

Low-dose PTH increases osteoblast activity via decreased *Mef2c/Sost* in senescent osteopenic mice

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Abstract

Intermittent administration of parathyroid hormone (PTH) 1–34 at a standard dose has been shown to induce anabolic effects in bone. However, whether low-dose PTH promotes bone formation during senescence is unknown. To address this issue, we determined the effects of low-dose PTH and analysed the underlying mechanisms in prematurely senescent mice that display osteopenia. Treatment of 9-week-old *Samp6* mice for 6 weeks with PTH at a standard dose (100 µg/kg per day) increased vertebral and femoral bone mass and improved bone microarchitecture as a result of increased bone-forming surfaces and mineral apposition rate (MAR). At a tenfold lower dose (10 µg/kg per day), PTH increased axial bone volume and trabecular thickness, as detected by bone histomorphometry but not by micro-computed tomography analysis. This anabolic effect resulted from increased osteoblast activity, as reflected by increased serum N-terminal propeptide of type 1 procollagen (P1NP) levels and MAR, with unchanged bone-forming surface or osteoblast surface. Mechanistically, low-dose PTH increased the expression of osteoblast markers in bone marrow stromal cells and mature osteoblasts, which was associated with increased expression of the Wnt effector *Wisp1*. Moreover, low-dose PTH decreased the expression of the *Mef2c* transcription factor, resulting in decreased *Sost* expression in osteoblasts/osteocytes. These results indicate that PTH at a low dose is effective at promoting bone formation and increased bone volume in senescent osteopenic mice through increased osteoblast activity and modulation of specific Wnt effectors, which raises the potential therapeutic use of intermittent PTH at low dose to increase bone forming activity and bone mass in skeletal senescence.

Key Words

- ▶ low-dose PTH
- ▶ senescent mice
- ▶ osteoblast activity
- ▶ Wnt signalling

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Introduction

Bone loss associated with ageing is characterised by decreased bone formation relative to bone resorption, resulting in altered bone microarchitecture, osteoporosis and increased risk of fractures (Khosla *et al.* 2011). The age-related decline in bone formation results from multiple intrinsic and extrinsic mechanisms that lead to decreased differentiation of bone marrow stromal cells (BMSCs) into osteoblasts and

decreased osteoblast number and activity (Manolagas & Parfitt 2010, Kassem & Marie 2011). One important challenge is therefore to improve the number and functioning of osteoblasts in order to prevent the decrease in bone formation and bone mass in skeletal senescence. Intermittent parathyroid hormone (PTH) administration is the only proved strategy for inducing an anabolic effect that is

available in clinics for the treatment of severe osteoporosis. This treatment has been shown to increase bone formation to a greater extent than bone resorption in animals and humans, resulting in improved bone mass and microarchitecture (Compston 2007). In adult mice, the anabolic effect of PTH on bone is mediated by increased numbers and activity of osteoblasts (Canalis *et al.* 2007, Jilka 2007). The PTH-induced increase in osteoblast number in adult mice is achieved by increased osteoblast differentiation, by converting quiescent lining cells to active osteoblasts and by reducing osteoblast apoptosis (Jilka 2007, Kim *et al.* 2012). The anabolic effect of PTH is mediated through direct and indirect mechanisms. PTH signals through G-protein-coupled type 1 PTH/PTH-related peptide receptor and subsequent activation of protein kinase A, protein kinase C and other pathways that control osteoblastogenesis (Jilka 2007). In addition, PTH interacts with fibroblast growth factor 2 (Fei *et al.* 2013) and insulin-like growth factor 1 signalling (Canalis *et al.* 1989) promoting bone formation, which shows that the anabolic effect of PTH is mediated in part through interaction with other signalling pathways.

Wnt signalling is an important regulator of osteoblast proliferation, differentiation and survival (Baron & Kneissel 2013). The Wnt canonical pathway involves Wnt binding to the co-receptors LRP5 and Frizzled, leading to the inhibition of glycogen synthase kinase 3 β and decreased phosphorylation, stabilisation and subsequent translocation of β -catenin into the nucleus. This results in the binding of β -catenin to T-cell-specific transcription factor/lymphoid enhancer-binding factor 1 transcription factor and activation of target genes (Clevers 2006). The Wnt canonical pathway is negatively regulated by extracellular antagonists such as soluble Frizzled-related proteins (sFRPs), Dickkopf (DKK) and sclerostin that interact with Wnt proteins or Wnt signalling partners to antagonise Wnt signalling (Kawano & Kypta 2003, Bodine *et al.* 2004, Yao *et al.* 2010). Results from several studies have indicated that PTH may interact with Wnt signalling (Kulkarni *et al.* 2005). Specifically, PTH upregulates Wnt-responsive genes in osteoblastic cells (Qin *et al.* 2003, Tobimatsu *et al.* 2006), and PTH1R interacts with the co-receptor LRP6 to increase β -catenin (Wan *et al.* 2008). PTH may also promote Wnt/ β -catenin signalling by inhibiting the Wnt antagonists sFRP1 and DKK1 (Bellido *et al.* 2005, Bodine *et al.* 2007, Guo *et al.* 2010). In addition, PTH decreases the expression of *Sost* in osteocytes (Bellido *et al.* 2005, Keller & Kneissel 2005). *Sost* encodes sclerostin, which binds to the Wnt co-receptors LRP4/5/6 and thereby antagonises Wnt/ β -catenin signalling (van Bezooijen *et al.* 2004, Li *et al.* 2005).

To date, only a few studies have analysed the anabolic effect of PTH in the ageing skeleton (Knopp *et al.* 2005, Brennan *et al.* 2009, Jilka *et al.* 2010). Moreover, the effect of PTH 1–34 in osteopenic mice has been tested at standard doses ranging from 80 to 100 μ g/kg per day, and it is not known whether PTH at a much lower dose (i.e. a tenfold lower dose) may be effective at promoting bone formation in senescent mice. To address this issue, we determined the skeletal response to low-dose PTH in prematurely senescent osteopenic mice and analysed the cellular and molecular mechanisms involved in this response. We show herein that intermittent PTH at a low dose is effective at increasing axial bone mass in prematurely senescent osteopenic mice by promoting osteoblast activity, through modulation of specific Wnt effectors.

Materials and methods

Animals and treatment

We used *Samp6* mice, a murine model of ageing in the P6 strain of senescence-accelerated mice which is characterised by decreased bone formation and low bone mass (Jilka *et al.* 1996, Clement-Lacroix *et al.* 2005). Seven-week-old *Samp6* mice (Harlan Laboratories, Derby, UK) were fed with mouse standard diet (A04 rat/mouse diet, SAFE, Augy, France) containing calcium (0.84%), phosphorus (0.57%) and vitamin D (1000 IU/kg), and were weighed once a week. Nine-week-old mice (eight mice per group) were treated with s.c. human PTH 1–34 at a high standard dose (100 μ g/kg BW) or a low dose (10 μ g/kg BW) (Sigma) or the vehicle, 5 days/week for 6 weeks. The rationale for choosing the low dose is that it is tenfold lower than the dose usually used in osteopenic mice (Knopp *et al.* 2005, Jilka *et al.* 2010, Hanyu *et al.* 2012, Kim *et al.* 2012). To label bone mineralisation fronts, control and treated mice were given tetracycline (20 mg/kg, Sigma) and calcein (10 mg/kg) by s.c. injection, respectively, at days 8 and 3 before being killed by injection of ketamine/xylazine. The protocol was conducted according to the guidelines of the Local Ethical Committee (ref. no. CEEALV/2011.11.01).

P1NP analysis

The animals were killed and blood was collected and spun in order to recuperate the serum. Serum aliquots were frozen before analysis of N-terminal propeptide of type 1 procollagen (P1NP) levels, an established marker of bone formation, by ELISA (R&D Systems, Lille, France).

Bone microarchitecture and histomorphometry

The animals were killed and, lumbar vertebrae and right femurs (distal metaphysis) were obtained for analysis of micro- and macro-structures. The bones were scanned using a high-resolution micro-computed tomography (microCT) system (SkyScan 1172, MicroPhotonics, Allentown, PA, USA) and analysed using a 3D morphometry evaluation program (NRecon reconstruction program). For histomorphometric analysis, the bones were embedded in methylmethacrylate and 5 μm sections were stained with aniline blue to analyse structural parameters (osteoblast surface, bone volume, trabecular number and thickness), as described previously (Haÿ *et al.* 2009). TRAP staining was carried out to evaluate the number of active osteoclasts (Haÿ *et al.* 2009). Unstained sections (8 μm thick) were used to assess dynamic parameters (mineral apposition rate (MAR), double labelled surface and bone formation rate (BFR); (Parfitt *et al.* 1987).

Quantitative PCR analysis

The animals were killed and the bone marrow was flushed from the tibia, yielding BMSCs and remaining bone containing mature osteoblasts and osteocytes. Total RNA was extracted from the two bone cell pools, and 1 μg of total RNA from each sample was reverse-transcribed using the Applied Biosystems Kit (High-Capacity cDNA RT Kit). The relative mRNA levels of osteoblast differentiation markers and Wnt effectors were evaluated by quantitative PCR analysis (LightCycler; Roche Applied Science) using a SYBR Green PCR Kit (ABGen, Courtabœuf, France) and specific primers (Haÿ *et al.* 2009, Andrews *et al.* 2012, Saidak *et al.* 2012). The signals were normalised to hypoxanthine phosphoribosyltransferase (HRPT) as an internal control.

Statistical analysis

Values are presented as the mean \pm S.E.M. of six to eight animal groups. Data were analysed with the unpaired two-tailed Student's *t*-test. A *P* value <0.05 was considered statistically significant.

Results

Low-dose PTH increases bone volume in senescent mice

We first compared the effect of the two doses of PTH on trabecular bone mass and microarchitecture as evaluated

by microCT analysis. As it might be expected, intermittent PTH at the standard dose increased vertebral bone mass and volume in *Samp6* mice (Fig. 1A and B). This effect resulted from increased trabecular thickness and number and decreased trabecular separation (Fig. 1C, D and E). Interestingly, the low dose of PTH was sufficient to increase trabecular bone thickness as evaluated by microCT analysis (Fig. 1C). To further investigate the anabolic effect of PTH at low doses, we performed a histomorphometric analysis of vertebral bone. We found that PTH at a low dose increased trabecular bone volume

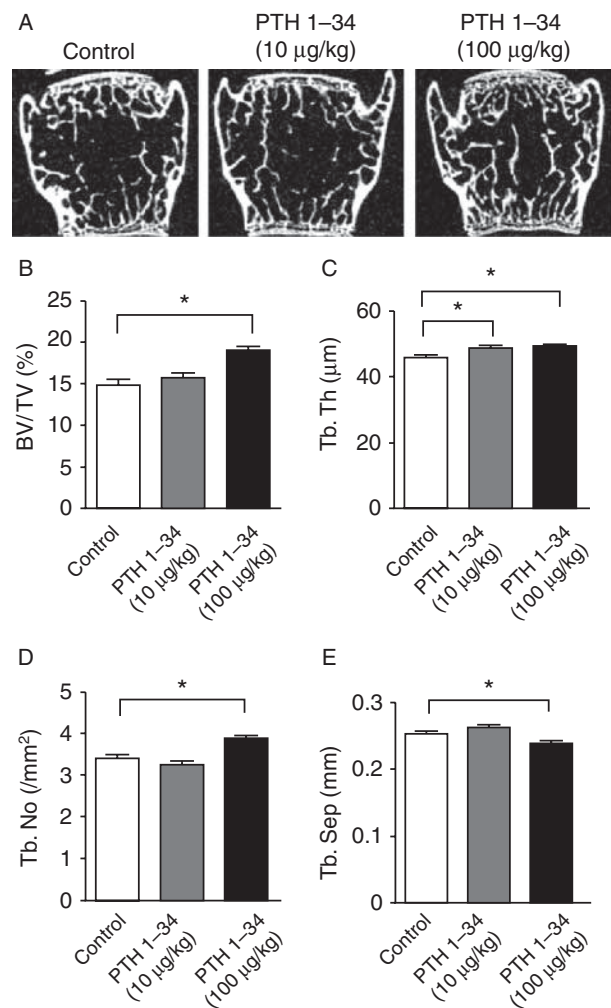


Figure 1

Distinct effects of standard and low-dose intermittent PTH on bone mass and microarchitecture in prematurely senescent osteopenic *Samp6* mice. Representative microCT images of lumbar vertebrae (A) showing that both doses of PTH increased bone mass in *Samp6* mice. (B, C, D and E) Analysis of bone microarchitecture parameters showing the distinct effects of PTH at the standard and low doses on bone volume (BV, expressed relative to trabecular volume (TV)), trabecular thickness (Tb. Th), trabecular number (Tb. No) and trabecular separation (Tb. Sep). Mean \pm S.D. of six to eight mice. *Statistically significant ($P < 0.05$).

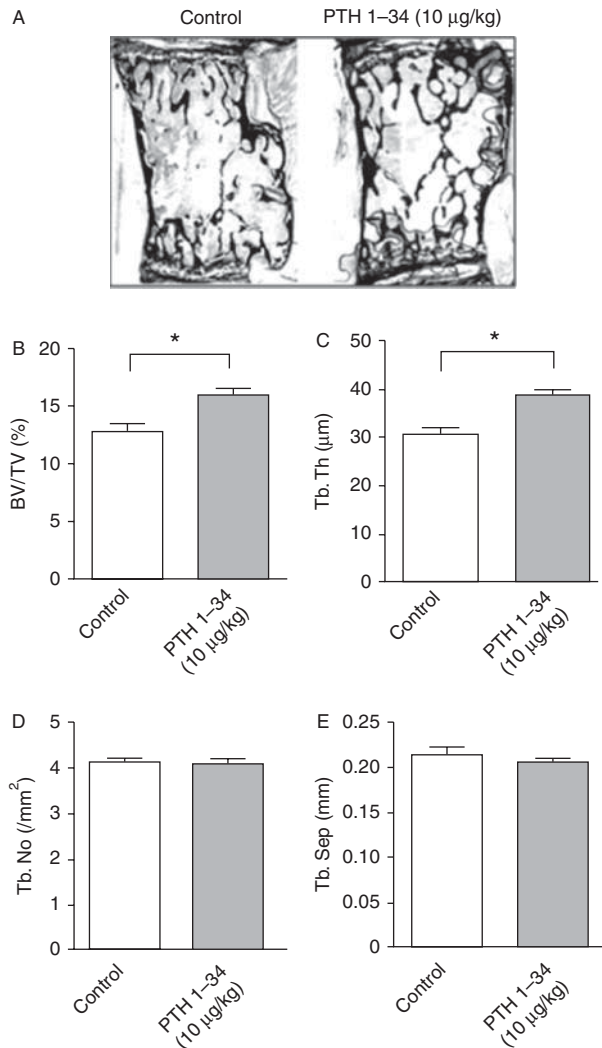


Figure 2

Histomorphometric analysis showing that intermittent low-dose PTH increased trabecular bone volume (A and B) and trabecular bone thickness (C) but not trabecular number (D) or separation (E) in vertebral bone in prematurely senescent osteopenic *Samp6* mice. Mean \pm s.d. of six to eight mice. *Statistically significant ($P < 0.05$).

in vertebrae (Fig. 2A and B) as a result of increased trabecular bone thickness (Fig. 2C), with no significant change in trabecular number or separation (Fig. 2D and E). The positive effect on bone volume was restricted to the axial bone because the standard dose, but not the low dose, increased bone volume and trabecular thickness and number and decreased trabecular separation in the femur (Supplementary Fig. 1A, B, C and D, see section on supplementary data given at the end of this article). The increased axial bone volume induced by low-dose PTH was not related to changes in bone resorption, because the trabecular separation was not affected (Figs 1 and 2)

and the number of TRAP⁺ osteoclasts was unchanged (Supplementary Fig. 2A). These results indicate that PTH at a low dose is effective at increasing trabecular bone thickness and bone volume in the axial skeleton in senescent osteopenic *Samp6* mice.

Low-dose PTH increases osteoblast function but not number in senescent mice

To determine whether the low-dose PTH increased bone mass by increasing bone formation, we analysed the levels of serum P1NP, an established marker of bone-forming activity. Both the standard-dose and the low-dose PTH increased serum P1NP levels, indicating an anabolic response at both doses (Fig. 3). To confirm that the low dose of PTH was efficient at promoting trabecular bone formation, histomorphometric analysis was performed in axial bone. PTH at low dose had no effect on the extent of double-labelled surface (Fig. 4A). Consistently, the osteoblast surface, which reflects osteoblast number, was unchanged (Supplementary Fig. 2B). In contrast, the low-dose PTH increased the bone MAR, which reflects the bone-forming activity (Fig. 4B) without change in the BFR (Fig. 4C). These effects were not restricted to axial bone because similar results were obtained in the femur (Supplementary Fig. 3, see section on supplementary data given at the end of this article). In contrast, the standard high-dose PTH increased bone MAR, double-labelled surface and BFR by twofold (Supplementary Fig. 3). These results indicate that PTH at a low dose is effective at increasing bone formation in senescent osteopenic mice by increasing the activity but not the number of active osteoblasts.

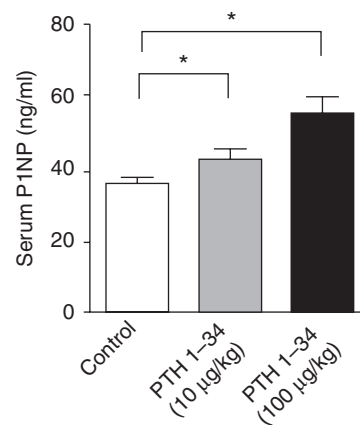
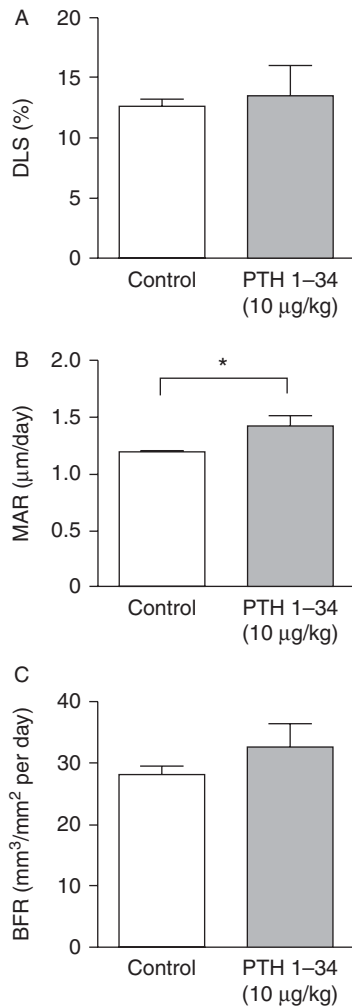


Figure 3

Intermittent PTH at low and standard doses increased serum P1NP levels in prematurely senescent osteopenic *Samp6* mice after 6 weeks of treatment. Mean \pm s.d. of six to eight mice. *Statistically significant ($P < 0.05$).

**Figure 4**

Intermittent low-dose PTH increased osteoblast activity but not bone forming surface in the vertebrae of prematurely senescent osteopenic *Samp6* mice. Low-dose PTH had no effect double-labelled surface (DLS) (A) or bone formation rate (BFR) (C) but increased the mineral apposition rate (MAR) (B). Mean \pm s.d. of six to eight mice. *Statistically significant ($P < 0.05$).

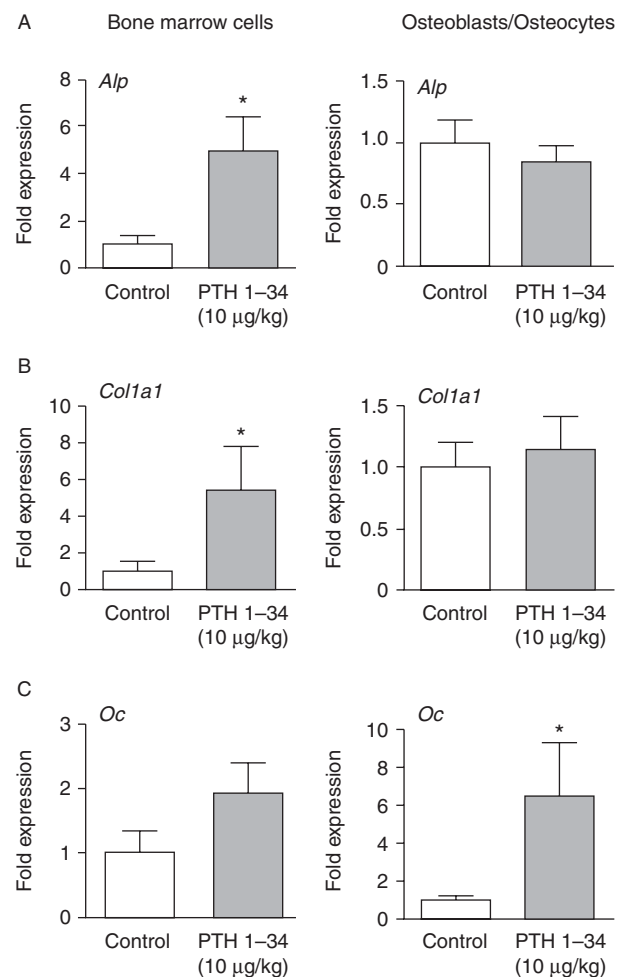
Low-dose PTH increases functional osteoblast markers in senescent mice

To determine the mechanisms by which the low-dose PTH increased osteoblast activity, we performed a molecular analysis of genes that characterise osteoblast function in osteoprogenitor cells present in the bone marrow stroma. At a low dose, PTH was effective at increasing the expression of alkaline phosphatase (*Alp*) and type 1 collagen (*Col1a1*) and tended to increase the expression of osteocalcin (*Oc*), a marker of mature osteoblasts (Fig. 5A–C). In the bone-marrow-free tibia that contains mature osteoblasts and osteocytes, *Alp* and *Col1a1* levels were not affected by low-dose PTH, whereas *Oc* was

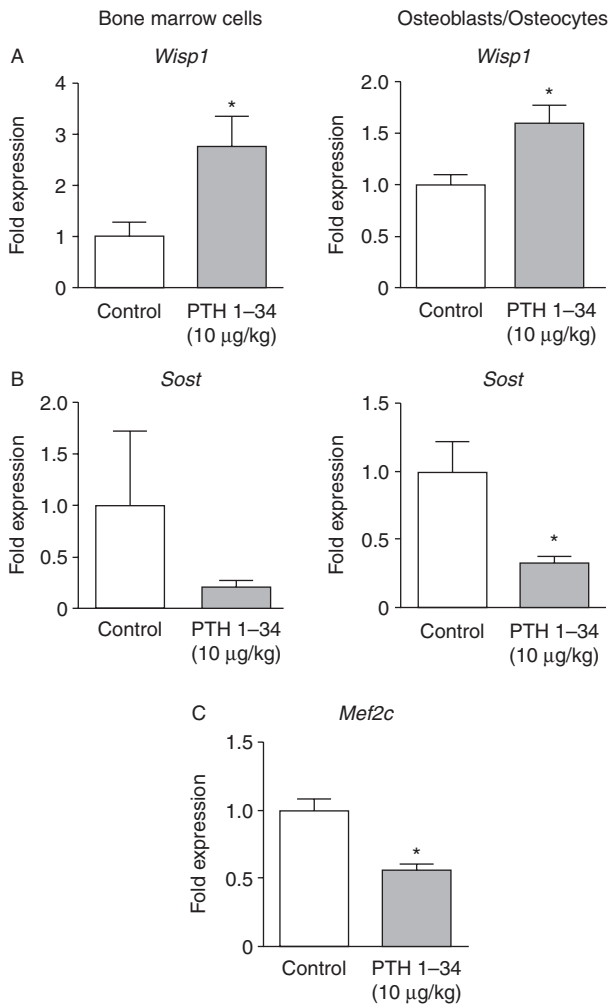
markedly increased (Fig. 5A–C). These results indicate that PTH at low dose acted by promoting osteoblast function in *Samp6* mice.

Low-dose PTH modulates specific Wnt effectors in senescent mice

We next determined the molecular mechanisms underlying the positive effect of the low-dose PTH on osteoblast activity in *Samp6* mice by determining the changes in direct Wnt effectors (Clevers 2006, Gordon & Nusse 2006). We first analysed the effect of low-dose PTH on

**Figure 5**

Intermittent low-dose PTH increased expression of the osteoblast differentiation markers *Alp* (A), *Col1a1* (B) and *Oc* (C) in prematurely senescent osteopenic *Samp6* mice. Quantitative RT-PCR analysis of osteoblast gene expression was carried out in the bone marrow stromal cells and osteoblasts/osteocytes extracted from long bones in control *Samp6* mice or mice treated with low-dose PTH for 6 weeks. The transcript levels were normalized to the values for HPRT. Mean \pm s.d. of six to eight mice. *Statistically significant ($P < 0.05$).

**Figure 6**

Intermittent low-dose PTH modulated Wnt signalling effectors *Wisp1* (A), *Sost* (B) and *Mef2c* (C) in prematurely senescent osteopenic *Samp6* mice. Quantitative RT-PCR analysis of Wnt effectors was carried out using bone marrow stromal cells and osteoblasts/osteocytes extracted from long bones (or pooled populations for *Mef2c*) of control *Samp6* mice or mice treated with PTH at a low dose for 6 weeks. The transcript levels were normalized to the values for HPRT. Mean \pm s.d. of six to eight mice. *Statistically significant ($P < 0.05$).

Wnt-induced secreted protein 1 (*Wisp1*) because it is a direct target of canonical Wnt signalling (Ono *et al.* 2011). The low-dose PTH increased *Wisp1* mRNA levels in both BMSCs and osteoblasts/osteocytes, indicating that Wnt signalling was activated in both osteoblast precursor cells and more mature osteoblasts/osteocytes (Fig. 6A).

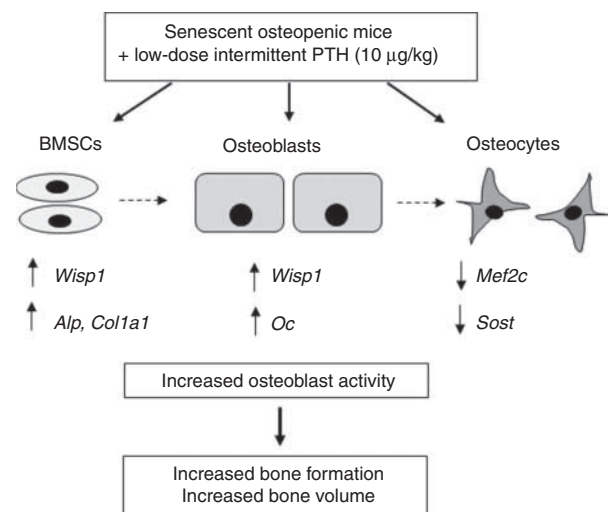
Wnt signalling is negatively regulated by inhibitors such as DKK1 and sFRP1 (Bodine *et al.* 2004). To investigate whether low-dose PTH acts by downregulating these Wnt inhibitors, we analysed the changes in *Dkk1* and *sFrp1* gene expression in bones of *Samp6* mice. We found that low-dose

PTH increased the expression of *Dkk1* in both BMSCs and osteoblasts/osteocytes (Supplementary Fig. 4A, see section on supplementary data given at the end of this article). In contrast, *sFrp1* mRNA levels remained unchanged (Supplementary Fig. 4B). This indicates that these Wnt antagonists were distinctly affected by low-dose PTH in *Samp6* mice. Importantly, we found that *Sost* mRNA levels tended to be reduced in BMSCs and were greatly reduced by PTH at a low dose in osteoblasts/osteocytes (Fig. 6B). To determine how the low-dose PTH may affect *Sost* expression in *Samp6* mice, we analysed the expression of *Mef2c* which upregulates *Sost* expression (Leupin *et al.* 2007, Kramer *et al.* 2010) and is regulated by PTH *in vitro* (Bonnet *et al.* 2012). We found that *Mef2c* in bones was decreased by low-dose PTH, which provides a mechanism by which low-dose PTH acts on *Sost* expression and promotes osteoblast activity in osteopenic senescent *Samp6* mice (Fig. 6C).

Overall, these results provide the first evidence that low-dose PTH treatment in *Samp6* mice is effective for increasing bone mass in the axial skeleton by increasing osteoblast activity, but not cell number, via upregulation of *Wisp1* and downregulation of *Mef2c*/*Sost* in bone (Fig. 7).

Discussion

Intermittent PTH at a standard dose is known to produce an anabolic effect on bone in adult animals. However, it is not known whether a low dose (i.e. tenfold lower than the

**Figure 7**

Proposed mechanism of the anabolic effect of low-dose PTH in prematurely senescent osteopenic *Samp6* mice. Low-dose PTH increased osteoblast activity, but not number, by modulating specific Wnt effectors, resulting in increased bone formation and axial bone volume in senescent osteopenic *Samp6* mice.

usual dose used) is effective in osteopenic ageing mice. In this study, we showed that low-dose PTH is efficient at increasing bone formation and axial bone volume in senescent osteopenic mice, and that this anabolic effect results from increased osteoblast activity and modulation of specific Wnt effectors. First, we showed that PTH treatment at a low dose increased trabecular bone thickness and volume in axial bone in *Samp6* mice, demonstrating that a low dose of PTH is sufficient to augment bone mass in senescent osteopenic mice. This effect was evidenced by histomorphometric but not microCT analysis. This discrepancy is probably due to the distinct resolution between the two techniques. Second, the results indicate that the increased bone volume induced by low-dose PTH results from increased osteoblast activity, as shown by the increased PINP levels and MAR. In contrast, the osteoblast surface and double-labelled surfaces were unchanged, indicating that low-dose PTH acted by increasing osteoblast activity, but not the number of active osteoblasts, in *Samp6* mice. This is in contrast with the effect produced by PTH at a standard dose, which increased the double-labelled bone forming surface in *Samp6* mice (this study), as has been found in WT ageing mice (Jilka 2007). These results support the notion that PTH at a low dose is effective at increasing bone volume in senescent osteopenic mice through enhanced osteoblast activity.

In WT mice, the anabolic effect of PTH at a standard dose results from activation of multiple signals (Canalis *et al.* 2007, Jilka 2007). In prematurely senescent osteopenic mice low-dose PTH increased the expression of *Alp* and *Col1a1*, which are targets of Wnt signalling (Gaur *et al.* 2005, Bodine & Komm 2006). Consistent with this finding, low-dose PTH increased the expression of *Wisp1*, a direct target of Wnt signalling, as has been also previously reported for WT mice treated with a standard dose of PTH (Jilka *et al.* 2010). WISP1 is a member of the CCN family that is upregulated by β -catenin (Xu *et al.* 2000) and that positively regulates bone formation (French *et al.* 2004, Ono *et al.* 2011). Our findings that low-dose PTH increased *Wisp1* expression in both BMSCs and osteoblasts/osteocytes therefore reveals one mechanism for the anabolic effect of low-dose PTH in senescent mice. We also found that low-dose PTH increased the expression of *Dkk1* in BMSCs and osteoblasts/osteocytes. This is surprising because DKK1 antagonises canonical Wnt signalling *in vivo* by binding to Wnt ligands, leading to the attenuation of Wnt/receptor activation and bone formation (Bovolenta *et al.* 2008). The increased expression of *Dkk1* by low-dose PTH

is likely to result secondarily from the activation of Wnt signalling which is known to increase *Dkk1* expression (Chamorro *et al.* 2005). This is consistent with a negative-feedback response restricting the exposure of bone cells to prolonged activation of Wnt signalling. The induction of *Dkk1* expression is unlikely to reduce the overall response to low-dose PTH in senescent *Samp6* mice because, although the suppression of Wnt signalling by DKK1 was found to attenuate PTH-mediated stromal cell response and new bone formation (Guo *et al.* 2010), targeted overexpression of *Dkk1* in osteoblasts does not impair the anabolic response to PTH in mice (Yao *et al.* 2011). In contrast to *Dkk1*, the level of *sFrp1*, a negative regulator of bone formation (Yao *et al.* 2010), was unchanged by low-dose PTH in *Samp6* mice. Results from previous studies have indicated that the bone anabolic effect of PTH is attenuated by either overexpression or deletion of *sFrp1* in mice (Bodine *et al.* 2007). The lack of change in *sFrp1* expression in PTH-treated *Samp6* senescent mice is thus unlikely to affect the anabolic effect of low-dose PTH in these mice.

Our finding that PTH at a low dose markedly decreased *Sost* levels in osteoblasts/osteocytes provides another mechanism for the anabolic effect of low-dose PTH in senescent *Samp6* mice. The decreased *Sost* level with a low-dose of PTH in *Samp6* mice is consistent with the effect of PTH at a standard dose in WT mice (Bellido *et al.* 2005, Keller & Kneissel 2005). *Sost* expression in osteocytes has been shown to be positively regulated by the transcription factor MEF2C (Leupin *et al.* 2007). Recent results have indicated that *Mef2c* invalidation results in reduced *Sost* expression and increased bone mass in mice (Kramer *et al.* 2010, 2012, Collette *et al.* 2012). In this study, we found that PTH at a low dose decreased *Mef2c* levels which was associated with decreased *Sost* levels, increased osteoblast activity and bone volume in senescent osteopenic *Samp6* mice. Our data thus support the notion that the anabolic response to low-dose PTH involves downregulation of *Mef2c* and subsequent reduction of *Sost*, leading to increased Wnt/ β -catenin signalling, osteoblast activity and bone mass in senescent osteopenic mice (Fig. 7). The finding that PTH at low dose is able to modulate specific Wnt signalling effectors in *Samp6* mice is particularly important in the context of skeletal senescence because results from current studies indicate that bone ageing is associated with alterations in the local expression of Wnt ligands (Rauner *et al.* 2008) among other mechanisms (Manolagas & Parfitt 2010, Marie 2014). We previously reported that the anti-osteoporotic agent strontium ranelate promotes Wnt signalling in osteoblasts and

thereby increases bone formation and bone mass in senescent osteopenic *Samp6* mice (Saidak *et al.* 2012). The results of the present study support the interest in targeting Wnt signalling for improving osteoblast activity and bone mass in skeletal ageing.

In summary, our results reveal that PTH at a low dose is effective at increasing bone volume in senescent osteopenic mice through activation of osteoblast activity and modulation of specific Wnt effectors, which supports the potential therapeutic interest in using intermittent PTH at a low dose for improving bone formation and bone mass in skeletal senescence.

Supplementary data

This is linked to the online version of the paper at <http://dx.doi.org/10.1530/JOE-14-0249>.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Author contribution statement

Z S and P J M designed the experiments, Z S, C L H, S A and C M performed the experiments and Z S and P J M wrote the manuscript.

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References

- Andrews SF, Dai X, Ryu BY, Gulick T, Ramachandran B & Rawlings DJ 2012 Developmentally regulated expression of MEF2C limits the response to BCR engagement in transitional B cells. *European Journal of Immunology* **42** 1327–1336. (doi:10.1002/eji.201142226)
- Baron R & Kneissel M 2013 WNT signaling in bone homeostasis and disease: from human mutations to treatments. *Nature Medicine* **19** 179–192. (doi:10.1038/nm.3074)
- Bellido T, Ali AA, Gubrij I, Plotkin LI, Fu Q, O'Brien CA, Manolagas SC & Jilka RL 2005 Chronic elevation of parathyroid hormone in mice reduces expression of sclerostin by osteocytes: a novel mechanism for hormonal control of osteoblastogenesis. *Endocrinology* **146** 4577–4583. (doi:10.1210/en.2005-0239)
- van Bezooijen RL, Roelen BA, Visser A, van der Wee-Pals L, de Wilt E, Karperien M, Hamersma H, Papapoulos SE, ten Dijke P & Lowik CW 2004 Sclerostin is an osteocyte-expressed negative regulator of bone formation, but not a classical BMP antagonist. *Journal of Experimental Medicine* **199** 805–814. (doi:10.1084/jem.20031454)
- Bodine PV & Komm BS 2006 Wnt signaling and osteoblastogenesis. *Reviews in Endocrine & Metabolic Disorders* **7** 33–39. (doi:10.1007/s11154-006-9002-4)
- Bodine PV, Zhao W, Kharode YP, Bex FJ, Lambert AJ, Goad MB, Gaur T, Stein GS, Lian JB & Komm BS 2004 The Wnt antagonist secreted Frizzled-related protein-1 is a negative regulator of trabecular bone formation in adult mice. *Molecular Endocrinology* **18** 1222–1237. (doi:10.1210/me.2003-0498)
- Bodine PV, Seestaller-Wehr L, Kharode YP, Bex FJ & Komm BS 2007 Bone anabolic effects of parathyroid hormone are blunted by deletion of the Wnt antagonist secreted Frizzled-related protein-1. *Journal of Cellular Physiology* **210** 352–357. (doi:10.1002/jcp.20834)
- Bonnet N, Conway SJ & Ferrari SL 2012 Regulation of β catenin signaling and parathyroid hormone anabolic effects in bone by the matricellular protein periostin. *PNAS* **109** 15048–15053. (doi:10.1073/pnas.1203085109)
- Bovolenta P, Esteve P, Ruiz JM, Cisneros E & Lopez-Rios J 2008 Beyond Wnt inhibition: new functions of secreted Frizzled-related proteins in development and disease. *Journal of Cell Science* **121** 737–746. (doi:10.1242/jcs.026096)
- Brennan TC, Rizzoli R & Ammann P 2009 Selective modification of bone quality by PTH, pamidronate, or raloxifene. *Journal of Bone and Mineral Research* **24** 800–808. (doi:10.1359/jbmr.081227)
- Canalis E, Centrella M, Burch W & McCarthy TL 1989 Insulin-like growth factor I mediates selective anabolic effects of parathyroid hormone in bone cultures. *Journal of Clinical Investigation* **83** 60–65. (doi:10.1172/JCI113885)
- Canalis E, Giustina A & Bilezikian JP 2007 Mechanisms of anabolic therapies for osteoporosis. *New England Journal of Medicine* **357** 905–916. (doi:10.1056/NEJMr067395)
- Chamorro MN, Schwartz DR, Vonica A, Brivanlou AH, Cho KR & Varmus HE 2005 FGF-20 and DKK1 are transcriptional targets of β -catenin and FGF-20 is implicated in cancer and development. *EMBO Journal* **24** 73–84. (doi:10.1038/sj.emboj.7600460)
- Clement-Lacroix P, Ai M, Morvan F, Roman-Roman S, Vayssiere B, Belleville C, Estrera K, Warman ML, Baron R & Rawadi G 2005 Lrp5-independent activation of Wnt signaling by lithium chloride increases bone formation and bone mass in mice. *PNAS* **102** 17406–17411. (doi:10.1073/pnas.0505259102)
- Clevers H 2006 Wnt/ β -catenin signaling in development and disease. *Cell* **127** 469–480. (doi:10.1016/j.cell.2006.10.018)
- Collette NM, Genetos DC, Economides AN, Xie L, Shahnazari M, Yao W, Lane NE, Harland RM & Loots GG 2012 Targeted deletion of *Sost* distal enhancer increases bone formation and bone mass. *PNAS* **109** 14092–14097. (doi:10.1073/pnas.1207188109)
- Compston JE 2007 Skeletal actions of intermittent parathyroid hormone: effects on bone remodelling and structure. *Bone* **40** 1447–1452. (doi:10.1016/j.bone.2006.09.008)
- Fei Y, Gronowicz G & Hurley MM 2013 Fibroblast growth factor-2, bone homeostasis and fracture repair. *Current Pharmaceutical Design* **19** 3354–3363. (doi:10.2174/1381612811319190002)
- French DM, Kaul RJ, D'Souza AL, Crowley CW, Bao M, Frantz GD, Filvaroff EH & Desnoyers L 2004 WISP-1 is an osteoblastic regulator expressed during skeletal development and fracture repair. *American Journal of Pathology* **165** 855–867. (doi:10.1016/S0002-9440(10)63348-2)
- Gaur T, Lengner CJ, Hovhannisyan H, Bhat RA, Bodine PV, Komm BS, Javed A, van Wijnen AJ, Stein JL, Stein GS *et al.* 2005 Canonical WNT signaling promotes osteogenesis by directly stimulating *Rumx2* gene expression. *Journal of Biological Chemistry* **280** 33132–33140. (doi:10.1074/jbc.M500608200)
- Gordon MD & Nusse R 2006 Wnt signaling: multiple pathways, multiple receptors, and multiple transcription factors. *Journal of Biological Chemistry* **281** 22429–22433. (doi:10.1074/jbc.R600015200)

- Guo J, Liu M, Yang D, Bouxsein ML, Saito H, Galvin RJ, Kuhstoss SA, Thomas CC, Schipani E, Baron R *et al.* 2010 Suppression of Wnt signaling by Dkk1 attenuates PTH-mediated stromal cell response and new bone formation. *Cell Metabolism* **11** 161–171. (doi:10.1016/j.cmet.2009.12.007)
- Hanyu R, Wehbi VL, Hayata T, Moriya S, Feinstein TN, Ezura Y, Nagao M, Saita Y, Hemmi H, Notomi T *et al.* 2012 Anabolic action of parathyroid hormone regulated by the β^2 -adrenergic receptor. *PNAS* **109** 7433–7438. (doi:10.1073/pnas.1109036109)
- Hayé E, Laplantine E, Geoffroy V, Frain M, Kohler T, Muller R & Marie PJ 2009 N-cadherin interacts with axin and LRP5 to negatively regulate Wnt/ β -catenin signaling, osteoblast function, and bone formation. *Molecular and Cellular Biology* **29** 953–964. (doi:10.1128/MCB.00349-08)
- Jilka RL 2007 Molecular and cellular mechanisms of the anabolic effect of intermittent PTH. *Bone* **40** 1434–1446. (doi:10.1016/j.bone.2007.03.017)
- Jilka RL, Weinstein RS, Takahashi K, Parfitt AM & Manolagas SC 1996 Linkage of decreased bone mass with impaired osteoblastogenesis in a murine model of accelerated senescence. *Journal of Clinical Investigation* **97** 1732–1740. (doi:10.1172/JCI118600)
- Jilka RL, Almeida M, Ambrogini E, Han L, Roberson PK, Weinstein RS & Manolagas SC 2010 Decreased oxidative stress and greater bone anabolism in the aged, when compared to the young, murine skeleton with parathyroid hormone administration. *Aging Cell* **9** 851–867. (doi:10.1111/j.1474-9726.2010.00616.x)
- Kassem M & Marie PJ 2011 Senescence-associated intrinsic mechanisms of osteoblast dysfunctions. *Aging Cell* **10** 191–197. (doi:10.1111/j.1474-9726.2011.00669.x)
- Kawano Y & Kypta R 2003 Secreted antagonists of the Wnt signalling pathway. *Journal of Cell Science* **116** 2627–2634. (doi:10.1242/jcs.00623)
- Keller H & Kneissel M 2005 SOST is a target gene for PTH in bone. *Bone* **37** 148–158. (doi:10.1016/j.bone.2005.03.018)
- Khosla S, Bellido TM, Drezner MK, Gordon CM, Harris TB, Kiel DP, Kream BE, LeBoff MS, Lian JB, Peterson CA *et al.* 2011 Forum on aging and skeletal health: summary of the proceedings of an ASBMR workshop. *Journal of Bone and Mineral Research* **26** 2565–2578. (doi:10.1002/jbmr.488)
- Kim SW, Pajevic PD, Selig M, Barry KJ, Yang JY, Shin CS, Baek WY, Kim JE & Kronenberg HM 2012 Intermittent parathyroid hormone administration converts quiescent lining cells to active osteoblasts. *Journal of Bone and Mineral Research* **27** 2075–2084. (doi:10.1002/jbmr.1665)
- Knopp E, Troiano N, Bouxsein M, Sun BH, Lostritto K, Gundberg C, Dziura J & Insogna K 2005 The effect of aging on the skeletal response to intermittent treatment with parathyroid hormone. *Endocrinology* **146** 1983–1990. (doi:10.1210/en.2004-0770)
- Kramer I, Loots GG, Studer A, Keller H & Kneissel M 2010 Parathyroid hormone (PTH)-induced bone gain is blunted in SOST overexpressing and deficient mice. *Journal of Bone and Mineral Research* **25** 178–189. (doi:10.1359/jbmr.090730)
- Kramer I, Baertschi S, Halleux C, Keller H & Kneissel M 2012 *Mef2c* deletion in osteocytes results in increased bone mass. *Journal of Bone and Mineral Research* **27** 360–373. (doi:10.1002/jbmr.1492)
- Kulkarni NH, Halladay DL, Miles RR, Gilbert LM, Frolik CA, Galvin RJ, Martin TJ, Gillespie MT & Onyia JE 2005 Effects of parathyroid hormone on Wnt signaling pathway in bone. *Journal of Cellular Biochemistry* **95** 1178–1190. (doi:10.1002/jcb.20506)
- Leupin O, Kramer I, Collette NM, Loots GG, Natt F, Kneissel M & Keller H 2007 Control of the SOST bone enhancer by PTH using MEF2 transcription factors. *Journal of Bone and Mineral Research* **22** 1957–1967. (doi:10.1359/jbmr.070804)
- Li X, Zhang Y, Kang H, Liu W, Liu P, Zhang J, Harris SE & Wu D 2005 Sclerostin binds to LRP5/6 and antagonizes canonical Wnt signaling. *Journal of Biological Chemistry* **280** 19883–19887. (doi:10.1074/jbc.M413274200)
- Manolagas SC & Parfitt AM 2010 What old means to bone. *Trends in Endocrinology and Metabolism* **21** 369–374. (doi:10.1016/j.tem.2010.01.010)
- Marie PJ 2014 Bone cell senescence: mechanisms and perspectives. *Journal of Bone and Mineral Research* **29** 1311–1321. (doi:10.1002/jbmr.2190)
- Ono M, Inkson CA, Kilts TM & Young MF 2011 WISP-1/CCN4 regulates osteogenesis by enhancing BMP-2 activity. *Journal of Bone and Mineral Research* **26** 193–208. (doi:10.1002/jbmr.205)
- Parfitt AM, Drezner MK, Glorieux FH, Kanis JA, Malluche H, Meunier PJ, Ott SM & Recker RR 1987 Bone histomorphometry: standardization of nomenclature, symbols, and units. Report of the ASBMR Histomorphometry Nomenclature Committee. *Journal of Bone and Mineral Research* **2** 595–610. (doi:10.1002/jbmr.5650020617)
- Qin L, Qiu P, Wang L, Li X, Swarthout JT, Soteropoulos P, Toliás P & Partridge NC 2003 Gene expression profiles and transcription factors involved in parathyroid hormone signaling in osteoblasts revealed by microarray and bioinformatics. *Journal of Biological Chemistry* **278** 19723–19731. (doi:10.1074/jbc.M212226200)
- Rauner M, Sipos W & Pietschmann P 2008 Age-dependent Wnt gene expression in bone and during the course of osteoblast differentiation. *Age* **30** 273–282. (doi:10.1007/s11357-008-9069-9)
- Saidak Z, Hayé E, Marty C, Barbara A & Marie PJ 2012 Strontium ranelate rebalances bone marrow adipogenesis and osteoblastogenesis in senescent osteopenic mice through NFATc/Maf and Wnt signaling. *Aging Cell* **11** 467–474. (doi:10.1111/j.1474-9726.2012.00804.x)
- Tobimatsu T, Kaji H, Sowa H, Naito J, Canaff L, Hendy GN, Sugimoto T & Chihara K 2006 Parathyroid hormone increases β -catenin levels through Smad3 in mouse osteoblastic cells. *Endocrinology* **147** 2583–2590. (doi:10.1210/en.2005-1627)
- Wan M, Yang C, Li J, Wu X, Yuan H, Ma H, He X, Nie S, Chang C & Cao X 2008 Parathyroid hormone signaling through low-density lipoprotein-related protein 6. *Genes and Development* **22** 2968–2979. (doi:10.1101/gad.1702708)
- Xu L, Corcoran RB, Welsh JW, Pennica D & Levine AJ 2000 WISP-1 is a Wnt-1- and β -catenin-responsive oncogene. *Genes and Development* **14** 585–595. (doi:10.1101/gad.14.5.585)
- Yao W, Cheng Z, Shahnazari M, Dai W, Johnson ML & Lane NE 2010 Overexpression of secreted Frizzled-related protein 1 inhibits bone formation and attenuates PTH bone anabolic effects. *Journal of Bone and Mineral Research* **25** 190–199. (doi:10.1359/jbmr.090719)
- Yao GQ, Wu JJ, Troiano N & Insogna K 2011 Targeted overexpression of Dkk1 in osteoblasts reduces bone mass but does not impair the anabolic response to intermittent PTH treatment in mice. *Journal of Bone and Mineral Metabolism* **29** 141–148. (doi:10.1007/s00774-010-0202-3)

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