dysfunction.

In temperate climates that experience seasonal variation in ambient UVR levels, reduced blood pressure (Kunes et al. 1991, Woodhouse et al. 1993), heart failure, thromboembolic events and stroke have been observed in summer (reviewed in Zittermann & Gummert 2010). Winter increases in body fat and plasma Hba1c have been reported in type 2 diabetics (Sohmiya et al. 2014). Other studies report that the incidence of type 2 diabetes and fasting glucose levels are lowest in summer (Doro et al. 2008). Positive latitude (distance from the equator) gradients, a surrogate for reduced sun exposure, have been reported for hypertension (Rostand 1997) and cardiovascular-related mortality (Baldassarre et al. 2010). Some studies also report a reduced risk of diabetes or obesity in people living at higher altitudes (reviewed in Hirschler 2016), where terrestrial UVR radiation levels are greater (Holick et al. 2007). Not all evidence from studies of season, latitude and altitude point to a protective effect of increased sun exposure on cardiometabolic disease risk (Rostand 1997, Shore-Lorenti et al. 2014, Hirschler 2016).

Other human studies have found inverse associations of obesity and outcomes of excessive sun exposure like skin cancer, even after adjusting for physical activity (Pothiawala et al. 2012, Tang et al. 2013). However, increased systolic blood pressure and risk of diabetes was observed in Korean adults obtaining >5h/day of sun exposure, who were older, more likely to be smokers and drink alcohol and less likely to have a college education (Ohn et al. 2014). In other epidemiological studies, women with active sunbathing habits or who used sun beds had reduced risk of type 2 diabetes (Lindqvist et al. 2010), thromboembolic events (Lindqvist et al. 2009) and all-cause mortality (Lindqvist et al. 2014) after adjusting for exercise and other confounders. Exposure to sub-erythemal UVA radiation reduced blood pressure in normotensive young (Liu et al. 2014) but not older adults (Krause et al. 1998, Scragg et al. 2011). The anti-hypertensive effects of UVA radiation may have been dependent on the release of nitric oxide from preformed skin stores (Liu et al. 2014). Two weeks of whole body treatments with erythemal UVB radiation (4 times in total) increased insulin secretion in healthy adults challenged with glucagon (Colas et al. 1989).

Vitamin D status can be used as a proxy for sun exposure. Skin exposure to UVB radiation results in dermal synthesis of vitamin D, and further hydroxylation events in the liver increase circulating 25-hydroxyvitamin D (25(OH)D). Vitamin D deficiency has been proposed as a risk factor for obesity and type 2 diabetes (Earthman et al. 2012). Serum 25(OH)D levels are reduced in obesity (Autier et al. 2014), but clinical trials have failed to conclusively show that vitamin D supplementation reduces weight gain (Mallard et al. 2016), type 2 diabetes or cardiovascular disease risk (Autier et al. 2014). The lack of success of these trials may be attributable to factors around study design (e.g. small sample size), the initial vitamin D status of participants (e.g. not being vitamin D-deficient at the start of the trial) and the amount and timing of vitamin D supplementation; however, the biological activity of non-vitamin D sun-induced mediators like nitric oxide (Feelisch et al. 2014, Liu et al. 2014, Fleury et al. 2016) may also explain the lack of effects observed.

We previously reported a protective effect of ongoing exposure to sub-erythemal UVR in controlling the development of signs of obesity and type 2 diabetes in C57Bl/6 mice fed a high-fat diet (Geldenhuys et al. 2014). UV-irradiated mice had reduced weight gain and diminished metabolic dysfunction including decreased fasting glucose and insulin levels, improved glucose tolerance, reduced insulin resistance and less liver steatosis (Geldenhuys et al. 2014). The beneficial effects of UVR on fasting glucose levels and liver steatosis were at least partially dependent on skin release of nitric oxide (Geldenhuys et al. 2014). We exposed mice to sub-erythemal UVR twice a week, from a source (FS40 sunlamps) that mainly emitted UVB radiation, for the 12 weeks from when mice first started eating the high-fat diet (Geldenhuys et al. 2014). In the study described below, we tested exposure to UVR as a potential way to limit the progression of overweight to obesity. We concentrated on sub-erythemal (non-burning) UVR, which is of low risk for skin cancer development and therefore could more easily be translated into policy or therapy. We compared the effects of the FS40 sunlamps (~65% UVB), with a light source that emits radiation that more closely mimics sunlight (Cleo sunlamps, 4% UVB (de Winter et al. 2001, Narbutt et al. 2005)). Finally, we examined a possible role for nitric oxide in mediating the effects of UVR.

Materials and methods

Mice

All experiments were performed according to the ethical guidelines of the National Health and Medical Research Council of Australia and with approval from the Telethon
Kids Institute Animal Ethics Committee. C57BL/6J(ARC) male mice were purchased from the Animal Resources Centre, Western Australia. The temperature (21°C) and lighting conditions (12-h light/darkness cycle) in the animal facility were controlled. Mice were housed under Perspex-filtered fluorescent lighting, which emitted no detectable ultraviolet (UV) B radiation as measured using a UV radiometer (UVX Digital Radiometer, Ultraviolet Products Inc., Upland, CA, USA). Mice were allowed access to food and acidified water ad libitum.

Diet

All diets were obtained from Specialty Feeds (Glen Forrest, Western Australia). The contents of these semi-pure low- (5% fat; canola oil) and high-fat (23%; lard (20.7%)) diets are described in Supplementary Table 1 (see section on supplementary data given at the end of this article). Neither diet was supplemented with dietary vitamin D, as the effects of UVR were reduced by this treatment in previous studies (Geldenhuys et al. 2014). Serum 25-hydroxyvitamin D levels in response to irradiation of C57BL/6J male mice to the same frequency (twice a week) and dose (1 kJ/m²) of UVB radiation from the FS40 sunlamps (as that used here, see below) have previously been reported in detail (Geldenhuys et al. 2014). All mice were fed the low-fat diet from 4 until 8 weeks of age, and one group was fed the low-fat diet until the end of the experiment. All other mice were fed the high-fat diet from 8 weeks of age for 12 weeks until mice were 20 weeks of age (Fig. 1).

Figure 1

The experimental approach. Four-week-old C57BL/6J male mice were fed a low-fat diet (LFD) for four weeks. At eight weeks of age, mice were either continued on this diet (treatment 1) or switched to a high-fat diet (HFD). After 6 weeks of feeding, mice fed the HFD were exposed to one of 5 treatments for another 6 weeks. The shaved dorsal skin of these mice were treated twice a week with (2) vehicle only and mock-irradiation (Mock UVR), (3) sub-erythemal FS40 UVR (1 kJ/m² UVB) and then vehicle (FS40 UVR), (4) topical SNAP (SNAP), (5) sub-erythemal FS40 UVR (1 kJ/m² UVB) and then topical cPTIO (FS40 UVR + cPTIO) or (6) CLEO UVR (1 kJ/m² UVB; CLEO UVR). The first treatment group of mice (1) were fed a low-fat diet and were administered the vehicle and mock-irradiated twice a week. Mice were treated for 6 weeks with these skin treatments until 20 weeks of age. There were a total of 6 treatments, with 18 mice per treatment. The first treatment group of mice (1) were fed a low-fat diet and were administered the vehicle and mock-irradiated twice a week. Mice were treated for 6 weeks with these skin treatments until 20 weeks of age. There were a total of 6 treatments, with 18 mice per treatment. The experiment was performed twice with results combined for both experiments. nb. One mouse was killed from the CLEO UVR treatment (at week 4), and another from the FS40 UVR + cPTIO treatment (at week 10) due to the development of severe dermatitis, which did not resolve, reducing the total number of animals to 17 in these two groups.
UV radiation and topical skin treatments

Two sources of UVR were used. The first was a bank of 40W FS40 lamps (Philips TL UV-B, Eindhoven, The Netherlands) emitting broadband UVR, 250–360 nm, with 65% of the output in the UVB range (280–315 nm), and the remaining UVR in the UVC (250–280 nm) and UVA (315–360 nm) ranges. The second was a bank of 100W Cleo Natural lamps (Philips) emitting light, which more closely mimics the terrestrial spectrum of solar radiation, with the spectral UVR bandwidth composed of 4% UVB and 96% UVA (de Winter et al. 2001, Narbutt et al. 2005). Clean-shaven dorsal skin (8 cm²) was exposed to light emitted from either the FS40 or CLEO sunlamps as previously described using PVC plastic to block wavelengths less than 280 nm (UVC radiation) (Gorman et al. 2007, Ng et al. 2013, Geldenhuys et al. 2014). Mice exposed to either light source were irradiated with the same dose of sub-erythemal UVB radiation (1 kJ/m²; (McGlade et al. 2007, Geldenhuys et al. 2014)) as determined using a handheld ultraviolet radiometer (UVX Digital Radiometer; Ultraviolet Products Inc.). The amount of UVA radiation delivered differed, with mice receiving an estimated dose of 0.5 or 24 kJ/m² of UVA radiation, when exposed to the FS40 or CLEO lamps, respectively. For other treatments, as previously described (Geldenhuys et al. 2014), dorsal skin was treated with 0.1 mmol SNAP (S-nitroso-N-acetyl-d,l-penicillamine, Sigma (Ikeyama et al. 2007)), a nitric oxide donor or a nitric oxide scavenger, cPTIO (carboxy-PTIO potassium salt, Sigma (Yasukawa et al. 2012), 0.1 mmol) immediately following delivery of FS40 UVR. For mock treatments, the dorsal skin of mice was shaved, and mice then placed in the same Perspex box (under standard fluorescent lighting) for the same amount of time used to irradiate other mice. After 6 weeks of feeding mice the high-fat diet (Fig. 1), one of five of the following skin treatments was administered twice a week to the shaved dorsal skin of mice: (1) vehicle and mock-irradiation (Mock UVR), (2) sub-erythemal FS40 UVR (1 kJ/m² UVB) and then vehicle (FS40 UVR); (3) topical SNAP (1 mM, SNAP); (4) sub-erythemal FS40 UVR (1 kJ/m² UVA) and then topical cPTIO (FS40 UVR+cPTIO) or (5) CLEO UVR (1 kJ/m² UBV; CLEO UVR) (Fig. 1). The SNAP-treated mice were also mock-irradiated. A final group of mice were fed the low-fat diet and treated with vehicle and mock-irradiated twice a week.

Measuring weight gain and tissue weights

Mice were weighed weekly on the same day in the morning using a digital scale (Ohaus Scout, >0.1 g resolution).

Percentage weight gain was calculated from 8 weeks of age. At the conclusion of the experiment, liver and gonadal deposits of white adipose tissue and interscapular deposits of brown adipose tissue were dissected from mice and their weights were determined using an analytical scale (Analytical Standard Electric Balance, Ohaus, NJ, USA; >0.0001 g resolution).

Glucose tolerance tests (GTT)

As previously described (Geldenhuys et al. 2014), but briefly, mice were fasted for 5 h before injected intraperitoneally with 1 g/kg glucose (Phebra, Lane Cove, NSW, Australia), with glucose levels serially monitored before and after the glucose challenge using a Accu-Chek Performa glucometer (Roche).

Serum metabolites

Serum cholesterol, HDL cholesterol, LDL cholesterol and triglyceride and fasting insulin, adiponectin and leptin were measured as previously described (Geldenhuys et al. 2014). Serum levels of activated aspartate aminotransferase (AST) levels were measured at PathWest Pathology, using the Clinical Chemistry kit as part of the Architect c System (Abbot Laboratories).

Histopathological assessment of liver pathology

The severity of non-alcoholic fatty liver disease (NAFLD) was assessed by grading formalin-fixed and H&E-stained liver sections as previously described (Geldenhuys et al. 2014), with the extent of fibrosis scored in Masson’s trichrome-stained sections (0 = none, 1 = fibrosis in some portal areas, 2 = fibrosis in most portal areas, 3 = fibrosis in most portal areas with portal to central bridging) (Kleiner et al. 2005). Steatosis and ballooning scores were added together for an overall steatosis score (≤6, (Geldenhuys et al. 2014)), and fibrosis scores were added to these for a combined histopathology score (≤10).

Statistical analyses

The experiment was performed twice, with n = 18 mice per treatment. Initial power calculations indicated that 36 mice per group would be required to observe a significant reduction of ≥35% in weight gain (power ≥0.8, P < 0.05; G*Power v3.1.3, 2009, based upon previously published data (Geldenhuys et al. 2014)). However, this number of
mice per group exceeded the logistical capacity of our research team. We expected that using an inbred strain under identical maintenance conditions would produce comparable results, and so, split this large experiment into two of equal size. Within the controlled confines of a single animal house, we controlled for the potential effects of seasonality (in particular temperature and lighting fluctuations), important factors in studying the effects of UVR on biological responses. Experiment 1 commenced in February 2015 and was completed in June 2015, whereas experiment 2 commenced in April 2015 and was completed in August 2015. The combined results from both experiments were compared using an analysis of variance (ANOVA) comparing between treatments using a Tukey post hoc analysis, which corrects for multiple comparisons. Area under the curve (AUC) was calculated for GTT using GraphPad Prism (v5) using 0 as the baseline. Results were considered as statistically significant for $P$ values <0.05.

**Results**

A high-fat diet induced signs of overweight and metabolic dysfunction after 6 weeks of feeding

C57Bl/6J male mice were fed a high- or low-fat diet from 8 weeks of age onward (Fig. 1). Mice fed the high-fat diet weighed more (Fig. 2A) and had increased weight gain (Fig. 2B) than those fed the low-fat diet after 1–2 weeks of feeding, weighing considerably more after 6 weeks. A GTT was performed after 5 weeks. Impaired glucose tolerance was observed in mice fed the high-fat diet with increased blood glucose levels observed at various times after glucose challenge (Fig. 2C), and increased AUC (GTT, Fig. 2D) observed in mice fed a high-fat diet.

**FS40 UVR but not CLEO UVR reduced weights and weight gain in mice fed a high-fat diet**

After 6 weeks of feeding mice the high-fat diet, one of five skin treatments was administered twice a week to the shaved dorsal skin of mice as described in Fig. 1. Another group of mice were fed the low-fat diet and treated with vehicle and mock-irradiated twice a week. All mice fed the high-fat diet weighed more (Fig. 3A and C) and gained more weight (Fig. 3B and D) than those fed the low-fat diet after 84 days (12 weeks) of feeding. Mice treated with FS40 UVR (but not CLEO UVR) had significantly reduced weights and weight gain compared to mice fed the high-fat diet after 6 weeks of treatment (Fig. 3). There was no difference in weights or weight gain after topical application of the nitric oxide donor SNAP (feeding mice a high-fat diet), compared to mice fed a high-fat diet (and mock treated) (Fig. 3). Topical treatment with the nitric oxide scavenger, cPTIO, prevented the suppressive effects of FS40 UVR on body weights and weight gain (Fig. 3).
UVR reduced fasting insulin levels in mice fed a high-fat diet

Mice fed the low-fat diet had significantly reduced fasting insulin and leptin levels in comparison to mice fed the high-fat diet only (mock UVR+vehicle) (Table 1) after 3 weeks of the skin interventions (or after 9 weeks of being fed a high-fat diet). Reduced fasting insulin levels were observed in mice exposed to UVR from either sunlamp (FS40 or CLEO), compared to those in mice fed a high-fat diet only (Table 1). However, there was no difference between fasting insulin levels observed in mice exposed to FS40 UVR, with or without cPTIO treatment (Table 1). There was no difference in adiponectin levels measured in mice from any treatment (Table 1). Mice fed the low-fat diet had significantly improved glucose tolerance (as measured by GTT) compared to mice fed the high-fat diet only (mock UVR+vehicle) (Table 1) after 4 weeks of the skin interventions (or after 10 weeks of being fed a high-fat diet). However, none of the UVR and/or topical treatments affected the extent of glucose intolerance observed or fasting glucose levels when comparing results observed in mice fed the high-fat diet only (Table 1).

Table 1  Expression levels of fasting insulin, leptin, adiponectin and glucose, and area under the curve values for glucose tolerance tests (GTT) measured 9–11 weeks after mice were initially fed the high-fat diet.

<table>
<thead>
<tr>
<th>Diet</th>
<th>UVR and/or skin treatment</th>
<th>Fasting insulin (ng/mL)</th>
<th>Fasting leptin (ng/mL)</th>
<th>Fasting adiponectin (ng/mL)</th>
<th>Fasting glucose (mM)</th>
<th>GTT (AUC, %basal glucose)</th>
</tr>
</thead>
<tbody>
<tr>
<td>LFD</td>
<td>Mock UVR + vehicle</td>
<td>0.5 ± 0.0*</td>
<td>1.0 ± 0.4*</td>
<td>10.2 ± 3.5</td>
<td>10.1 ± 0.4</td>
<td>1550 ± 42*</td>
</tr>
<tr>
<td>HFD</td>
<td>Mock UVR + vehicle</td>
<td>4.1 ± 1.2†</td>
<td>15.2 ± 2.7†</td>
<td>15.7 ± 3.3</td>
<td>10.1 ± 0.3</td>
<td>1973 ± 62†</td>
</tr>
<tr>
<td>FS40 UVR</td>
<td>Mock UVR + vehicle</td>
<td>12.0 ± 2.0*</td>
<td>11.9 ± 2.3†</td>
<td>14.5 ± 4.5</td>
<td>8.8 ± 0.4</td>
<td>1857 ± 81†</td>
</tr>
<tr>
<td>SNAP</td>
<td>Mock UVR + vehicle</td>
<td>1.8 ± 0.4</td>
<td>8.8 ± 1.5†</td>
<td>13.0 ± 3.7</td>
<td>9.1 ± 0.4</td>
<td>1915 ± 65†</td>
</tr>
<tr>
<td>FS40 UVR</td>
<td>Mock UVR + vehicle</td>
<td>1.6 ± 0.4</td>
<td>13.8 ± 3.9†</td>
<td>10.7 ± 3.7</td>
<td>10.0 ± 0.4</td>
<td>2103 ± 122†</td>
</tr>
<tr>
<td>CLEO UVR</td>
<td>Mock UVR + vehicle</td>
<td>1.2 ± 0.2*</td>
<td>15.0 ± 2.9†</td>
<td>23.8 ± 9.3</td>
<td>9.5 ± 0.3</td>
<td>1774 ± 42</td>
</tr>
</tbody>
</table>

*P < 0.05 relative to HFD (mock UVR+vehicle), one-way ANOVA (Tukey post hoc analysis); †P < 0.05 relative to LFD (mock UVR+vehicle), one-way ANOVA (Tukey post hoc analysis).

AUC, area under curve; cPTIO, carboxy-PTIO; HFD, high-fat diet; LFD, low-fat diet; SNAP, S-nitroso-N-acetyl-L:-penicillamine; UVR, ultraviolet radiation.
Table 2  Circulating lipid levels and adipose tissue weights at the end of the experiment.

<table>
<thead>
<tr>
<th>Diet</th>
<th>UVR and/or skin treatment</th>
<th>LDL cholesterol (mM)</th>
<th>HDL cholesterol (mM)</th>
<th>Total-cholesterol (mM)</th>
<th>Triglyceride (mM)</th>
<th>Gonadal WAT (g)</th>
<th>Inter-scapular BAT (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>LFD Mock UVR + vehicle</td>
<td><strong>0.13 ± 0.01</strong></td>
<td>1.5 ± 0.1</td>
<td>2.2 ± 0.2</td>
<td>0.9 ± 0.1</td>
<td><strong>0.6 ± 0.0</strong></td>
<td>0.15 ± 0.01</td>
</tr>
<tr>
<td>2</td>
<td>HFD Mock UVR + vehicle</td>
<td>0.22 ± 0.01†</td>
<td>1.7 ± 0.1</td>
<td>3.0 ± 0.2</td>
<td>1.1 ± 0.1</td>
<td>1.4 ± 0.1†</td>
<td>0.20 ± 0.02</td>
</tr>
<tr>
<td>3</td>
<td>HFD FS40 UVR + vehicle</td>
<td>0.20 ± 0.02†</td>
<td>1.7 ± 0.2</td>
<td>3.2 ± 0.3†</td>
<td>0.8 ± 0.1</td>
<td>1.1 ± 0.1†</td>
<td>0.15 ± 0.01</td>
</tr>
<tr>
<td>4</td>
<td>HFD SNAP</td>
<td>0.19 ± 0.01†</td>
<td>1.7 ± 0.1</td>
<td>3.1 ± 0.2†</td>
<td>1.1 ± 0.2</td>
<td>1.5 ± 0.1†</td>
<td>0.18 ± 0.01</td>
</tr>
<tr>
<td>5</td>
<td>HFD FS40 UVR+cPTIO</td>
<td>0.18 ± 0.02</td>
<td>1.7 ± 0.1</td>
<td>3.1 ± 0.1</td>
<td>0.9 ± 0.0</td>
<td>1.3 ± 0.1†</td>
<td>0.17 ± 0.02</td>
</tr>
<tr>
<td>6</td>
<td>HFD CLEO UVR + vehicle</td>
<td><strong>0.12 ± 0.01</strong></td>
<td>1.6 ± 0.1</td>
<td>2.6 ± 0.1</td>
<td>0.8 ± 0.1</td>
<td>1.2 ± 0.1†</td>
<td><strong>0.14 ± 0.01</strong></td>
</tr>
<tr>
<td>n/treatment</td>
<td></td>
<td>14–17</td>
<td>14–17</td>
<td>28–31</td>
<td></td>
<td>14–17</td>
<td>28–31</td>
</tr>
</tbody>
</table>

*P < 0.05 relative to HFD (mock UVR + vehicle), one-way ANOVA (Tukey post hoc analysis); †P < 0.05 relative to LFD (mock UVR + vehicle), one-way ANOVA (Tukey post hoc analysis).

Table 3  Liver histology observations at the end of the experiment.

<table>
<thead>
<tr>
<th>Diet</th>
<th>Liver function enzyme, aspartate aminotransferase (AST) (IU/L)</th>
<th>Triglyceride (mM)</th>
<th>Total-cholesterol (mM)</th>
<th>HDL cholesterol (mM)</th>
<th>LDL cholesterol (mM)</th>
<th>Hepatic steatosis</th>
<th>Fatty liver index</th>
<th>Collagen content</th>
<th>Liver weight (g)</th>
</tr>
</thead>
</table>

diet treatments compared to other treatment groups (Table 2). Weights of gonadal WAT were reduced in mice fed the low-fat diet, compared to high-fat diet treatments, with no significant reduction observed in mice exposed to UVR from either source (compared to the high-fat diet only treatment) (Table 2). Weights of interscapular BAT levels were significantly reduced in mice administered CLEO UVR (and fed a high-fat diet), compared to mice fed a high-fat diet only (Table 2). There was no effect of any of the skin treatments on serum triglyceride or HDL cholesterol or total cholesterol levels (Table 2).

**UVR reduced liver steatosis in mice fed a high-fat diet**

At the end of each experiment (or after 6 weeks of UVR intervention), significant steatosis and mild fibrosis were observed in livers of mice fed a high-fat diet (Fig. 4 and Table 3). Unexpectedly, there was significant steatosis in mice fed the low-fat diet, even though liver weights were significantly lower than those observed in mice fed a high-fat diet only (Table 3). Mice fed a high-fat diet and also exposed to FS40 or CLEO UVR or topically treated with the nitric oxide donor (SNAP) had significantly reduced liver steatosis compared to mice fed a high-fat diet only (Fig. 4 and Table 3). The effects of FS40 UVR on liver steatosis were reversed by immediate treatment with the nitric oxide donor cPTIO (Table 3, P < 0.05). In addition, there was some evidence (a trend; t test, P = 0.06) that the FS40 UVR treatment reduced circulating levels of the liver function enzyme, aspartate aminotransferase (AST) when compared to levels observed in mice fed the high-fat diet only (Table 3). Increased AST levels are generally

Figure 4  Representative liver histology sections stained with Masson's trichrome. The experiment is described in detail in Fig. 1. Data are representative of 2 experiments, with red arrows used to indicate steatosis, blue arrows for ballooning and green arrow for fibrosis for livers obtained at the 12-week endpoint.
regarded as an initial sign of liver dysfunction and may be used as part of a diagnosis of NAFLD (Clark 2006). There was less evidence for reduced AST levels ($P=0.2$) in mice treated with UVR and cPTIO, compared to those in mice treated with UVR and vehicle (Table 3). Altogether, these results suggest that regular exposure to low-dose UVR may reduce signs of NAFLD in mice fed a high-fat diet through mechanisms, which may be partially dependent on nitric oxide.

**Discussion**

Here, we observed the beneficial effects of ongoing exposure to low-dose (sub-erythmal) UVR as an intervention to reduce the severity of metabolic dysfunction in overweight mice fed a high-fat diet. Both sources of UVR reduced fasting insulin levels and the extent of liver steatosis. Applying the nitric oxide scavenger cPTIO to skin prevented the beneficial effects of FS40 UVR on liver steatosis but not fasting insulin, suggesting that different mediator(s) induced by UVR may be responsible. Furthermore, CLEO UVR (or SNAP) had an additional benefit of reducing circulating LDL cholesterol levels. Those mice treated with UVR from the CLEO lamps were exposed to $\geq 10$-fold more UVA radiation (than emitted by FS40 sunlamps), suggestive of a suppressive role for UVA-induced mediators in curbing serum LDL cholesterol levels. UVR from FS40 (but not CLEO) sunlamps reduced body weight and weight gain in mice fed a high-fat diet. These effects were reversed by topical treatment with the nitric oxide donor, cPTIO, suggesting a dependence on skin release of nitric oxide induced by UVR from the FS40 sunlamps. Put together, these findings indicate a potential benefit for UVR and sun exposure in limiting weight gain and metabolic dysfunction induced by excessive weight gain and obesity.

There are a limited number of preclinical studies that report on the effects of ongoing exposure to UVR on weight gain and signs of metabolic dysfunction (Nakano et al. 2011, Geldenhuys et al. 2014). A strength of our preclinical approach is that we measured the direct effects of ongoing UVR exposure of a known dose. Sun exposure is not easy to quantify in humans, especially over the long time frame required for obesity development. In addition, many other factors can be readily controlled in animal studies, including genetics (inbred mice), environment (in particular temperature and duration of light/darkness cycles), diet and exercise, which are all considerably more challenging in human studies. Even so, a number of difficult-to-control factors could affect the results, which are hard to identify but may have included the effects of different batches of diet or mice, and where breeding may be seasonally affected. We also acknowledge that mice have different skin to humans, with substantially more hair/fur and a thinner epidermis. To test for potential translatability of results, it will therefore be important to reproduce the findings of this study in a human setting. Our findings are similar to those of Al-Daghri and coworkers, in which 59 adults from Saudi Arabia underwent a year-long intervention, which promoted sun exposure (5–30 min) two times a week and increased consumption of vitamin D-rich foods (Al-Daghri et al. 2012). The prevalence of metabolic syndrome reduced from 25% to 13%, accompanied by reduced dyslipidemia (Al-Daghri et al. 2012). The participants of this study had an initial mean BMI of 29.2, suggesting that many were overweight and obese (Al-Daghri et al. 2012). Together with our new findings, these results suggest that low-level sun exposure may

<table>
<thead>
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<th>Table 3</th>
<th>Liver weights and histopathology scores at the end of the experiment.</th>
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<tbody>
<tr>
<td>Diet</td>
<td>UVR and/or skin treatment</td>
</tr>
<tr>
<td>---------</td>
<td>---------------------------</td>
</tr>
<tr>
<td>1 LFD</td>
<td>Mock UVR + vehicle</td>
</tr>
<tr>
<td>2 HFD</td>
<td>Mock UVR + vehicle</td>
</tr>
<tr>
<td>3 HFD</td>
<td>FS40 UVR + vehicle</td>
</tr>
<tr>
<td>4 HFD</td>
<td>SNAP</td>
</tr>
<tr>
<td>5 HFD</td>
<td>FS40 UVR + cPTIO</td>
</tr>
<tr>
<td>6 HFD</td>
<td>CLEO UVR + vehicle</td>
</tr>
<tr>
<td>n/treatment</td>
<td>29–31</td>
</tr>
</tbody>
</table>

* $P<0.05$ relative to HFD (mock UVR + vehicle), one-way ANOVA (Tukey post hoc analysis); † $P<0.05$ relative to LFD (mock UVR + vehicle), one-way ANOVA (Tukey post hoc analysis).

AST, aspartate aminotransferase; AUC, area under curve; cPTIO, carboxy-PTIO; HFD, high fat diet; LFD, low fat diet; SNAP, S-nitroso-N-acetyl-DL-penicillamine; UVR, ultraviolet radiation.
reduce adiposity and improve cardiometabolic outcomes in overweight and/or obese people.

The protective effects of CLEO UVR in suppressing circulating LDL cholesterol levels are supported by studies in humans. Sun exposure for 15 days reduced the LDL/HDL cholesterol ratio in the serum of adults with psoriasis undergoing heliotherapy (Osmancevic et al. 2009). Oxidized LDL cholesterol levels increased with latitude in men with stable coronary heart disease (Grau et al. 2007). Dietary administration of the eNOS substrate l-arginine to healthy elderly human volunteers for 2 weeks lowered circulating LDL but not HDL cholesterol levels (Hurson et al. 1995), suggesting that systemic elevation of nitric oxide exerts effects consistent with the ones we here observed with UVR. Combined with observations from the literature, our findings suggest that safe sun exposure might be a way of reducing serum LDL cholesterol levels in patients with cardiometabolic dysfunction.

The effects of FS40 UVR on weight, weight gain and liver steatosis were reversed by immediate skin treatment with the nitric oxide scavenger cPTIO, suggesting that skin release of nitric oxide mediates the protective effects of UVR on these signs of adiposity. In the current study, we did not observe all the effects of nitric oxide donor SNAP, as previously found, when it suppressed body weight and liver steatosis (Geldenhuys et al. 2014) (compared to only liver steatosis in the current study). A difference between these findings might be accounted for by reduced treatment time in the current study, whereas previously SNAP was administered to skin from when mice were first fed a high-fat diet (Geldenhuys et al. 2014). In addition, our results suggest that skin exposure to UVR is more effective (than SNAP) at increasing the bioactivity of nitric oxide and related metabolites from skin. Human and mouse skin contain large stores of nitrogen oxides, which are mobilized into the circulation by exposure of skin to sub-erythemal UVR (Geldenhuys et al. 2014, Liu et al. 2014). The precise mechanism for this process is yet to be described. The results of other preclinical studies using eNOS (endothelial nitric oxide synthase)-deficient mice (Nozaki et al. 2015) or chemical inhibitors of eNOS (Sheldon et al. 2015), suggest that nitric oxide can reduce liver steatosis impairing hepatic blood flow (Nozaki et al. 2015) and/or reducing hepatic mitochondrial activity (Sheldon et al. 2015). Further work is required to determine the mechanism(s) through which the skin release of nitric oxide by UVR prevents the excessive accumulation of fat in the liver.

We have previously shown that the effects of UVR on reducing weight gain and the development of signs of metabolic dysfunction in male mice fed a high fat diet are independent of vitamin D (Geldenhuys et al. 2014). Male mice have significantly impaired capacity to increase circulating 25-hydroxyvitamin D levels in response to UVR (Gorman et al. 2012, Geldenhuys et al. 2014, Xue et al. 2015). This may be because male mice have reduced epidermal stores of 7-dehydrocholesterol (Gorman et al. 2012, Xue et al. 2015), which are suppressed in an androgen-specific fashion (Xue et al. 2015). Vitamin D-deficient male mice also have increased renal levels of the vitamin D breakdown enzyme, 24-hydroxylase (CYP24A1) (Gorman et al. 2012). We previously reported that dietary vitamin D₃ (2280IU/kg) did not have anti-obesogenic effect when administered alone, and when combined with FS40 UVR, dietary vitamin D prevented the suppressive effects of FS40 UVR on weight gain, WAT weight and fasting glucose levels (Geldenhuys et al. 2014). Although dietary vitamin D improved liver steatosis, FS40 UVR was more effective (Geldenhuys et al. 2014). In human epidemiological studies, obesity is associated with poorer vitamin D status, but low 25(OH)D levels may be caused by obesity rather than vice versa (Vimaleswaran et al. 2013). The effects of vitamin D supplementation on improving cardiometabolic function in humans are uncertain with inconclusive results reported in a meta-analyses of randomized controlled trials that tested the efficacy of vitamin D to modulate weight gain (Mallard et al. 2016), cardiovascular disease, stroke, blood pressure, blood lipids and glucose metabolism (Aytier et al. 2014).

Feeding C57BL/6J mice a high-fat diet usually increases the extent of hepatic steatosis (Duval et al. 2016, Gavito et al. 2016, Song et al. 2016). However, unlike our previous studies (Geldenhuys et al. 2014), significant liver steatosis was observed in mice fed a low-fat diet (Fig. 4 and Table 3). Similarly, we also observed higher than expected levels of AST in the serum of mice fed the low-fat diet, with levels comparable to those fed the high-fat diet (Table 3). We used the same diets as those in previously published studies, with an identical experimental approach for the mice fed the high- and low-fat diets only (Geldenhuys et al. 2014). Therefore, it is difficult to understand why our previous observations were not reproduced in the current study. Mice were housed in a similar fashion (open-topped cages with 6 mice per cage). However, it is possible that there may have been a different strategy used to breed the mice by our supplier (e.g. age of dams, diet) or through animal handling (e.g. sex of technician/researcher (Sorge et al. 2014)) within our bioresources facility (e.g. time of year of study) that may account for this unexpected observation.
The mice in the current study were housed at 21°C. It would be interesting to determine the effects of regular skin exposure to UVR on the development of obesity in mice housed in warmer, thermoneutral conditions. Thermoneutrality may promote adiposity in mice fed a high-fat diet (Cui et al. 2016). The effects of exposure to sub-erythemal UVA radiation in reducing arterial blood pressure of young adult male volunteers were independent of temperature, with similar increases in the skin temperatures of individuals exposed to the UVA or sham irradiation protocols (Liu et al. 2014). In our studies, we used sub-erythemal doses of UVR, which did not induce edema or burn the skin of the mice. Whether there is an anti-obesogenic role for heat production in the skin after exposure to UVR is uncertain.

In conclusion, our studies demonstrate that low-dose (sub-erythemal) UVR reduces the expression of risk factors of adiposity and cardiometabolic dysfunction in already overweight mice. We observed a reduction in weight gain of ~10%, but a much larger reduction (>50%) in circulating fasting insulin levels with UVR treatment. These observations are similar to those from weight loss studies in humans, where small weight losses of ~5% can correspond to significant improvements in insulin sensitivity and reductions in hepatic fat (Magkos et al. 2016). Further work is needed in the form of clinical trials to assess the efficacy of safe sun exposure and/or UVR phototherapy to reduce the cardiometabolic risk in susceptible people.

Supplementary data
This is linked to the online version of the paper at doi.org/10.1530/JOE-16-0616.

Declaration of interest
Prof. Fee lis ch and Dr Weller are members of the Scientific Advisory Board of AOBiome LLC, a company commercializing ammonia-oxidizing bacteria for use in inflammatory skin disease. Dr Weller is also a Director of and Prof. Fee lis ch a Scientific Advisor for RelaxSol Ltd, a company developing novel sunscreen and skincare products. We have no further disclosures or conflicts of interest to declare.

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Author contribution statement
S G conceived and designed this study with input from N F, M F, P H H, R B W, J S and V M. N F acquired and analyzed the data for the study with help from J S and S G. All authors have contributed toward the interpretation of findings from this study, have played a role in drafting the article or revising it critically for its intellectual content and have given their final approval for this version of the paper to be published.

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