LEPTIN RESENSITISATION: A REVERSION OF LEPTIN-RESISTANT STATES

Maria F Andreoli1,2, Jose Donato Jr3, Isin Cakir4,5 and Mario Perello6

1Laboratory of Experimental Neurodevelopment, Institute of Development and Paediatric Research (IDIP), La Plata Children’s Hospital and Scientific Research Commission, Province of Buenos Aires (CIC-PBA), La Plata, Buenos Aires, Argentina
2Argentine Research Council (CONICET), La Plata, Buenos Aires, Argentina
3Department of Physiology and Biophysics, Institute of Biomedical Sciences, University of São Paulo, São Paulo, São Paulo, Brazil
4Life Sciences Institute, University of Michigan, Ann Arbor, Michigan, USA
5Department of Molecular Physiology and Biophysics, Vanderbilt University, Nashville, Tennessee, USA
6Laboratory of Neurophysiology, Multidisciplinary Institute of Cell Biology (IMBICE, Argentine Research Council (CONICET), National University of La Plata and Scientific Research Commission, Province of Buenos Aires (CIC-PBA)), La Plata, Buenos Aires, Argentina

Correspondence should be addressed to M F Andreoli: mfandreoli@fbcb.unl.edu.ar

Abstract

Leptin resistance refers to states in which leptin fails to promote its anticipated effects, frequently coexisting with hyperleptinaemia. Leptin resistance is closely associated with obesity and also observed in physiological situations such as pregnancy and in seasonal animals. Leptin resensitisation refers to the reversion of leptin-resistant states and is associated with improvement in endocrine and metabolic disturbances commonly observed in obesity and a sustained decrease of plasma leptin levels, possibly below a critical threshold level. In obesity, leptin resensitisation can be achieved with treatments that reduce body adiposity and leptinaemia, or with some pharmacological compounds, while physiological leptin resistance reverts spontaneously. The restoration of leptin sensitivity could be a useful strategy to treat obesity, maintain weight loss and/or reduce the recidivism rate for weight regain after dieting. This review provides an update and discussion about reversion of leptin-resistant states and modulation of the molecular mechanisms involved in each situation.

Key Words
- diet-induced obesity
- leptin resistance
- obesity
- leptin resensitisation

Introduction

Obesity results from an imbalance between energy intake and energy expenditure and represents a major threat to health across the globe. It is a chronic and multifactorial condition related to numerous diseases that often leads to premature disability and mortality. The discovery of the anorexigenic hormone leptin in 1994 (Zhang et al. 1994) opened a new field within the therapeutic strategies to combat obesity. Leptin, a product of the obesity (ob) gene, displays key anti-obesity actions as it inhibits food intake and induces energy expenditure (Pelleymounter et al. 1995). This anti-obesity effect was demonstrated in individuals bearing congenital leptin deficiencies (Farooqi et al. 1999) and raised expectations regarding its potential as a drug to reduce body weight in obese patients. However, leptin therapy turned out to be disappointing as a weight loss strategy since most obese individuals have hyperleptinaemia associated with loss of responsiveness to this hormone. Such lack of sensitivity to leptin inspired the notion of leptin resistance (Frederich et al. 1995). Leptin resistance has been described in several conditions (from obese individuals to pregnant females and seasonal animals) and encompasses numerous molecular and cellular mechanisms that have not yet been fully elucidated. The restoration of leptin sensitivity
could be a useful strategy to treat obesity, maintain weight loss and/or reduce the recidivism rate for weight regain after dieting (Chhabra et al. 2016). The idea of leptin resensitisation emerges as a new concept that refers to the reversion of leptin-resistant states, usually linked to reduction of body adiposity and leptinaemia. This review summarises the current knowledge about the reversion of leptin-resistant states.

**General aspects of leptin and its receptor: molecular leptin signalling**

Leptin is a 167-residue peptide hormone mainly produced by adipocytes and acts in the central nervous system to primarily coordinate the metabolic adaptations to fasting (Zhang et al. 1994, Margetic et al. 2002). Leptin is also expressed in other tissues, but their contribution to the total leptinaemia is negligible (Guilmeau et al. 2004, Odle et al. 2014). Except during prolonged fasting, leptinaemia correlates with the mass of adipose tissue (Frederich et al. 1995), thereby representing a key marker of energy storage. As discussed below, leptinaemia falls when energy stores decrease (e.g., starvation), increasing appetite and decreasing energy use. Conversely, leptinaemia increases with adequate energy stores.

Newly synthesised leptin is sorted into the secretory pathway and secreted at the same rate as it is synthesised; as a consequence, the stimulated secretion of leptin mainly depends on the transcription and translation rates of the ob gene (Coleman & Herrmann 1999, Cammisotto et al. 2006). Thus, leptinaemia shows little short-term variation and requires several hours (or even days) to modify in response to changes in the nutritional status (Myers 2006). Leptin content in adipose tissue directly depends on the fat cell size and is regulated by the hormonal milieu: exposure to high insulinaemia and glucocorticoid levels increases leptin production (Lee et al. 2007). Besides, the sympathetic nervous system, via activation of beta-adrenergic receptors, inhibits leptin production (Ricci & Fried 1999).

Leptin acts via its specific receptor, named LepR. Murine Lepr gene is alternatively spliced to give rise to six isoforms, LepRa-LepRf. LepR isoforms, except LepRe, differ in the length of the intracellular domain, and therefore, in their physiological role. LepRa isoform transports plasma leptin into the brain (Elmqist et al. 1998, Hileman et al. 2002). Only LepRb contains all intracellular motifs required for full activation of the signalling pathways and, consequently, is essential for leptin action (Tartaglia et al. 1995). LepR belongs to class 1 cytokine receptor family and lacks intrinsic enzymatic activity.

The binding of leptin to LepRb activates the tyrosine kinase janus kinase 2 (JAK2), which is associated with LepRb and phosphorylates the tyrosine residues Y985, Y1077 and Y1138 on LepRb (Fig. 1). These phosphorylated residues act as docking sites for signalling molecules that contain Src homology 2 (SH2)-domain and mediate the subsequent intracellular events (Banks et al. 2000). Phosphorylated Y1138 recruits the signal transducer and activator of transcription (STAT) 3, a latent transcription factor mediating major effects of LepRb (Bates et al. 2003, Gao et al. 2004). STAT3 is then phosphorylated by JAK2, generating pSTAT3 that undergoes dimerisation and translocates into the cell nucleus to act as an active transcription factor. Phosphorylated Y985 recruits a SH2-containing tyrosine phosphatase (SHP2), which activates mitogen-activated protein kinase (MAPK) and extracellular signal-regulated kinases (ERK) signalling pathways (Banks et al. 2000). Phosphorylated Y985 also binds to the suppressor of cytokine signalling 3 (SOCS3), whose gene transcription depends on pSTAT3 and exerts an inhibitory effect on LepRb signalling (Bjorbaek et al. 1999). Phosphorylated Y1077 recruits STAT5, which mediates some effects of leptin on energy balance and reproduction as well as effects of prolactin (Tups 2009). Leptin-induced JAK2 activation also leads to the phosphatidylinositol 3-kinase (PI3K) activation, which is the major signalling pathway utilised by insulin and insulin-like growth factors (Hekerman et al. 2005). Notably, JAK2/STAT pathway mainly regulates gene expression while JAK2/PI3K pathway regulates not only gene expression but also fast cellular events by regulating ion channels (Donato et al. 2010, Gavello et al. 2016).

**Biological effects of leptin**

Leptin is a pleiotropic hormone that displays a variety of effects that seem to depend on its circulating level. Leptinaemia decreases under fasting, playing a critical role initiating the neuroendocrine response to starvation, including limiting procreation, decreasing thyroid thermogenesis and increasing secretion of stress steroids, which together are likely to have survival value during prolonged nutritional deprivation (Ahima et al. 1996). Leptin replacement blunts some of these fasting-induced adaptations, mainly concerning the gonadal, adrenal and thyroid axes (Ahima et al. 1996). Thus, studying the effects of systemic administration of leptin to fasted animals,
Leptin resensitisation

M F Andreoli et al.

Leptin resensitisation

Review

M F Andreoli et al. Leptin resensitisation

in doses that increase leptinaemia to levels similar to those found in fed animals, has helped to clarify some of the biologically relevant effects of leptin and supported the notion that leptinaemia in the lower concentration range is a key coordinator of the adaptation to negative energy balance conditions (Ahima et al. 1996). In contrast, the extent to which leptinaemia in the higher concentration range affects some physiological functions is still a matter of debate. Some groups suggest that hyperleptinaemia displays some actions in obesity that prevent further body weight gain (Myers et al. 2012, Ottaway et al. 2015), while others argue that the anti-obesity effect of hyperleptinaemia has not yet been clearly demonstrated (Ahima et al. 1996, Flier & Maratos-Flier 2017). In addition, some effects of leptin have been unmasked by administering pharmacological doses of leptin or by using non-physiological strategies of administration (e.g. chronic infusion, central infusions, infusions in specific brain areas); thus, the physiological implications of these observations are uncertain. Bearing these considerations in mind, some of the described biological effects of leptin are the following:

Leptin reduces food intake


The anorectic effect of leptin is mainly mediated by the hypothalamic arcuate nucleus (ARH) (Munzberg et al. 2004). In the ARH, leptin activates anorexigenic neurons that express proopiomelanocortin (POMC), which is cleaved into different neuropeptides, including α-melanocyte-stimulating hormone (α-MSH). α-MSH inhibits food intake via the melanocortin receptor 3 and 4 (MC3R/MC4R) (Coppari et al. 2005). In the ARH, leptin also inhibits orexigenic neurons that produce neuropeptide Y (NPY), agouti-related protein (AgRP) and gamma-aminobutyric acid (GABA) (Coppari et al. 2005). Leptin-responsive neurons outside the ARH could also

Figure 1

Leptin signalling. Binding of leptin to its receptor, LepRb, induces the activation of JAK2, which in turn phosphorylates the intracellular domain of LepRb, especially at tyrosines Y985, Y1077 and Y1138. Phosphorylated Y985 serves as a docking site for SHP2, activating MAPK/ERK signalling pathway. Phosphorylated Y1077 recruits STAT5. Phosphorylated Y1138 recruits STAT3, inducing its phosphorylation, dimerisation and nuclear translocation to activate the transcription of target molecules such as SOCS3. Induction of JAK2 can also stimulate PI3K, which regulates fast cellular events by regulating ion channels. LepRb also receives inhibitory signals from multiple negative feedback loops such as SOCS3 and PTP1B. ERK, extracellular signal-regulated kinase; JAK2, Janus kinase 2; LepRb, leptin receptor b; MAPK, mitogen-activated protein kinase; PI3K, phosphatidylinositol 3-kinase; SHP2, Src homology 2-containing tyrosine phosphatase; SOCS3, suppressor of cytokine signalling 3; STAT3, signal transducer and activator of transcription 3; STAT5, signal transducer and activator of transcription 5. A full colour version of this figure is available at https://doi.org/10.1530/JOE-18-0606.
mediate the anorectic effects of the hormone. Particularly, the ventromedial nucleus (VMH) (Dhillon et al. 2006), the dorsomedial nucleus (DMH) and the paraventricular nucleus (PVH) of the hypothalamus are responsive to leptin and work as an interconnected circuit (Myers et al. 2009, Perello & Raingo 2013, Sutton et al. 2016). Other proposed targets of the anorexigenic effects of leptin include glutamatergic neurons of the median preoptic area (MPO) (Yu et al. 2016), dopamine neurons of the ventral tegmental area (VTA) (Fulton et al. 2006) and neurons of the nucleus of the solitary tract, which is an important relay of gastrointestinal sensory inputs (Garfield et al. 2012). In spite of these studies, the relative physiological relevance of these non-ARH areas mediating anorexigenic effects of leptin is still unclear. Regulation of food intake by leptin largely depends on pSTAT3, as point mutation of Y1138 or deletion of brain STAT3 lead to hyperphagic obesity (Bates et al. 2003). LepRb-induced PI3K signalling appears to be rather important for acute suppression of food intake (Xu et al. 2010).

**Leptin increases energy expenditure and thermogenesis**

A single intracerebroventricular (icv) injection or peripheral infusion of leptin slightly increase or have no effect on energy expenditure (Scarpace et al. 1997, Doring et al. 1998, Gautron & Elmquist 2011). Chronic administration of leptin to mimic the leptinaemia kinetics observed in obesity slightly decreases energy expenditure (Ravussin et al. 2014), and higher doses of leptin induce long-lasting effects that can completely deplete body fat stores in animals (Halas et al. 1997, Monteza et al. 2005). In addition, leptin deficiency causes a reduction in metabolic rate in ob/ob mice (Breslow et al. 1999). Thus, endogenous leptinaemia seems to be able to affect energy expenditure and thermogenesis under normal circumstances.

The effects of leptin on energy expenditure are mediated by both the autonomic nervous system and neuroendocrine hypothalamic–pituitary–thyroid (HPT) axis (Pandit et al. 2017). Leptin upregulates the activity of the sympathetic nervous system (Haynes et al. 1997) presumably via its action on multiple neuronal targets that include not only ARH POMC and NPY/AgRP/GABA neurons (Cowley et al. 2001, Harlan et al. 2011) but also MPO (Yu et al. 2016), VMH (Dhillon et al. 2006) and DMH (Enriori et al. 2011). Leptin activates the HPT axis via its direct action on thyrotropin-releasing hormone (TRH) neurons of the PVH (Perello et al. 2006) and also via an indirect action through ARH POMC and NPY/AgRP/GABA neurons that provide potent stimulatory and inhibitory inputs, respectively, to PVH TRH neurons (Fekete et al. 2000).

**Leptin increases heart rate and blood pressure**

LepRb is expressed in brain regions and peripheral organs (e.g. heart, kidneys and adrenals) that are important in cardiovascular control and blood pressure regulation (Fruhbeck 2002). Central administration of leptin or direct injections in the DMH or the VMH increase both mean arterial pressure and/or heart rate in rodents (Dunbar et al. 1997, Casto et al. 1998, Marsh et al. 2003). However, the physiological significance of these observations is uncertain. In humans, chronic administration of leptin does not elevate blood pressure (Simonds et al. 2017).

**Leptin decreases glycaemia**

Leptin administration at a dose that does not affect body weight and food intake normalises blood glucose and insulin levels in otherwise hyperglycemic ob/ob mice (Pelleymounter et al. 1995). Leptin decreases glycaemia by sensitising metabolically relevant tissues to insulin but also in an insulin-independent manner (Fujikawa & Coppari 2015). Leptin improves the glycaemic control via its effects at both the central and peripheral level, where leptin suppresses the production of glucagon and corticosterone, increases glucose uptake and inhibits hepatic glucose output (D’Souza et al. 2017). The central effects of leptin on glucose homeostasis strongly depend on the ARH (Coppari et al. 2005).

**Leptin is a permissive factor for puberty and fertility**

The administration of physiological amounts of leptin prevents the fasting-induced delay in ovulation (Ahima et al. 1996). Leptin signalling is required to enter puberty (de Luca et al. 2005). LepRb is not expressed in gonadotropin-releasing hormone neurons. Therefore, leptin indirectly controls the reproductive function via interneurons located at the ventral premammillary nucleus or through ARH POMC and NPY/AgRP/GABA neurons (Donato et al. 2011, Donato & Elias 2011).

**Leptin resistance**

Leptin resistance refers to the states in which leptin fails to promote its anticipated effects, frequently coexisting with
marked hyperleptinaemia. A schematic representation of the development of leptin resistance can be seen in Figs 2 and 3. The assessment of leptin resistance encompasses diverse aspects. The standard biochemical marker for cellular LepRb action is leptin-induced STAT3 phosphorylation (Myers et al. 2010), and impairment of this induction is usually interpreted as an indication of leptin resistance. The measurement of the acute or chronic ability of exogenously-administered leptin to reduce body weight, adiposity and/or food intake is also used to estimate the sensitivity to leptin. Practically speaking, ‘leptin resistance’ is a broadly applied and context-dependent term with no universal, quantifiable and clinically useful definition (Myers et al. 2012).

Since a major physiological function of leptin is to signal energy deficiency, the implications of hyperleptinaemia and the concomitant notion of ‘leptin resistance’ become controversial. Besides, much of the evidence for leptin resistance relies on pharmacological studies that use non-physiological doses or routes of leptin administration. A variety of arguments suggest that a ‘leptin resistance’ underlies the development of obesity; however, it has been also considered that leptin action naturally faces a ceiling effect, beyond which it promotes little additional effect (Ahima et al. 1996, Myers et al. 2010, Rosenbaum & Leibel 2014, Flier & Maratos-Flier 2017). The concept of leptin resistance is also dependent on which of the biological effects of leptin are affected. The fact that in some forms...
of obesity there may be resistance to the anorectic and weight-reducing actions of leptin but preservation of hypertension led to the concept of selective leptin resistance (Rahmouni et al. 2005, Mark 2013).

**Cellular and molecular mechanisms of leptin resistance**

**Brain accessibility**

Leptin resistance may involve a limited accessibility of leptin to the brain. Leptin accesses the brain via a saturated transport system (Banks et al. 1996) and cerebrospinal fluid/serum leptin ratio has been shown to be decreased in obesity (Caro et al. 1996, Schwartz et al. 1996). Central administration of leptin to obese animals reduces food intake more potently than peripheral administration of the hormone, suggesting that leptin transport to the brain is impaired in obesity (Halas et al. 1997, Van Heek et al. 1997). Interestingly, leptin transport across the blood–cerebrospinal fluid barrier (Kleinert et al. 2018), rather than across the blood–brain barrier (Harrison et al. 2018), seems to be impaired in obese animals. Notably, rats lacking functional LepRa display decreased transport of leptin to the brain and develop obesity (Kastin et al. 1999).

**Hyperleptinaemia-induced downregulation of LepRb expression**

Reduced hypothalamic LepRb expression is found in some models displaying hyperleptinaemia (Martin et al. 2000), and leptin resistance has been hypothesised to be secondary to the hyperleptinaemia found in obesity (Knight et al. 2010). In support of this notion, chronic hyperleptinaemia induced by transgene overexpression decreases hypothalamic LepRb levels and impairs leptin signalling (Wilsey et al. 2002). In addition, chronic high central leptin desensitises its physiological effects over time rendering lean rodents more susceptible to obesity (Scarpace et al. 2005).

**Alteration of signalling cascade**

Leptin resistance seems to involve intracellular proteins that impair LepR signalling. As stated above, SOCS3 blocks LepRb signalling. Since high hypothalamic SOCS3 expression is found in most hyperleptinaemic conditions, it is hypothesised that upregulation of SOCS3 in obesity impairs leptin-induced pSTAT3 (Bjorbaek et al. 1999). The key role of SOCS3 regulating leptin sensitivity has been confirmed by its either selective inactivation (Bjornholm et al. 2007, Pedroso et al. 2016) or overexpression (Reed et al. 2010). Leptin action is also downregulated by protein tyrosine phosphatases (PTPs, such as PTP1B), which block leptin signalling through the dephosphorylation of LepRb, JAK2 or pSTAT3 and are increased in some models that display leptin resistance (St-Pierre & Tremblay 2012). Recently, matrix metalloproteinase-2 (Mmp-2) has been suggested as a key mediator of leptin resistance. Obesity promotes the hypothalamic Mmp-2 activity and its secretion from the astrocytes and AgRP neurons (Mazor et al. 2018). Mmp-2 cleaves the extracellular domain of LepRb and diminishes leptin action (Mazor et al. 2018).

**Hypothalamic inflammation and oxidative stress**

Leptin resistance has been linked to the hypothalamic inflammation, which involves the IκB kinase-β/nuclear factor-κB (IKKβ/NF-κB) signalling (de Git & Adan 2015). Activation of IKKβ/NF-κB pathways induces SOCS3 expression and production of proinflammatory cytokines, such as interleukin (IL) 1 and 6 and tumour necrosis factor-α (Zhang et al. 2008). Proinflammatory cytokines, in turn, increase SOCS3 and PTP1B, leading to leptin resistance (de Git & Adan 2015). Notably, constitutive activation of hypothalamic IKKβ impairs leptin signalling and increases weight gain and food intake, while genetic or pharmacological inhibition of IKKβ activity protects against obesity and improves leptin sensitivity in obese mice (Cakir & Nillni 2018a). Besides, it was recently shown that hypothalamic oxidative stress impairs the function of POMC neurons and suppresses leptin signalling in the hypothalamus, resulting in the development of systemic leptin resistance and obesity (Yagishita et al. 2017).

**Endoplasmic reticulum (ER) stress**

Impairment of leptin signalling has been linked to ER stress, which is caused by an excessive accumulation of unfolded proteins that activate the unfolded protein response (Cakir et al. 2013). This response promotes an improvement of the ER protein-folding capacity, the degradation of misfolded proteins and the reduction of the load of new proteins entering the ER to resolve protein-folding defects. Obesity-induced hypothalamic ER stress reduces the post-transcriptional processing of POMC and impairs the biosynthesis of α-MSH (Cakir et al. 2013). Notably, pharmacologically induced hypothalamic ER stress in lean animals increases SOCS3 and PTP1B and promotes leptin resistance (Cakir et al. 2013). Thus, obesity-related hypothalamic ER stress can promote central leptin resistance (Cakir & Nillni 2018b).
**Table 1** Conditions displaying leptin resistance and their strategies of reversion.

<table>
<thead>
<tr>
<th>Condition</th>
<th>Experimental model</th>
<th>Main features (key references)</th>
<th>Tested strategies to reverse leptin-resistant state (key references)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Obesity</td>
<td>DIO</td>
<td>Most DIO models display hyperleptinaemia, higher basal pSTAT3, diminished leptin-induced pSTAT3, increased SOCS3 and ER stress ([Munzberg et al. 2004, Enriori et al. 2007, Cakir et al. 2013, Ottaway et al. 2015]. Features of DIO models depend on experimental factors such as species, diet composition and duration of the manipulation)</td>
<td>• Fasting ([Pedroso et al. 2016, Caron et al. 2018])</td>
</tr>
<tr>
<td>Monogenic mutations</td>
<td></td>
<td>Several genetic models are useful tools to study leptin resistance. Most of them display one or more features of diminished leptin sensitivity ([Bates et al. 2003, Gao et al. 2004, Elmqquist et al. 2005, Myers et al. 2010, Liao et al. 2012, Lutz &amp; Woods 2012])</td>
<td>• Energy restriction ([Morabito et al. 2017])</td>
</tr>
<tr>
<td>Obesity-prone rats</td>
<td></td>
<td>Obesity-prone rats are early leptin-resistant and display diminished leptin-induced pSTAT3 and reduced LepRb expression without hyperleptinaemia prior to the development of obesity ([Levin et al. 1997, 2003, Irani et al. 2007, Bouret et al. 2008])</td>
<td>• Switch to low fat diets ([Enriori et al. 2007, Morabito et al. 2017])</td>
</tr>
<tr>
<td>Early overfeeding</td>
<td></td>
<td>Reduction of the litter size leads to decreased pSTAT3 and increased SOCS3 ([Plagemann et al. 2012])</td>
<td>• Fasting ([Shapiro et al. 2008])</td>
</tr>
<tr>
<td>ARH ablation</td>
<td></td>
<td>Monosodium glutamate-treated rodents display increased adiposity, hyperleptinaemia and resistance to the anorexigenic effects of leptin ([Perello et al. 2003])</td>
<td>• Exercise ([Shapiro et al. 2011, Kang et al. 2013])</td>
</tr>
<tr>
<td>Age-related obesity</td>
<td></td>
<td>Ageing is associated with body weight gain, central leptin resistance, upregulation of hypothalamic SOCS3, reduced leptin-induced pSTAT3 and diminished LepR expression ([Scarpace &amp; Tumer 2001])</td>
<td>• Normal body weight is restored in obese pomc-knockout mice if POMC is re-expressed after a reduction in leptinaemia ([Bumaschny et al. 2012, Chhabra et al. 2016])</td>
</tr>
<tr>
<td>Pregnancy</td>
<td>Pregnant rodents</td>
<td>Increase in food intake and adiposity are associated to hyperleptinaemia and leptin resistance compromising the VMH. Prolactin and placental lactogens induce impaired leptin-induced pSTAT3 and pSTAT5, increased expression of SOCS3, reduced LepRb expression and impairment of leptin transport ([Ladyman &amp; Grattan 2005, Tups 2009, Ladyman et al. 2010, Trujillo et al. 2011])</td>
<td>• Total reversion by partition ([Ladyman &amp; Grattan 2005, Tups 2009, Ladyman et al. 2010, Trujillo et al. 2011])</td>
</tr>
<tr>
<td>Seasonal animals</td>
<td>Siberian hamsters and field voles</td>
<td>Seasonal animals show hyperleptinaemia and high food intake during long day photoperiod (summer) with impairment of leptin-induced pSTAT3, prolactin-induced increase of pSTAT5 and increased SOCS3 and PTP1B ([Anderson et al. 2006, Tups et al. 2004, Tups 2009])</td>
<td>• Total reversion by switch to short day photoperiod ([Tups et al. 2004, Tups 2009])</td>
</tr>
</tbody>
</table>

DIO, diet-induced obesity; ER, endoplasmic reticulum; LepRb, leptin receptor b; POMC, proopiomelanocortin; PTP1B, protein tyrosine phosphatase 1 B; SOCS3, suppressor of cytokine signalling 3; STAT3, signal transducer and activator of transcription 3; STAT5, signal transducer and activator of transcription 5; VMH, ventromedial nucleus.

**Conditions displaying leptin resistance**

**Obese animals**

Several rodent models of obesity are used to investigate the molecular basis of leptin resistance ([Myers et al. 2010]). Such models include diet-induced obesity (DIO), genetic models, obesity-prone models, early overfeeding, age-related obesity and animals with hypothalamic lesions (Table 1).

DIO is likely the most frequent animal model used to investigate leptin resistance because it shares many characteristics with the common form of human obesity,
including an attenuated response to the anorexigenic effect of leptin (Van Heek et al. 1997, Myers et al. 2010). DIO models are obtained by feeding animals with highly palatable and hypercaloric diets for different periods of time. Notably, obesity per se in DIO models is not sufficient for the development of leptin resistance, and the presence of hyperleptinaemia is also required (Knight et al. 2010). An evidence for this notion comes from ob/ob mice implanted with osmotic pumps to bring their leptin levels to those observed in lean wild-type mice: this DIO normoleptinaemic ob/ob model remains leptin sensitive despite developing obesity comparable to a DIO wild-type model (Knight et al. 2010). Notably, only 8 days of high-dose leptin treatment are necessary to induce leptin resistance in wild-type mice (Montez et al. 2005). Besides, mice chronically overexpressing central leptin show leptin resistance and increased susceptibility to high-fat (HF) diet (Scarpace et al. 2005).

Molecular changes linked to hypothalamic leptin resistance in DIO models (Fig. 2) include higher basal pSTAT3 (Cakir et al. 2013), diminished leptin-induced pSTAT3 (Munzberg et al. 2004), increased SOCS3 (Enriori et al. 2007, Ottaway et al. 2015) and ER stress (Cakir et al. 2013). Hypothalamic inflammation has also been detected as early as 24 h after HF diet and prior to any weight gain (Thaler et al. 2012). Interestingly, hypothalamic expression of PTPs is increased in obese mice and their selective ablation improves leptin sensitivity and partially prevents obesity (St-Pierre & Tremblay 2012).

Importantly, specific features of DIO models strongly depend on factors such as animal model (strains of mice or rats), diet composition and duration of the manipulation, among other factors. Commercially available diets used to induce DIO include HF or HF/high sugar content. Although it is difficult to discern the effects of each macronutrient, it is known that saturated fats strongly impair leptin signalling in the hypothalamus (de Git & Adan 2015) and that high fructose intake also leads to leptin resistance (Shapiro et al. 2008). The hypercaloric diet period also impacts on the phenotype and the molecular features detected. In mice, the development of DIO as a consequence of HF diet has been divided into three stages: (a) an early stage in which mice gain weight and remain sensitive to leptin; (b) a further stage in which mice have increased leptin production and still retain central sensitivity to leptin and (c) a later stage in which central leptin sensitivity is decreased (Lin et al. 2000). However, it has been reported that reduced leptin sensitivity can be detected as early as after 6 days on HF diet, when the increase in body weight and circulating leptin is first detectable (Munzberg et al. 2004). Another study showed that 24 h of HF diet led to impaired leptin signalling in the ARH, suggesting that a reduction of leptin sensitivity in the ARH precedes the increase in body weight induced by HF diet (Rizwan et al. 2017).

It is interesting to note that DIO mice are considerably less obese than ob/ob mice and maintain their reproductive function, energy expenditure, sympathetic outflow and other leptin-regulated processes, in contrast to ob/ob- or LepR-deficient (db/db) mice (Rahmouni et al. 2005, Myers et al. 2010). Also, the administration of a LepRb antagonist to DIO mice induces similar effects on energy balance as seen in lean mice (Ottaway et al. 2015). These observations suggest that leptin resistance in DIO models selectively affects the anorectic effect of the hormone, while endogenous hyperleptinaemia is able to exert several effects biologically relevant in DIO animals (Myers et al. 2010). In line with this possibility, it has been shown that the degree of leptin resistance in DIO models differs among brain areas. Thus, leptin fails to decrease food intake or increase pSTAT3 in the ARH of DIO mice while its action in other hypothalamic regions is retained (Munzberg et al. 2004). The action of endogenous leptin in the DMH has been proposed to increase blood pressure in DIO mice (Simonds et al. 2014). Similarly, DIO rats are resistant to the anorectic effect of leptin, although endogenous hyperleptinaemia acts in the PVH TRH neurons upregulating the HPT axis and increasing thermogenesis (Perello et al. 2010). In addition, oxytocin-producing neurons of the PVH remain sensitive to leptin and mediate leptin effects on body weight in DIO rats (Perello & Raingo 2013).

Animal models of obesity concerning monogenic mutations in the leptin pathway have also been used to study leptin resistance (Bates et al. 2003, Gao et al. 2004). These models include db/db mice and their counterpart, the Zucker rats. Additionally, POMC-deficient mice, MC4R-deficient mice and Otsuka Long Evans Tokushima Fatty rats, which lack functional receptors for cholecystokinin, display diminished leptin sensitivity (Elmqquist et al. 2005, Myers et al. 2010, Lutz & Woods 2012). Interestingly, mice with truncations of the brain-derived neutrophic factor gene are obese, hyperphagic, hyperleptinaemic and resistant to the anorectic action of leptin, but display normal leptin-induced pSTAT3 in the hypothalamus suggesting that some biochemical markers of LepR signalling do not necessarily reflect the physiological response to leptin (Liao et al. 2012).

A different model is the obesity-prone rat, a polygenic substrain developed by selectively breeding outbred
Sprague–Dawley rats highly prone to develop DIO (Levin et al. 1997). Obesity-prone rats are early (possibly inborn) leptin resistant. Prior to the development of obesity, they show diminished hypothalamic leptin-induced pSTAT3 (Bouret et al. 2008) and reduced LepRb expression (Levin et al. 2003) without hyperleptinaemia (Irani et al. 2007).

Another obesity model displaying leptin resistance is early overfeeding by reduction of the litter size, a manoeuvre that increases the availability of breast milk in the postnatal period and leads to decreased pSTAT3 and increased SOCS3 in the hypothalamus (Plagemann et al. 2012).

Some animal models of obesity are generated by selective hypothalamic lesions, such as ARH-ablated animals, which are obtained using postnatal administration of monosodium glutamate to rodents (Perello et al. 2003). ARH-ablated animals display increased adiposity, hyperleptinaemia and resistance to the anorexigenic effects of leptin presumably due to the absence of one of the key hypothalamic targets of leptin (Perello et al. 2003).

Age-related obesity is also associated with body weight gain, central leptin resistance, upregulation of hypothalamic SOCS3, reduced leptin-induced pSTAT3 and diminished LepR expression (Scarpace & Tumer 2001).

**Pregnancy**

Gestation is associated to hyperleptinaemia and leptin resistance (Table 1). Gestational leptin resistance is characterised by impaired leptin-induced pSTAT3 in the hypothalamus and likely plays a role in the adaptations observed during pregnancy, including increase in food intake and adiposity (Ladyman et al. 2010). Some evidence indicates that prolactin and placental lactogens are involved in gestational leptin resistance (Ladyman et al. 2010). Prolactin levels are elevated during the first half of pregnancy, while placental lactogens progressively increase during late gestation (Augustine & Grattan 2008). Many hypothalamic LepR-expressing neurons are directly responsive to prolactin, which increases hypothalamic expression of SOCS3 via activation of pSTAT5 and abolishes the anorexigenic effects of leptin (Trujillo et al. 2011). Notably, ablation of Socs3 gene in LepRb-expressing cells of female mice does not affect fertility but increases leptin sensitivity during gestation and mitigates pregnancy-induced metabolic changes (Zampieri et al. 2015).

Gestational leptin resistance could also include reduced hypothalamic expression of LepRb mRNA (Ladyman & Grattan 2005) and impairment of leptin transport into the brain due to the sustained hyperprolactinemia (Trujillo et al. 2011). Unlike DIO models, impairment of leptin sensitivity during pregnancy seems to compromise the VMH (Ladyman & Grattan 2005, Tups 2009). However, SOCS3 ablation in steroidogenic factor-1 neurons of the VMH only causes modest metabolic effects during pregnancy (Ramos-Lobo et al. 2017).

**Small seasonal animals**

Seasonal mammals, such as Siberian hamsters and field voles, exhibit a natural body weight cycle, accompanied by a biannual reversible switch in leptin sensitivity (Table 1). Such seasonal leptin resistance is observed during long day photoperiod (summer), when the animals show hyperleptinaemia and high food intake (Tups et al. 2004). At a molecular level, seasonal leptin resistance is characterised by an impairment of leptin-induced pSTAT3 in the ARH. This phenomenon seems to be also mediated by a prolactin-induced increase of pSTAT5, which in turn increases SOCS3 and PTP1B and further blunts leptin’s central action (Anderson et al. 2006, Tups 2009). Therefore, hypothalamic SOCS3 is employed to sensitise the brain to a different reading of leptin signal in opposite photoperiods (Tups et al. 2004, Tups 2009).

**Leptin resensitisation**

Leptin resensitisation refers to the reversion of leptin-resistant states (Fig. 3). Normalisation of circulating leptin levels after chronic hyperleptinaemia has been shown to affect the sensitivity to the hormone (Montez et al. 2005). In obese animals, leptin resistance can be reversed with treatments that reduce body adiposity and leptinaemia, while physiological leptin resistance reverts spontaneously when pregnant animals give birth or when seasonal animals are exposed to a short-day photoperiod. The reversion of leptin-resistant states is achieved through the modulation of diverse molecular mechanisms depending on the case (Fig. 2 and Table 1).

**Reduction of circulating leptin after a chronic exogenously induced hyperleptinaemia**

Since hyperleptinaemia is required for the development of leptin resistance, normalisation of leptin levels after chronic hyperleptinaemia should improve leptin sensitivity. To the best of our knowledge, only two studies investigated the effects of the withdrawal of leptin after its pharmacological administration. One study showed that mice treated with leptin for 8 days decrease their food

---

Table 1

<table>
<thead>
<tr>
<th>Condition</th>
<th>Outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hyperleptinaemia</td>
<td>Increased adiposity, hyperleptinaemia and resistance to anorexigenic effects of leptin</td>
</tr>
<tr>
<td>Pregnancy</td>
<td>Increased food intake and adiposity during gestation, reduced leptin-induced pSTAT3 in the hypothalamus</td>
</tr>
<tr>
<td>Seasonal mammals</td>
<td>Biannual reversible switch in leptin sensitivity during long day photoperiod</td>
</tr>
<tr>
<td>Small seasonal animals</td>
<td>Reversible switch in leptin sensitivity during long day photoperiod</td>
</tr>
</tbody>
</table>

---
intake in the first 4 days and then normal food intake is recovered (Montez et al. 2005). After leptin withdrawal, food intake transiently increases the first 3 days, while body weight returns to initial values. Thus, mice seem to develop a transient leptin-resistant state after few days of hyperleptinaemia that quickly reverts when the treatment is over. Notably, food intake increments and weight gain can be partially suppressed depending on the magnitude of the leptinaemia reduction, showing that leptin sensitivity is recovered at least partially in the first few days (Montez et al. 2005). A more recent study also showed that the initial weight gain after leptin withdrawal is not sustained in time and that mice slightly gain weight and adiposity after ~4 months of leptin withdrawal (Ravussin et al. 2014). Strikingly, leptinaemia remains elevated in this period (Ravussin et al. 2014). These findings suggest that after chronic exogenously induced hyperleptinaemia, the reversion of leptin resistance and the reduction of circulating leptin are not immediate. Unfortunately, none of these studies was designed to investigate leptin resistance per se, nor the molecular mechanisms involved in the adaptations from hyperleptinaemia to normoleptinaemia.

**Reversion of leptin resistance in obesity models**

Since DIO leads to leptin resistance, it is expected that treatments that reduce body adiposity and hyperleptinaemia should improve leptin sensitivity. Dietary interventions, including fasting, energy restriction and the switch from HF to low-fat diets, induce leptin resensitisation. One day fasting robustly decreases leptinaemia and partially restores leptin responsiveness in obese mice, and fasting-induced hyperphagia and weight regain involve changes in SOCS3 expression (Pedroso et al. 2016). Fasting-induced decrease in leptinaemia is partly regulated by sympathetic outflow to the white adipose tissue regulating leptin biosynthesis and secretion (Caron et al. 2018). Energy restriction also reduces leptinaemia and restores leptin-induced pSTAT3 response in several brain regions including VMH (Morabito et al. 2017). Additionally, diet switch from HFD to regular chow reverses leptin resistance in some DIO models. In particular, decreasing the dietary fat content in mice fed with HFD for 20 weeks reverses obesity and hypothalamic leptin sensitivity in the ARH after 7 weeks of diet switch (Enriori et al. 2007). Another study showed that 12 weeks of low-fat diet after 30 weeks of DIO not only decreases body weight and leptinaemia to values similar to lean mice but also recovers leptin-induced pSTAT3 levels in some brain regions to a greater extent than energy restriction (Morabito et al. 2017). However, ARH and DMH do not recover leptin sensitivity neither with energy restriction nor with dietary switch (Morabito et al. 2017). Diet composition can also affect leptin resensitisation. When rats become leptin-resistant on HF/high-fructose diet, the removal of fructose from this diet reverses hyperleptinaemia and improves leptin sensitivity (Shapiro et al. 2008). Notably, not all studies have found that dietary manipulations restore leptin sensitivity. It is likely that type and duration of HF diet intake, degree of obesity achieved and duration of the dietary intervention impacts on the degree of leptin resensitisation. Besides, sleeve gastrectomy in obese rats induces not only body weight loss but also an improvement of leptin sensitivity, which is secondary to weight loss since pair-fed groups show similar response to leptin (Stefater et al. 2010).

Exercise also decreases body weight and leptinaemia and improves leptin sensitivity by activating leptin sensitive neurons in the VMH and increasing pSTAT3 in the VTA. This effect seems to be independent of fat mass loss (Shapiro et al. 2011, Kang et al. 2013).

Genetically modified mice provide a powerful strategy to study the reversion of obesity and leptin resistance. For instance, Pomc-knockout mice display early-onset extreme obesity; however, normal body weight can be restored if POMC is re-expressed in young reactivatable Pomc-knockout mice. If POMC re-expression is induced in aged Pomc-knockout mice, elevated body weight and leptin resistance are observed (Bumaschny et al. 2012, Chhabra et al. 2016); nevertheless, normal body weight can be restored only if POMC is re-expressed after leptinaemia is first reduced by calorie restriction. This fact suggests that a critical threshold of leptinaemia exists in order to reverse obesity (Chhabra et al. 2016).

In the case of obesity-prone rats, obesity and leptin resistance can be reversed by rearing in large litters, limiting the pups’ nutrient supply during the suckling period. Large litter rearing reduces body weight and enhances leptin sensitivity by lowering plasma leptin levels, protecting the animals from becoming obese (Patterson et al. 2010). In this obesity model brief postweaning exercise can also increase leptin sensitivity and reduce body weight (Patterson et al. 2009).

Neonatal leptin resistance induced by litter size reduction seems to partially reverse in adulthood, as exogenous leptin injection suppresses food intake, despite body weight and circulating leptin remain elevated (Sominsky et al. 2017). Interestingly, long-term energy restriction in ARH-ablated animals decreases not only
adiposity and plasma leptin but also normalises peripheral sensitivity to leptin (Perello et al. 2009). In contrast, fasted ARH-ablated animals show a reduction of leptin levels but adiposity and leptin sensitivity are not affected, suggesting that a short-term decrease of leptinaemia alone is not sufficient to restore leptin sensitivity (Perello et al. 2009). ARH-ablated animals subjected to adrenal enucleation, a strategy to transiently reduce adiposity and leptinaemia due to a reduction of glucocorticoid levels, also display a transient leptin resensitisation that ends when plasma glucocorticoid and leptin return to high levels (Perello et al. 2003). In aged animals, food restriction reduces SOCS3 expression and prevents leptin resistance (Moyse et al. 2012). Unfortunately, the molecular events mediating leptin resensitisation in these cases are poorly known.

Reversion of leptin resistance in physiological models

The transitory state of leptin resistance caused by pregnancy is spontaneously reversed once the hormonal milieu of pregnant animals returns to basal levels, and leptin sensitivity of postpartum females is normalised. Therefore, the end of pregnancy is a physiological model of leptin resensitisation. Regarding small seasonal animals, the switch from long to short day photoperiod is associated with a state of increased leptin sensitivity subsequent to reduction of SOCS3 and PTP1B following prolactin reduction (Tups et al. 2004, Tups 2009).

Leptin-sensitising compounds

Several pharmacological compounds have been reported to modulate leptin sensitivity, and some compounds naturally found in foods also act as leptin sensitisers in DIO models. Phenolic compounds (e.g., resveratrol) suppress leptin expression from adipocytes and attenuate hyperleptinaemia and leptin resistance developed in the context of early weaning or maternal HF diet exposure (Franco et al. 2016). Teasaponin and ginsenoside reduce food intake and body weight and increase hypothalamic pSTAT3 levels and Pomc expression in rodents fed HF diet (Franco et al. 2016). These effects are achieved after 3 or 4 weeks of treatment with these compounds.

Pharmacological leptin sensitisers could be broadly grouped into two categories. The first group includes small molecules that induce marginal weight loss when administered alone but increase the anorectic effect of exogenous leptin. Such compounds include meta-chlorophenylpiperazine, which is a serotonin receptor agonist and acts as a serotonin reuptake inhibitor (Yan et al. 2015), metformin, which is a commonly used anti-diabetic medication (Kim et al. 2006) and betulinic acid (Choi et al. 2013), which presumably causes leptin sensitisation through PTP1B inhibition.

The second group of molecules induce weight loss in hyperleptinaemic animals even when administered alone, suggesting that they restore leptin signalling and resensitise obese mice to their endogenous hyperleptinaemia. Such molecules include amylin, predominantly secreted from the pancreatic β cells but also produced by hypothalamic neurons (Le Foll et al. 2015), and pramlintide, an amylin analogue in clinical use for the treatment of diabetes. The mechanism by which amylin works is unclear but it seems to increase IL-6 production in VMH microglia that in turn activates pSTAT3 signalling in LepR neurons (Le Foll et al. 2015). Amylin also exerts a direct effect in enhancing leptin signalling via activation of the ERK/MAPK pathway on POMC neurons (Lutz et al. 2018). Glucagon-like peptide-1 (GLP-1) also increases central leptin action and appears to exert its anorectic effects synergistically with leptin. The acute anorectic effect of GLP-1 is attenuated in LepR-deficient rats, and GLP-1 action is also partially mediated by central induction of IL-6 production (Shirazi et al. 2013). Accordingly, IL-6 appears to be a common mediator of leptin-sensitising factors in the brain, and central or systemic overexpression of IL-6 attenuates DIO in a leptin-dependent manner. Peripherally restricted cannabinoid receptor 1 (CB1R) inverse agonists act as potent weight loss agents in DIO models and lack the psychiatric effects induced by centrally acting cannabinoid antagonists, such as rimonabant. Peripheral action of CB1R inverse agonists, such as JD5037, work in a leptin-dependent manner since they do not induce weight loss in ob/ob or db/db mice and their anti-obesity effects in DIO models are attenuated by LepRb antagonists. Notably, CB1R inverse agonists seem to act by decreasing hyperleptinaemia since their effect increasing leptin clearance and suppressing leptin production precedes weight loss. Heat shock protein 90 (HSP90) inhibitors display leptin sensitisser properties (Desarzens et al. 2014, Ozcan et al. 2017). At least some HSP90 inhibitors were shown to induce weight loss in DIO mice but had a blunted effect in ob/ob or db/db mice. Some of these HSP90 inhibitors are natural compounds, such as celastrol and gambogic acid, which have pleiotropic effects including PTP1B inhibitory and anti-inflammatory activities (Tan et al. 2017). Induction of heat shock response is proposed to exert anti-obesity effects through a peripheral mechanism that involves
adipose tissue and skeletal muscle mediated energy expenditure (Ma et al. 2015). Whether and how peripheral heat shock response couples to central leptin sensitisation is currently unknown. HSP90 inhibitors activate ER stress, and some of them also inhibit ER chaperones (Fribley et al. 2015). For example, the pentacyclic triterpene celastrol is a potent inducer of ER stress (Fribley et al. 2015). However, whether this effect is involved in the leptin sensitising role of these molecules is not known.

Concluding remarks

Experimental evidence shows that leptin resistance can be reversed, at least to some extent, in several animal models of obesity (Fig. 2). Such leptin resensitisation is associated with improvement in endocrine and metabolic disturbances commonly observed in obesity and a sustained decrease of plasma leptin levels, possibly below a critical threshold level. In some cases, leptin resensitisation is not immediate or complete, neither proportional to the decrease of body weight. At a molecular level, the recovery of leptin sensitivity shows a different timing depending on the brain region. Reversion of leptin resistance is total and immediate in physiological conditions, such as during pregnancy and the switch from long to short day photoperiod (Fig. 2). Notably, aged obese mice are less able to fully reverse the leptin-resistant states. Based upon current understandings, several pharmacological compounds are proposed to be promising therapeutic targets for the treatment of leptin resistance associated with DIO.

Declaration of Interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

Funding
This work was supported by Universidad Nacional del Litoral (CAID 2016 N° 50020150100047L) and Roemmers Foundation Research Grant (2018) to M F A, the National Agency of Scientific and Technological Promotion of Argentina (PICT2016-1084 to M P) and by the São Paulo Research Foundation (FAPESP: 17/02983-2 to J D).

Acknowledgements
The authors would like to thank the Department of Science Communication, Direction of Publishing and Special Contents of the Scientific Research Commission, Province of Buenos Aires (CIC-PBA), especially to designer Denisse A Patrault, for their kind assistance in the graphic design of the figures.

References
Leptin resensitisation


Gao Q, Wolfgang MJ, Neschen S, Morino K, Horvath TL, Shulman GI & Fu XY 2004 Disruption of neural signal transducer and activator of transcription 3 causes obesity, diabetes, infertility, and thermal
dysregulation. PNAS 101 4661–4666. (https://doi.org/10.1073/pnas.0303992101)


Halas JL, Boozer C, Blair-West J, Fidahusein N, Denton DA & Friedman JM 1997 Physiological response to long-term peripheral and central leptin infusion in lean and obese mice. PNAS 94 8878–8883. (https://doi.org/10.1073/pnas.94.16.8878)


Haynes WG, Morgan DA, Walsh SA, Mark AL & Sivitz WI 1997 Receptor-mediated regional sympathetic nerve activation by leptin. Journal of Clinical Investigation 100 270–278. (https://doi.org/10.1172/JCI91532)


Hileman SM, Pierroz DD, Masuzaki H, Bjorbaek C, El-Haschimi K, Banks WA & Flier JS 2002 Characterization of short isoforms of the leptin receptor in rat cerebral microvessels and of brain uptake of leptin in mouse models of obesity. Endocrinology 143 775–783. (https://doi.org/10.1210/endo.143.3.8669)


Perello M & Raingo J 2013 Leptin activates oxytoxin neurons of the hypothalamic paraventricular nucleus in both control and diet-induced obese rodents. *Plos ONE* **8** e69625. (https://doi.org/10.1371/journal.pone.0069625)


Perello M & Raininger J 2013 Leptin activates oxytoxin neurons of the hypothalamic paraventricular nucleus in both control and diet-induced obese rodents. *Plos ONE* **8** e69625. (https://doi.org/10.1371/journal.pone.0069625)


Perello M, Cakir I, Cakir I & Cakir S 2013 Leptin activates oxytoxin neurons of the hypothalamic paraventricular nucleus in both control and diet-induced obese rodents. *Plos ONE* **8** e69625. (https://doi.org/10.1371/journal.pone.0069625)


Perello M, Cakir I, Cakir I & Cakir S 2013 Leptin activates oxytoxin neurons of the hypothalamic paraventricular nucleus in both control and diet-induced obese rodents. *Plos ONE* **8** e69625. (https://doi.org/10.1371/journal.pone.0069625)


Simonds SE, Pryor JT & Cowley MA 2017 Does leptin cause an increase in blood pressure in animals and humans? *Current Opinion in Nephrology and Hypertension* **26** 20–25. ([https://doi.org/10.1097/MNH.0000000000000287](https://doi.org/10.1097/MNH.0000000000000287))


St-Pierre J & Tremblay ML 2012 Obesity is associated with hypothalamic injury in rodents and humans. *Journal of Clinical Investigation* **122** 153–162. ([https://doi.org/10.1172/JCI59660](https://doi.org/10.1172/JCI59660))


