

The role of bile acids in metabolic regulation

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Abstract

Bile acids (BA), long believed to only have lipid-digestive functions, have emerged as novel metabolic modulators. They have important endocrine effects through multiple cytoplasmic as well as nuclear receptors in various organs and tissues. BA affect multiple functions to control energy homeostasis, as well as glucose and lipid metabolism, predominantly by activating the nuclear farnesoid X receptor and the cytoplasmic G protein-coupled BA receptor TGR5 in a variety of tissues. However, BA also are aimed at many other cellular targets in a wide array of organs and cell compartments. Their role in the pathogenesis of diabetes, obesity and other 'diseases of civilization' becomes even more clear. They also interact with the gut microbiome, with important clinical implications, further extending the complexity of their biological functions. Therefore, it is not surprising that BA metabolism is substantially modulated by bariatric surgery, a phenomenon contributing favorably to the therapeutic effects of these surgical procedures. Based on these data, several therapeutic approaches to ameliorate obesity and diabetes have been proposed to affect the cellular targets of BA.

Key Words

- ▶ bile acids
- ▶ diabetes
- ▶ obesity
- ▶ energy homeostasis
- ▶ bariatric surgery

Journal of Endocrinology
(2016) **228**, R85–R96

Introduction

Bile acids (BA), which for decades were considered to only be involved in lipid digestion in the intestinal lumen and cholesterol solubilization in the bile, now seem to have pleiotropic effects: contributing to the homeostasis of lipids, glucose and other metabolic substrates (Li & Chiang 2014, Qi *et al.* 2015), affecting immune system functions (Sipka & Bruckner 2014) as well as gut microbiome composition (Ridlon *et al.* 2014). By binding to multiple cytoplasmic as well as nuclear receptors in various organs and tissues, they act as real hormones. Based on these facts, a paradigm for BA endocrine functions has recently been postulated (Houten *et al.* 2006).

Bile acid metabolism

BA are amphiphilic molecules derived from cholesterol in the hepatocytes. The principal metabolic changes are

comprised of shortening of the cholesterol side chain plus hydroxylation of the core of the molecule at specific carbon atoms, forming the primary BA, cholic acid and chenodeoxycholic acid respectively (Hofmann 1984). These primary BA, after conjugation with glycine or taurine in the liver, are actively secreted into the bile via the bile salt export pump (BSEP, ABCB11, OMIM *603201). The BA pool in the human body is maintained by the efficient enterohepatic circulation (Hofmann 1984), preserving as much as 95% of conjugated BA. The active reabsorption of conjugated BA occurs in the distal ileum through the ASBT (also known as ISBT/IBAT/NTCP2, OMIM *601295) (Modica *et al.* 2010), and is profoundly deteriorated by inflammatory conditions affecting the ileal mucosa, such as in Crohn's disease (Lenicek *et al.* 2011, Vitek 2015). Unconjugated BA, formed after glycine/taurine hydrolysis by intestinal bacteria, can also

be absorbed by passive diffusion from both the small and large intestine; however, their transport is much less effective via this route. The so-called 'BA gut–liver axis' is regulated by the farnesoid X receptor (FXR), the intracellular BA sensor. In the ileocytes, FXR controls for BA uptake, their intracellular trafficking as well as BA basolateral efflux (mediated by organic solute transporters α/β (OST α/β)) (Zwicker & Agellon 2013). Simultaneously, ileal enterocytes, upon stimulation of FXR with BA, also secrete FGF19, which downregulates BA synthesis in the hepatocytes (Holt *et al.* 2003). However, FGF19 secreted from the small intestine in response to feeding also has insulin-like functions, whereas FGF21, a counterpart to FGF19, secreted from the liver in response to prolonged fasting, has glucagon-like effects (Potthoff *et al.* 2012). In fact, while insulin/glucagon serve as immediately acting fed-state and fasted-state hormones, FGF19 and FGF21 can be considered late-acting hormones (Potthoff *et al.* 2012). Interestingly, when administered in pharmacological doses, both FGF19 and FGF21 have insulin-sensitizing and hypolipidemic effects in rodent models of obesity and type 2 diabetes (T2DM) (Tomlinson *et al.* 2002, Kharitonov *et al.* 2005).

Apart from the FGF19 signaling pathway, hepatic BA synthesis is also controlled by another FXR-dependent mechanism, which is the small heterodimer partner (SHP, another orphan nuclear receptor) mediated downregulation of the *CYP7A1* gene, coding for cholesterol 7 α -hydroxylase, the rate-limiting enzyme in BA synthesis from cholesterol (Chiang 2009). In addition to this important function, SHP serves as a versatile corepressor of gene expression by inhibiting numerous transcriptional factors in diverse metabolic, proliferative and inflammatory pathways (Seok *et al.* 2013).

Cellular targets of BA

The emerging role of BA in various metabolic processes is mediated through several membrane and nuclear receptors (Zhou & Hylemon 2014). These involve specific nuclear receptors. Apart from FXR, there are also vitamin D receptor (VDR), constitutive androstane receptor (CAR), pregnane X receptor (PXR) as well as the cytoplasmic receptors TGR5, muscarinic receptors and sphingosine 1-phosphate receptor 2 (S1PR2) (Table 1).

FXR, in addition to its essential role in cholesterol/BA metabolism, also contributes to triacylglycerol (Fuchs *et al.* 2013) and glucose metabolism. In fact, BA, via activation of FXR present in pancreatic β cells, are capable of stimulating insulin production (Schittenhelm *et al.*

2015). Furthermore, FXR is also involved in the control of glucose homeostasis via its direct interaction with carbohydrate-responsive element-binding protein (ChREBP) acting as an important transcription factor of glycolytic genes (Benhamed *et al.* 2014). Surprisingly, FXR is also expressed in cardiovascular organs such as the heart and arterial system including coronary arteries, aorta as well as atherosclerotic arteries (Bishop-Bailey *et al.* 2004). The same is also true for the expression of VDR (Mathew *et al.* 2008) and PXR (Wang *et al.* 2013), pointing to the complexity of BA action on various organs and systems.

Although the role of PXR (another intracellular BA sensor in the pathogenesis of obesity and other metabolic disorders) awaits definite clarification, its effect on energy metabolism has been proven (Gao & Xie 2012). Interestingly, PXR is also involved in the modulation of the innate immunity system. This is demonstrated even in vascular cells, protecting them against the harmful effects of xenobiotics (Wang *et al.* 2014).

CAR, originally reported as a nuclear receptor regulating the response to xenobiotics, is another nuclear receptor activated by BA (Zhang *et al.* 2004, Huang *et al.* 2006, Sipka & Bruckner 2014). It is interesting to note that CAR has been marked as an anti-obesity nuclear receptor improving insulin sensitivity (Gao *et al.* 2009) as well as lipid metabolism (Maglich *et al.* 2009) and thyroid functions (Maglich *et al.* 2004).

TGR5 (*GPBAR1*), a member of the rhodopsin-like subfamily of G protein-coupled receptors, is expressed in the enteroendocrine small-intestinal cells as well as in the thyroid gland, brown adipose tissue (Zhou & Hylemon 2014), macrophages (Perino *et al.* 2014) and in many other organs (Duboc *et al.* 2014). An increasing body of evidence shows TGR5's important role in energy homeostasis, glucose metabolism (Thomas *et al.* 2009) and the modulation of immune functions (Perino *et al.* 2014). In addition, recently published data also demonstrated TGR5 expression in pancreatic β cells, with a direct effect on insulin secretion (Kumar *et al.* 2012) (in a similar manner as described above for FXR) and also in cardiomyocytes (Desai *et al.* 2010).

BA have also been reported to activate specific muscarinic receptors (Raufman *et al.* 2003). Although this phenomenon has primarily been discussed in relationship to possible gastrointestinal pathologies (Zhou & Hylemon 2014), stimulation of muscarinic receptors in endothelial cells has been shown to attenuate atherosclerosis in an experimental animal model (Zhou *et al.* 2014). Furthermore, muscarinic M3 receptors are also expressed in the adipose tissue (Yang *et al.* 2009) as well as

Table 1 Cellular targets of bile acids

Mechanism of action	Specific target ^a	Organ/tissue ^a	Function	References
Nuclear receptors	FXR	Liver Intestine Pancreatic β cells Cardiovascular system Liver Cardiovascular system Liver	Bile acid, lipid and glucose metabolism Bile acid metabolism Insulin secretion Cardiovascular system protection Lipid metabolism Vasoprotection	Mazuy <i>et al.</i> (2015) Mazuy <i>et al.</i> (2015) Schittenhelm <i>et al.</i> (2015) Bishop-Bailey <i>et al.</i> (2004) Chow <i>et al.</i> (2014) Mathew <i>et al.</i> (2008) Gao & Xie (2012)
	VDR	Vascular system Immune system Liver	Regulation of innate immunity system Regulation of innate immunity system Lipid and glucose metabolism	Wang <i>et al.</i> (2014) Wang <i>et al.</i> (2014) Maglich <i>et al.</i> (2009) and Gao <i>et al.</i> (2009)
	PXR	Liver	Induction of GLP-1 secretion Energy expenditure via activation of triiodothyronine	Parker <i>et al.</i> (2012) Watanabe <i>et al.</i> (2006)
	CAR	Brown adipose tissue/ skeletal muscles Pancreatic β cells Cardiac cells Immune system Intestine Endothelium Adipose tissue Pancreas Liver	Glucose metabolism Cardiac functions Anti-inflammatory action GI physiology Anti-atherogenic effects Obesity control Glucose metabolism Lipid metabolism Vasorelaxation Various effects	Kumar <i>et al.</i> (2012) Desai <i>et al.</i> (2010) Perino <i>et al.</i> (2014) Zhou & Hylemon (2014) Zhou <i>et al.</i> (2014) Yang <i>et al.</i> (2009) Hauge-Evans <i>et al.</i> (2014) Zhou & Hylemon (2014) Dopico <i>et al.</i> (2002) Hylemon <i>et al.</i> (2009)
	TGR5	Primarily liver	Multiple effects	Hylemon <i>et al.</i> (2009)
Cytoplasmic receptors	Muscarinic receptors	Primarily liver	Multiple effects	Hylemon <i>et al.</i> (2009)
	S1PR2	Primarily liver	Multiple effects	Hylemon <i>et al.</i> (2009)
Non-receptor-driven mechanisms	Large conductance Ca^{2+} -activated K^{+} channels	Vascular system Primarily liver	Glucose metabolism Lipid metabolism Vasorelaxation Various effects	Hylemon <i>et al.</i> (2009)
	Modulation of signalization cascades and enzymes regulating posttranslational modifications	Primarily liver	Multiple effects	Hylemon <i>et al.</i> (2009)

CAR, constitutive androstane receptor; EGFR, epidermal growth factor receptor; FXR, farnesoid X receptor; M3R, muscarinic M3 receptor; PXR, pregnane X receptor; S1PR2, sphingosine 1-phosphate receptor 2; TGR5, G protein-coupled bile acid receptor; VDR, vitamin D receptor.

^aOnly targets, organs and tissues believed to affect energy homeostasis are described.

the pancreas (Hauge-Evans *et al.* 2014), contributing significantly to glucose homeostasis.

Interestingly, BA also activate S1PR2 in liver cells, a mechanism believed to significantly regulate hepatic lipid metabolism (Zhou & Hylemon 2014). Sphingosine-1 phosphate, another natural ligand of S1PR2, also binds to apolipoprotein M, whose expression is under the influence of FXR, and it plays an important role in the pathogenesis of atherosclerosis and diabetes (Ren *et al.* 2015). Finally, deoxycholic acid, a secondary BA, has been surprisingly demonstrated to activate the epidermal growth factor receptor (EGFR) in hepatocytes (Qiao *et al.* 2001). EGFR is a factor known to be associated with the progression of vascular dysfunction in diabetes (Benter *et al.* 2015).

Apart from receptor-mediated actions, BA exert multiple functions through additional non-receptor-driven mechanisms involving the activation of large conductance Ca^{2+} -activated K^{+} channels that regulate arterial tone (Dopico *et al.* 2002), as well as many other less well-defined effects, including apoptosis, angiogenesis/neovascularization, NO metabolism and/or inflammatory processes (for a review, see Khurana *et al.* (2011)). This suggests the complexity of the totality of possible biological functions of BA.

This wide array of BA-mediated functions results in the modulation of multiple signaling pathways including JNK1/2, ERK1/2 or AKT1/2, with many possible biochemical, pathophysiological and clinical consequences (Hylemon *et al.* 2009). BA (particularly UDCA) have been reported to even modulate the miR-34a/sirtuin1/p53 pro-apoptotic pathway in non-alcoholic fatty liver disease (NAFLD; Castro *et al.* 2013). It also modulates sirtuin 1, a histone deacetylase, which is an important sensor in regulating energy homeostasis (Li 2013) as well as in diabetes pathophysiology (Kitada & Koya 2013). Indeed, BA, via multiple transcriptional cofactors such sirtuin 1 or SHP, have been suggested to behave as epigenomic cofactors affecting the posttranslational modification of histones (Kemper 2011, Smith *et al.* 2013), with deep impacts on the possible pathophysiological consequences. Indeed, BA-induced phosphorylation of SHP has been demonstrated to posttranslationally regulate hepatic metabolic genes (Seok *et al.* 2013). BA also inhibit lysine-specific histone demethylase 1 (LD1) (Kim *et al.* 2015), an enzyme playing an important role in adipogenesis (Musri *et al.* 2010) as well as in the development of diabetes (Brasacchio *et al.* 2009). Furthermore, BA have been shown to be involved in the posttranscriptional modification of

HMG-CoA reductase, a rate-limiting gene in cholesterol biosynthesis (Duckworth *et al.* 1991).

BA have also been shown to have potent immunosuppressive effects (Sipka & Bruckner 2014). Obesity is associated with a chronic, low-grade inflammation (termed metabolic inflammation), which is an important contributor to the initiation and progression of NAFLD, insulin resistance, T2DM and atherosclerosis (Gregor & Hotamisligil 2011). Thus, BA may also exert their potential anti-obesity actions via these anti-inflammatory mechanisms.

BA, gut microbiota and diabetes

Recently published data strongly suggests an important role of the gut microbiota in the development of obesity and NAFLD (Park *et al.* 2013). Gut microbiota cover more than 2000 species of commensal bacteria (Neish 2009), but only the recent progress in molecular techniques has revealed the vast diversity of gut microbiota, with Firmicutes and Bacteroidetes being the predominant bacterial phyla (Neish 2009).

Animal studies have proven that colonization of lean germ-free mice with the cecal microbiota of obese counterparts increases hepatic triacylglycerol accumulation, most likely through an increase in short-chain fatty acid (SCFA) fermentation in the intestinal lumen, leading to the stimulation of *de novo* synthesis of hepatic triacylglycerols (Backhed *et al.* 2004). Indeed, the gut microbiota of obese humans have a higher proportion of energy-harvesting Firmicutes bacteria, which is believed to increase the energy yield from the intestinal contents and accelerate fat accumulation in the human body (Turnbaugh *et al.* 2006). In fact, the ratio between the Firmicutes and Bacteroidetes phyla is important for SCFA production and is linked to obesity (Fernandes *et al.* 2014). These recent observations are also the rationale for several clinical trials recently initiated to treat obesity, NAFLD and/or diabetes with fecal bacterial transplantation ((Vrieze *et al.* 2012); and www.clinicaltrials.gov, accessed Oct 31, 2015). Further support for these data is reinforced by the results of gastric bypass studies, which have shown marked changes in the gut microbiota, correlating with weight loss (Zhang *et al.* 2009, Furet *et al.* 2010, Li *et al.* 2011).

Besides obesity, the pathogenesis of diabetes also seems to be closely linked with gut microbiota. Based on metagenome-wide association studies, an increase in sulfate-reducing bacteria and a decrease in butyrate-producing species have been detected in T2DM (for a

review, see Tilg & Moschen (2014)). It should be mentioned that sulfate-reducing bacteria utilize taurine for sulfur reduction. Taurine-conjugated BA are closely associated with a Western type of diet, contrasted to glycine-conjugated BA predominance (e.g. in native African populations), which are also devoid of 'diseases of civilization' (McGarr *et al.* 2005). Indeed, the Western diet has been reported to induce the taurine-conjugated BA pool, with important changes to the gut microbiome (Devkota *et al.* 2012). In addition, expansion of the Firmicutes phyla in the gut lumen is related to the intestinal BA pool (Islam *et al.* 2011). The relationship between BA metabolism and the intestinal microbiome is mutual. It is not only gut bacteria that are capable of metabolizing BA, but BA also importantly influence the gut's microbiota composition. This influence is mediated through direct antimicrobial effects on gut microbes (Begley *et al.* 2005), via production of antimicrobial peptides mediated by activated FXR in ileocytes (Inagaki *et al.* 2006), or by inhibiting intestinal absorption of bacterial endotoxins (Kocsar *et al.* 1969, Parlesak *et al.* 2007). However, these potential antimicrobial effects are still poorly understood (Hofmann & Eckmann 2006).

It is also interesting to note that gut microbiome diversity is an important factor, capable to differentiate between lean and obese human subjects (Le Chatelier *et al.* 2013). The gut microbiome gene richness efficiently responds to dietary interventions (Cotillard *et al.* 2013), suggesting promising therapeutic approaches for obese patients.

BA, besides their 'classical' lipid digestive and respective TGR5-mediated metabolic functions, may also exert many other effects within the intestinal lumen. These, in turn, affect the susceptibility to obesity, the metabolic syndrome and/or diabetes.

One of these additional mechanisms may involve BA-mediated modulation of innate intestinal immunity. It has been demonstrated that activation of FXR modulated TLR4 of the intestinal myeloid cells results in anti-inflammatory effects in murine models of colitis (Vavassori *et al.* 2009). Also, *vice versa*, activation of TLR4/9 on monocytes has potent modulating effects on FXR, indicating a close interplay between FXR and effectors of innate immunity (Renga *et al.* 2013). Although it has been proposed that these mechanisms are implicated in intestinal inflammatory diseases, it is highly likely that they can also affect energy homeostasis as well as the risks of obesity, NAFLD and diabetes. These are all conditions where TLRs are believed to play an important pathogenic role (Jia *et al.* 2014, Ferreira *et al.* 2015).

As mentioned above, both *TGR5* and *FXR* (*NR1H4*) are also functionally expressed in pancreatic β -cells, where they regulate insulin secretion (Renga *et al.* 2010, Seyer *et al.* 2013). In fact, *Fxr*-deficient mice develop insulin resistance (Zhang *et al.* 2006); on the other hand, activation of FXR reverses this phenomenon in various animal models (Zhang *et al.* 2006, Cipriani *et al.* 2010). Based on these observations, FXR activation has been proposed as a promising therapeutic target for diabetic patients (Zhang *et al.* 2006). Indeed, treatment of patients having NAFLD and T2DM with obeticholic acid (a potent FXR agonist) has been demonstrated to increase insulin sensitivity in a recent human trial (Mudaliar *et al.* 2013); however, this was not confirmed in another study (Neuschwander-Tetri *et al.* 2015).

Furthermore, GLUT4, the main insulin-responsive glucose transporter, playing a critical role in maintaining systemic glucose homeostasis and contributing to insulin resistance, can be induced in hepatocyte- and adipocyte-like cells by chenodeoxycholic acid, which is a natural FXR agonist (Shen *et al.* 2008). Nevertheless, BA might also exert protective effects via FXR-independent action by the suppression of hepatic fatty acid and triacylglycerol gene expression (Wu *et al.* 2014).

However, not all recent data are fully supportive of the concept of the beneficial effects of FXR activation on energy homeostasis. It has recently been reported that selective disruption of intestinal FXR mediates gut microbiota-associated NAFLD development via the ceramide axis, pointing out the complexity of the entirety of FXR-mediated actions (Jiang *et al.* 2015). Additionally, in a mouse model, it has been demonstrated that alteration of the gut microbiota can antagonize the intestinal FXR via increased production of taurine-conjugated muricholic acid (Li *et al.* 2013, Sayin *et al.* 2013). It remains to be confirmed whether human BA conjugates may also exert the same inhibitory action on intestinal FXR.

BA metabolism in constipation and possible link to metabolic diseases

The interrelationship between BA homeostasis and metabolic diseases is far more complex. For example, it is known that patients with chronic constipation have a higher risk for cardiovascular disease (Shakir *et al.* 2007, Salmoirago-Blotcher *et al.* 2011) and T2DM (Talley *et al.* 2003, Salmoirago-Blotcher *et al.* 2011). In fact, decreased fecal BA output, a phenomenon associated with chronic constipation (Abrahamsson *et al.* 2008, Hofmann *et al.* 2008), has been reported in patients with coronary

atherosclerosis (Charach *et al.* 1998, 2011). Constipation not only seems to be a consequence of autonomic diabetic neuropathy (Vinik *et al.* 2003), but, based on the evidence discussed above, is instead a contributing factor, via impaired intestinal and hepatic BA metabolism.

BA, thyroid functions and energy expenditure

Apart from in the small intestine, TGR5 is expressed in numerous tissues and organs including the thyroid gland, brown adipose tissue, skeletal cardiac muscle, liver and pancreas (Duboc *et al.* 2014). This fact led to investigations of the effects of BA on these organs. Surprisingly, it turned out that BA-activated TGR5 in brown fat in mice (as well as in human myocytes) stimulated intracellular cAMP formation and activated type 2 iodothyronine deiodinase (D2), which is responsible for the conversion of T₄ to T₃, and mediated thermogenic effects of BA (Watanabe *et al.* 2006). This observation was also confirmed in an experimental study by da-Silva *et al.* (2011), who observed exactly the same effects on D2 activity and energy expenditure with tauroursodeoxycholic acid. However, these effects may be due to the modulation of intracellular pathways unrelated to TGR5 activation (Malisova *et al.* 2013). In fact, thermogenic effects of BA were also verified in a human study by Ockenga *et al.* (2012), who reported a positive association in subjects of venous BA concentrations with energy expenditure; however, this was not observed in obese subjects (Brufau *et al.* 2010). The effect of BA on thyroid function seems to be more complex. A negative association between BA and TSH levels has been reported in certain patients (Patti *et al.* 2009, Ockenga *et al.* 2012) as well as in healthy subjects (Song *et al.* 2015). This is most likely due to the effect of BA on TGR5 expressed in the pituitary gland (Doignon *et al.* 2011, Ockenga *et al.* 2012). Importantly, BA sequestrants have been shown to efficiently ameliorate hyperthyroidism (Shakir *et al.* 1993, Hagag *et al.* 1998, Kaykhaei *et al.* 2008), even in refractory patients (Sebastian-Ochoa *et al.* 2008, Alswat 2015, Yang *et al.* 2015). This effect is believed to be mediated by impaired reabsorption of thyroid hormones (de Luis *et al.* 2002), but is probably much more complex.

In this context, it is also interesting to note a feedback effect of TSH/thyroxine on BA production, primarily mediated by the modulation of CYP7A1 in the liver tissue (Ellis 2006, Song *et al.* 2015). However, not all data are conclusive, and the role of thyroid hormones in the BA biosynthetic pathway still awaits further elucidation.

BA, incretins and glucose homeostasis

Incretin hormones, specifically glucagon-like peptide 1 (GLP-1) and glucose-dependent insulinotropic peptide (GIP), are intestine-derived hormones that increase insulin secretion and decrease glucagon secretion upon stimulation by food ingestion (Holst & Gromada 2004), thus significantly contributing to the regulation of glucose metabolism (Holst *et al.* 2008). A majority of patients with T2DM have a combination of reduced GLP-1 secretion and partial resistance to its effects (Nauck *et al.* 2011). Both of these defects contribute to impairments of glucose metabolism in T2DM. Pharmacological approaches, which either increase endogenous GLP-1 or use the analogues of GLP-1 with a longer half-life, are now routinely used in the treatment of T2DM (Martin *et al.* 2011). Glucagon-like peptide 2 (GLP-2) is another intestine-derived hormone, which, in contrast to GLP-1 and GIP, does not have incretin effects. It is an important regulator of gastric motility, gastric acid secretion and intestinal hexose transport, with enhancing effects on the barrier function of the gut epithelium (Yusta *et al.* 2012).

BA have been shown to directly promote GLP-1 and GLP-2 secretion in small-intestinal enteroendocrine cells through TGR5 (Parker *et al.* 2012). Furthermore, their effect appears to be synergistic to that of glucose (Parker *et al.* 2012). Studies have shown that increased BA concentrations after malabsorptive bariatric surgery procedures (details provided in the next section) correlate with peak GLP-1 levels and fasting GIP levels (Patti *et al.* 2009, Pournaras *et al.* 2012, Kohli *et al.* 2013a). Similar associations have also been found in some other types of surgical manipulations. Importantly, numerous experimental studies have also found a positive correlation between increased BA levels after bariatric surgery and improvements in glucose homeostasis (Penney *et al.* 2015).

How BA contribute to the effect of bariatric surgery on energy homeostasis

Bariatric surgery not only substantially decreases body weight, but also markedly improves glucose metabolism, frequently leading to a complete remission of diabetes, as evidenced in both experimental and clinical studies (Ashrafian *et al.* 2010). Nevertheless, different types of operations clearly differ in their rates of diabetes remission as well as in the timing of metabolic improvements (Dixon *et al.* 2012). In general, bariatric operations can be classified as restrictive procedures such as gastric banding, gastric plication and sleeve gastrectomy (LSG) – or malabsorptive

or combined procedures (e.g. gastric bypass, biliopancreatic diversion) (O'Brien 2010). In the former group, the decreased food intake and weight loss is achieved through the restriction of the stomach size without further modifications of digestive tract anatomy (Pories 2008). In the majority of malabsorptive procedures, the stomach size is also partially restricted, and a direct connection of the stomach and the lower part of the small intestine is created, thus bypassing a significant portion of the intestine. Studies have shown that restrictive procedures do not alter circulating BA concentrations, with a majority of the studies reporting either no change of BA after gastric banding (Kohli *et al.* 2013a) or inconsistent results after sleeve gastrectomy (Haluzikova *et al.* 2013, Myronovych

et al. 2014). On the contrary, malabsorptive operations such as gastric bypass, which are usually more effective toward improvements of glucose metabolism, increase circulating BA levels (Kohli *et al.* 2013a). Plasma BA levels have also been found to increase after ileal interposition surgery, where the ileum is repositioned distal to the duodenum (Kohli *et al.* 2010). This manipulation is also associated with improvement of the components of the metabolic syndrome in rats with diet-induced obesity. Furthermore, the insertion of a duodenal-jejunal bypass liner (an endoscopically implanted device that eliminates the duodenum and proximal jejunum contact with digested food) has also been found to be associated with increased BA concentration in an experimental study

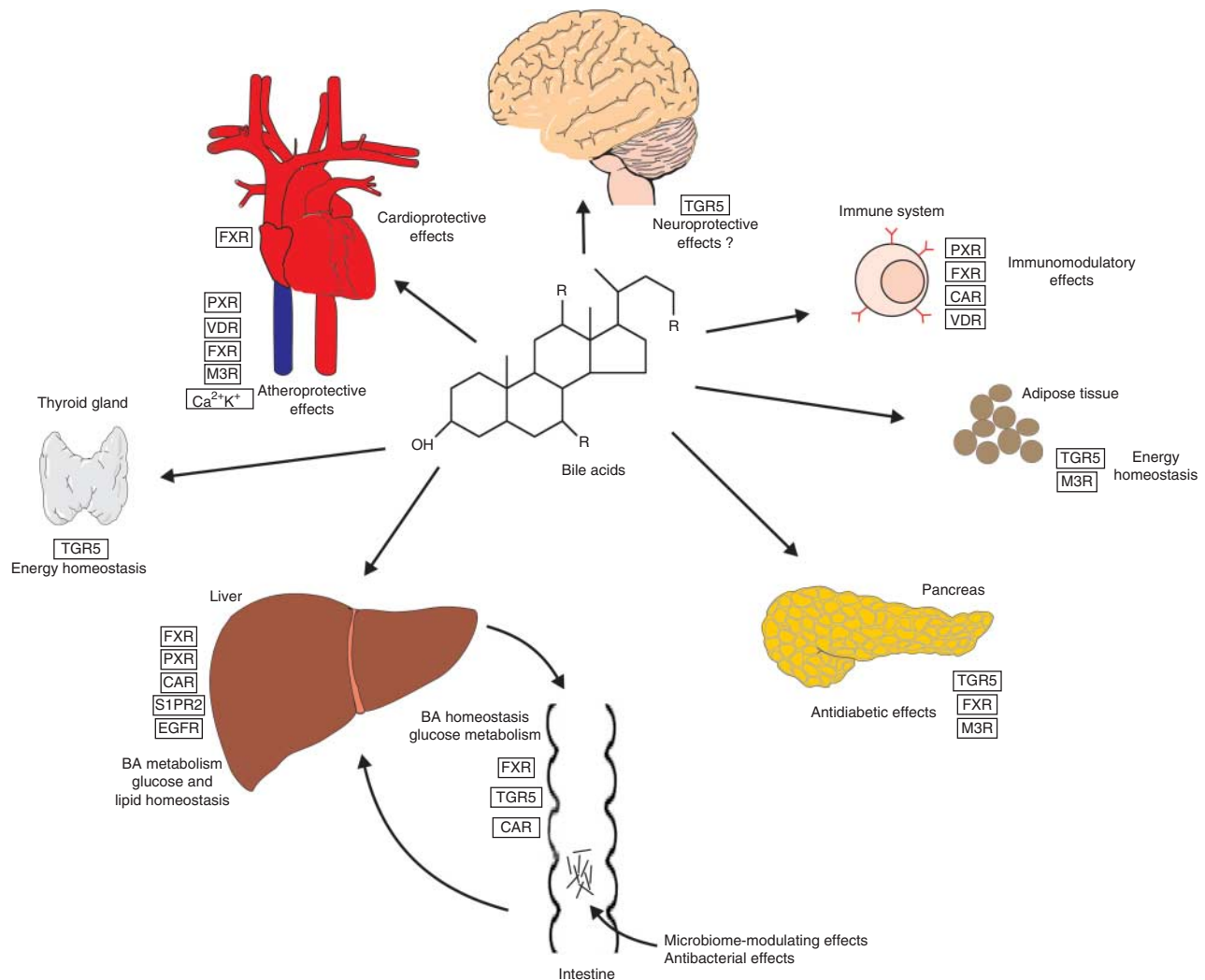


Figure 1

Receptor-mediated effects of bile acids on various tissues and organs involved in energy homeostasis. CAR, constitutive androstane receptor; EGFR, epidermal growth factor receptor; FXR, farnesoid X receptor;

M3R, muscarinic M3 receptor; PXR, pregnane X receptor; S1PR2, sphingosine 1-phosphate receptor 2; TGR5, G protein-coupled bile acid receptor; VDR, vitamin D receptor.

(Habegger *et al.* 2014). We recently observed a similar increase in patients with type 2 diabetes 6 months after the implantation of a duodenal–jejunal bypass liner (Kavalkova P, Mraz M, Trachta P, Haluzikova D, Lacinová Z, Benes M, Vlasakova Z, Petr T, Vitek L, Pelikanova T & Haluzik M, unpublished observations). Two experimental studies have directly demonstrated the importance of BA in post-bariatric surgery metabolic improvements by experimental diversion of bile to the distal gut, using either a catheter placed into the common bile duct of male obese rats to divert BA to the more distal jejunum (Kohli *et al.* 2013b) or a surgical manipulation to divert BA into the ileum (Pournaras *et al.* 2012) respectively. Both manipulations were associated with increased serum BA, postprandial GLP-1 secretion and improved glucose metabolism. Finally, *Fxr*-null mice have been shown to exhibit significantly blunted weight loss and improvements in glucose metabolism after bariatric surgery, suggesting an important role of FXR-mediated BA signaling after bariatric surgery (Ryan *et al.* 2014). Taken together, BA concentrations are significantly increased after malabsorptive bariatric surgery procedures. Multiple studies have shown that this increase significantly contributes to improvements in glucose homeostasis through modulations of GLP-1 secretion from the gut, changes of gut microbiota and endocrine effects of circulating BA in various organs and tissues (Fig. 1).

Conclusion

Our knowledge on the effects of BA on energy homeostasis and metabolism has dramatically expanded during the last decade. It is likely that further surprises are on the horizon, and BA will appear to have even more profound metabolic impacts. One example might be found in the observation that administration of BA sequestrants (a completely different approach from that described above) increases insulin sensitivity (Staels & Kuipers 2007, Suzuki *et al.* 2007a). This most likely takes place by delaying fatty acid absorption (Suzuki *et al.* 2007b) and increased production of secondary BA derived from sequestrant-trapped primary BA (Harach *et al.* 2012), thus leading to the stimulation of GLP-1 release. The same mechanism also seems to be in play for the ASBT inhibitor elobixibat, which may provide positive metabolic side effects (in addition to positive effects on the symptoms and signs of constipation mentioned above), reducing the risk for CVD and T2DM. Indeed, decreased LDL-cholesterol and increased GLP-1 levels have been reported in patients with dyslipidemia treated with elobixibat (Rudling *et al.* 2015). Interestingly,

the hypoglycemic effect of metformin has been suggested to be at least partly accounted for by decreased intestinal BA absorption (Carter *et al.* 2003). Thus, it is not surprising that colesevelam, a potent BA sequestrant, has been approved by the FDA for the treatment of T2DM. An increasing body of evidence demonstrates the important therapeutic potential of BA metabolism modulation, either through the direct regulation of a wide array of their specific receptors or as a consequence of bariatric surgical procedures. BA interaction with the intestinal microbiome is also important, although in this respect, our knowledge is still far from complete.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

Funding

This work was supported by grants PRV0UK 4102280002 from the Czech Ministry of Education, and RVO VFN64165 and NT13151-4 from the Czech Ministry of Health.

References

- Abrahamsson H, Ostlund-Lindqvist AM, Nilsson R, Simren M & Gillberg PG 2008 Altered bile acid metabolism in patients with constipation-predominant irritable bowel syndrome and functional constipation. *Scandinavian Journal of Gastroenterology* **43** 1483–1488. (doi:10.1080/00365520802321212)
- Alswat KA 2015 Role of cholestyramine in refractory hyperthyroidism: a case report and literature review. *American Journal of Case Reports* **16** 486–490. (doi:10.12659/AJCR.893821)
- Ashrafiyan H, Bueter M, Ahmed K, Suliman A, Bloom SR, Darzi A & Athanasiou T 2010 Metabolic surgery: an evolution through bariatric animal models. *Obesity Reviews* **11** 907–920. (doi:10.1111/j.1467-789X.2009.00701.x)
- Backhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF & Gordon JI 2004 The gut microbiota as an environmental factor that regulates fat storage. *PNAS* **101** 15718–15723. (doi:10.1073/pnas.0407076101)
- Begley M, Gahan CG & Hill C 2005 The interaction between bacteria and bile. *FEMS Microbiology Reviews* **29** 625–651. (doi:10.1016/j.femsre.2004.09.003)
- Benhamed F, Filhoulaud G, Caron S, Lefebvre P, Staels B & Postic C 2014 O-GlcNAcylation links ChREBP and FXR to glucose-sensing. *Frontiers in Endocrinology* **5** 230. (doi:10.3389/fendo.2014.00230)
- Benter IF, Sarkhou F, Al-Khalidi AT, Chandrasekhar B, Attur S, Dhaunsi GS, Yousif MH & Akhtar S 2015 The dual targeting of EGFR and ErbB2 with the inhibitor Lapatinib corrects high glucose-induced apoptosis and vascular dysfunction by opposing multiple diabetes-induced signaling changes. *Journal of Drug Targeting* **23** 506–518. (doi:10.3109/1061186X.2015.1057150)
- Bishop-Bailey D, Walsh DT & Warner TD 2004 Expression and activation of the farnesoid X receptor in the vasculature. *PNAS* **101** 3668–3673. (doi:10.1073/pnas.0400046101)
- Brasacchio D, Okabe J, Tikellis C, Balcerzyk A, George P, Baker EK, Calkin AC, Brownlee M, Cooper ME & El-Osta A 2009 Hyperglycemia

- induces a dynamic cooperativity of histone methylase and demethylase enzymes associated with gene-activating epigenetic marks that coexist on the lysine tail. *Diabetes* **58** 1229–1236. (doi:10.2337/db08-1666)
- Brufau G, Bahr MJ, Staels B, Claudel T, Ockenga J, Boker KH, Murphy EJ, Prado K, Stellaard F, Manns MP *et al.* 2010 Plasma bile acids are not associated with energy metabolism in humans. *Nutrition and Metabolism* **7** 73. (doi:10.1186/1743-7075-7-73)
- Carter D, Howlett HC, Wiernsperger NF & Bailey CJ 2003 Differential effects of metformin on bile salt absorption from the jejunum and ileum. *Diabetes, Obesity and Metabolism* **5** 120–125. (doi:10.1046/j.1463-1326.2003.00252.x)
- Castro RE, Ferreira DM, Afonso MB, Borralho PM, Machado MV, Cortez-Pinto H & Rodrigues CM 2013 miR-34a/SIRT1/p53 is suppressed by ursodeoxycholic acid in the rat liver and activated by disease severity in human non-alcoholic fatty liver disease. *Journal of Hepatology* **58** 119–125. (doi:10.1016/j.jhep.2012.08.008)
- Charach G, Rabinovich PD, Konikoff FM, Grosskopf I, Weintraub MS & Gilat T 1998 Decreased fecal bile acid output in patients with coronary atherosclerosis. *Journal of Medicine* **29** 125–136.
- Charach G, Grosskopf I, Rabinovich A, Shochat M, Weintraub M & Rabinovich P 2011 The association of bile acid excretion and atherosclerotic coronary artery disease. *Therapeutic Advances in Gastroenterology* **4** 95–101. (doi:10.1177/1756283X10388682)
- Chiang JY 2009 Bile acids: regulation of synthesis. *Journal of Lipid Research* **50** 1955–1966. (doi:10.1194/jlr.R900010-JLR200)
- Chow EC, Magomedova L, Quach HP, Patel R, Durk MR, Fan J, Maeng HJ, Irondi K, Anakk S, Moore DD *et al.* 2014 Vitamin D receptor activation down-regulates the small heterodimer partner and increases CYP7A1 to lower cholesterol. *Gastroenterology* **146** 1048–1059. (doi:10.1053/j.gastro.2013.12.027)
- Cipriani S, Mencarelli A, Palladino G & Fiorucci S 2010 FXR activation reverses insulin resistance and lipid abnormalities and protects against liver steatosis in Zucker (fa/fa) obese rats. *Journal of Lipid Research* **51** 771–784. (doi:10.1194/jlr.M001602)
- Cotillard A, Kennedy SP, Kong LC, Prifti E, Pons N, Le Chatelier E, Almeida M, Quinquis B, Levenez F, Galleron N *et al.* 2013 Dietary intervention impact on gut microbial gene richness. *Nature* **500** 585–588. (doi:10.1038/nature12480)
- Desai MS, Shabier Z, Taylor M, Lam F, Thevananther S, Kusters A & Karpen SJ 2010 Hypertrophic cardiomyopathy and dysregulation of cardiac energetics in a mouse model of biliary fibrosis. *Hepatology* **51** 2097–2107. (doi:10.1002/hep.23585)
- Devkota S, Wang Y, Musch MW, Leone V, Fehlner-Peach H, Nadimpalli A, Antonopoulos DA, Jabri B & Chang EB 2012 Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in H10-/- mice. *Nature* **487** 104–108. (doi:10.1038/nature11225)
- Dixon JB, le Roux CW, Rubino F & Zimmet P 2012 Bariatric surgery for type 2 diabetes. *Lancet* **379** 2300–2311. (doi:10.1016/S0140-6736(12)60401-2)
- Doignon I, Julien B, Serriere-Lanneau V, Garcin I, Alonso G, Nicou A, Monnet F, Gigou M, Humbert L, Rainteau D *et al.* 2011 Immediate neuroendocrine signaling after partial hepatectomy through acute portal hyperpressure and cholestasis. *Journal of Hepatology* **54** 481–488. (doi:10.1016/j.jhep.2010.07.012)
- Dopico AM, Walsh JV Jr & Singer JJ 2002 Natural bile acids and synthetic analogues modulate large conductance Ca²⁺-activated K⁺ (BKCa) channel activity in smooth muscle cells. *Journal of General Physiology* **119** 251–273. (doi:10.1085/jgp.20028537)
- Duboc H, Tache Y & Hofmann AF 2014 The bile acid TGR5 membrane receptor: from basic research to clinical application. *Digestive and Liver Disease* **46** 302–312. (doi:10.1016/j.dld.2013.10.021)
- Duckworth PF, Vlahcevic ZR, Studer EJ, Gurley EC, Heuman DM, Beg ZH & Hylemon PB 1991 Effect of hydrophobic bile acids on 3-hydroxy-3-methylglutaryl-coenzyme A reductase activity and mRNA levels in the rat. *Journal of Biochemical Chemistry* **266** 9413–9418.
- Ellis EC 2006 Suppression of bile acid synthesis by thyroid hormone in primary human hepatocytes. *World Journal of Gastroenterology* **12** 4640–4645. (doi:10.3748/wjg.v12.i29.4640)
- Fernandes J, Su W, Rahat-Rozenbloom S, Wolever TM & Comelli EM 2014 Adiposity, gut microbiota and faecal short chain fatty acids are linked in adult humans. *Nutrition & Diabetes* **4** e121. (doi:10.1038/nutd.2014.23)
- Ferreira DF, Fiamoncini J, Prist IH, Ariga SK, de Souza HP & de Lima TM 2015 Novel role of TLR4 in NAFLD development: modulation of metabolic enzymes expression. *Biochimica Biophysica Acta* **1851** 1353–1359. (doi:10.1016/j.bbali.2015.07.002)
- Fuchs C, Claudel T & Trauner M 2013 Bile acid-mediated control of liver triglycerides. *Seminars in Liver Disease* **33** 330–342. (doi:10.1055/s-0033-1358520)
- Furet JP, Kong LC, Tap J, Poitou C, Basdevant A, Bouillot JL, Mariat D, Corthier G, Dore J, Henegar C *et al.* 2010 Differential adaptation of human gut microbiota to bariatric surgery-induced weight loss: links with metabolic and low-grade inflammation markers. *Diabetes* **59** 3049–3057. (doi:10.2337/db10-0253)
- Gao J & Xie W 2012 Targeting xenobiotic receptors PXR and CAR for metabolic diseases. *Trends in Pharmacological Sciences* **33** 552–558. (doi:10.1016/j.tips.2012.07.003)
- Gao J, He J, Zhai Y, Wada T & Xie W 2009 The constitutive androstane receptor is an anti-obesity nuclear receptor that improves insulin sensitivity. *Journal of Biochemical Chemistry* **284** 25984–25992. (doi:10.1074/jbc.M109.016808)
- Gregor MF & Hotamisligil GS 2011 Inflammatory mechanisms in obesity. *Annual Review of Immunology* **29** 415–445. (doi:10.1146/annurev-immunol-031210-101322)
- Habegger KM, Al-Massadi O, Heppner KM, Myronovych A, Holland J, Berger J, Yi CX, Gao Y, Lehti M, Ottaway N *et al.* 2014 Duodenal nutrient exclusion improves metabolic syndrome and stimulates villus hyperplasia. *Gut* **63** 1238–1246. (doi:10.1136/gutjnl-2013-304583)
- Hagag P, Nissenbaum H & Weiss M 1998 Role of colestipol in the treatment of hyperthyroidism. *Journal of Endocrinological Investigation* **21** 725–731. (doi:10.1007/BF03348036)
- Haluzikova D, Lacinova Z, Kavalkova P, Drapalova J, Krizova J, Bartlova M, Mraz M, Petr T, Vitek L, Kasalicky M *et al.* 2013 Laparoscopic sleeve gastrectomy differentially affects serum concentrations of FGF-19 and FGF-21 in morbidly obese subjects. *Obesity* **21** 1335–1342. (doi:10.1002/oby.20208)
- Harach T, Pols TW, Nomura M, Maida A, Watanabe M, Auwerx J & Schoonjans K 2012 TGR5 potentiates GLP-1 secretion in response to anionic exchange resins. *Scientific Reports* **2** 430. (doi:10.1038/srep00430)
- Hauge-Evans AC, Reers C, Kerby A, Franklin Z, Amisten S, King AJ, Hassan Z, Vilches-Flores A, Tippu Z, Persaud SJ *et al.* 2014 Effect of hyperglycaemia on muscarinic M3 receptor expression and secretory sensitivity to cholinergic receptor activation in islets. *Diabetes, Obesity and Metabolism* **16** 947–956. (doi:10.1111/dom.12301)
- Hofmann AF 1984 Chemistry and enterohepatic circulation of bile acids. *Hepatology* **4** 4S–14S. (doi:10.1002/hep.1840040803)
- Hofmann AF & Eckmann L 2006 How bile acids confer gut mucosal protection against bacteria. *PNAS* **103** 4333–4334. (doi:10.1073/pnas.0600780103)
- Hofmann AF, Loening-Baucke V, Lavine JE, Hagey LR, Steinbach JH, Packard CA, Griffin TL & Chatfield DA 2008 Altered bile acid metabolism in childhood functional constipation: inactivation of secretory bile acids by sulfation in a subset of patients. *Journal of Pediatric Gastroenterology and Nutrition* **47** 598–606. (doi:10.1097/MPG.0b013e31816920a6)
- Holst JJ & Gromada J 2004 Role of incretin hormones in the regulation of insulin secretion in diabetic and nondiabetic humans. *American Journal of Physiology-Endocrinology and Metabolism* **287** E199–E206. (doi:10.1152/ajpendo.00545.2003)

- Holst JJ, Deacon CF, Vilsboll T, Krarup T & Madsbad S 2008 Glucagon-like peptide-1, glucose homeostasis and diabetes. *Trends in Molecular Medicine* **14** 161–168. (doi:10.1016/j.molmed.2008.01.003)
- Holt JA, Luo G, Billin AN, Bisi J, McNeill YY, Kozarsky KF, Donahee M, Wang DY, Mansfield TA, Kliewer SA *et al.* 2003 Definition of a novel growth factor-dependent signal cascade for the suppression of bile acid biosynthesis. *Genes and Development* **17** 1581–1591. (doi:10.1101/gad.1083503)
- Houten SM, Watanabe M & Auwerx J 2006 Endocrine functions of bile acids. *EMBO Journal* **25** 1419–1425. (doi:10.1038/sj.emboj.7601049)
- Huang W, Ma K, Zhang J, Qatanani M, Cuvillier J, Liu J, Dong B, Huang X & Moore DD 2006 Nuclear receptor-dependent bile acid signaling is required for normal liver regeneration. *Science* **312** 233–236. (doi:10.1126/science.1121435)
- Hylemon PB, Zhou H, Pandak WM, Ren S, Gil G & Dent P 2009 Bile acids as regulatory molecules. *Journal of Lipid Research* **50** 1509–1520. (doi:10.1194/jlr.R900007-JLR200)
- Inagaki T, Moschetta A, Lee YK, Peng L, Zhao G, Downes M, Yu RT, Shelton JM, Richardson JA, Repa JJ *et al.* 2006 Regulation of antibacterial defense in the small intestine by the nuclear bile acid receptor. *PNAS* **103** 3920–3925. (doi:10.1073/pnas.0509592103)
- Islam KB, Fukiya S, Hagio M, Fujii N, Ishizuka S, Ooka T, Ogura Y, Hayashi T & Yokota A 2011 Bile acid is a host factor that regulates the composition of the cecal microbiota in rats. *Gastroenterology* **141** 1773–1781. (doi:10.1053/j.gastro.2011.07.046)
- Jia L, Vianna CR, Fukuda M, Berglund ED, Liu C, Tao C, Sun K, Liu T, Harper MJ, Lee CE *et al.* 2014 Hepatocyte Toll-like receptor 4 regulates obesity-induced inflammation and insulin resistance. *Nature Communications* **5** 3878. (doi:10.1038/ncomms4878)
- Jiang C, Xie C, Li F, Zhang L, Nichols RG, Krausz KW, Cai J, Qi Y, Fang ZZ, Takahashi S *et al.* 2015 Intestinal farnesoid X receptor signaling promotes nonalcoholic fatty liver disease. *Journal of Clinical Investigation* **125** 386–402. (doi:10.1172/JCI76738)
- Kaykhaei MA, Shams M, Sadegholvad A, Dabbaghmanesh MH & Omrani GR 2008 Low doses of cholestyramine in the treatment of hyperthyroidism. *Endocrine* **34** 52–55. (doi:10.1007/s12020-008-9107-5)
- Kemper JK 2011 Regulation of FXR transcriptional activity in health and disease: emerging roles of FXR cofactors and post-translational modifications. *Biochimica Biophysica Acta* **1812** 842–850. (doi:10.1016/j.bbadis.2010.11.011)
- Kharitonov A, Shiyanova TL, Koester A, Ford AM, Micanovic R, Galbreath EJ, Sandusky GE, Hammond LJ, Moyers JS, Owens RA *et al.* 2005 FGF-21 as a novel metabolic regulator. *Journal of Clinical Investigation* **115** 1627–1635. (doi:10.1172/JCI23606)
- Khurana S, Raufman JP & Pallone TL 2011 Bile acids regulate cardiovascular function. *Clinical and Translational Science* **4** 210–218. (doi:10.1111/j.1752-8062.2011.00272.x)
- Kim YC, Fang S, Byun S, Seok S, Kemper B & Kemper JK 2015 Farnesoid X receptor-induced lysine-specific histone demethylase reduces hepatic bile acid levels and protects the liver against bile acid toxicity. *Hepatology* **62** 220–231. (doi:10.1002/hep.27677)
- Kitada M & Koya D 2013 SIRT1 in type 2 diabetes: mechanisms and therapeutic potential. *Diabetes & Metabolism Journal* **37** 315–325. (doi:10.4093/dmj.2013.37.5.315)
- Kocsar LT, Bertok L & Varteresz V 1969 Effect of bile acids on the intestinal absorption of endotoxin in rats. *Journal of Bacteriology* **100** 220–223.
- Kohli R, Kirby M, Setchell KD, Jha P, Klustaitis K, Woollett LA, Pfluger PT, Balistreri WF, Tso P, Jandacek RJ *et al.* 2010 Intestinal adaptation after ileal interposition surgery increases bile acid recycling and protects against obesity-related comorbidities. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **299** G652–G660. (doi:10.1152/ajpgi.00221.2010)
- Kohli R, Bradley D, Setchell KD, Eagon JC, Abumrad N & Klein S 2013a Weight loss induced by Roux-en-Y gastric bypass but not laparoscopic adjustable gastric banding increases circulating bile acids. *Journal of Clinical Endocrinology and Metabolism* **98** E708–E712. (doi:10.1210/jc.2012-3736)
- Kohli R, Setchell KD, Kirby M, Myronovych A, Ryan KK, Ibrahim SH, Berger J, Smith K, Toure M, Woods SC *et al.* 2013b A surgical model in male obese rats uncovers protective effects of bile acids post-bariatric surgery. *Endocrinology* **154** 2341–2351. (doi:10.1210/en.2012-2069)
- Kumar DP, Rajagopal S, Mahavadi S, Mirshahi F, Grider JR, Murthy KS & Sanyal AJ 2012 Activation of transmembrane bile acid receptor TGR5 stimulates insulin secretion in pancreatic β cells. *Biochemical and Biophysical Research Communications* **427** 600–605. (doi:10.1016/j.bbrc.2012.09.104)
- Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, Almeida M, Arumugam M, Batto JM, Kennedy S *et al.* 2013 Richness of human gut microbiome correlates with metabolic markers. *Nature* **500** 541–546. (doi:10.1038/nature12506)
- Lenicek M, Duricova D, Komarek V, Gabrysova B, Lukas M, Smerhovsky Z & Vitek L 2011 Bile acid malabsorption in inflammatory bowel disease: assessment by serum markers. *Inflammatory Bowel Diseases* **17** 1322–1327. (doi:10.1002/ibd.21502)
- Li X 2013 SIRT1 and energy metabolism. *Acta Biochimica et Biophysica Sinica* **45** 51–60. (doi:10.1093/abbs/gms108)
- Li T & Chiang JY 2014 Bile acid signaling in metabolic disease and drug therapy. *Pharmacological Review* **66** 948–983. (doi:10.1124/pr.113.008201)
- Li JV, Ashrafian H, Bueter M, Kinross J, Sands C, le Roux CW, Bloom SR, Darzi A, Athanasiou T, Marchesi JR *et al.* 2011 Metabolic surgery profoundly influences gut microbial-host metabolic cross-talk. *Gut* **60** 1214–1223. (doi:10.1136/gut.2010.234708)
- Li F, Jiang C, Krausz KW, Li Y, Albert I, Hao H, Fabre KM, Mitchell JB, Patterson AD & Gonzalez FJ 2013 Microbiome remodelling leads to inhibition of intestinal farnesoid X receptor signalling and decreased obesity. *Nature Communications* **4** 2384. (doi:10.1038/ncomms3384)
- de Luis DA, Duenas A, Martin J, Abad L, Cuellar L & Aller R 2002 Light symptoms following a high-dose intentional L-thyroxine ingestion treated with cholestyramine. *Hormone Research* **57** 61–63. (doi:10.1159/000057950)
- Maglich JM, Watson J, McMillen PJ, Goodwin B, Willson TM & Moore JT 2004 The nuclear receptor CAR is a regulator of thyroid hormone metabolism during caloric restriction. *Journal of Biochemical Chemistry* **279** 19832–19838. (doi:10.1074/jbc.M313601200)
- Maglich JM, Lobe DC & Moore JT 2009 The nuclear receptor CAR (NR1H3) regulates serum triglyceride levels under conditions of metabolic stress. *Journal of Lipid Research* **50** 439–445. (doi:10.1194/jlr.M800226-JLR200)
- Malisova L, Kovacova Z, Koc M, Kracmerova J, Stich V & Rossmeislova L 2013 Ursodeoxycholic acid but not tauroursodeoxycholic acid inhibits proliferation and differentiation of human subcutaneous adipocytes. *PLoS ONE* **8** e82086. (doi:10.1371/journal.pone.0082086)
- Martin JH, Deacon CF, Gorrell MD & Prins JB 2011 Incretin-based therapies—review of the physiology, pharmacology and emerging clinical experience. *Internal Medicine Journal* **41** 299–307. (doi:10.1111/j.1445-5994.2011.02439.x)
- Mathew S, Lund RJ, Chaudhary LR, Geurs T & Hruska KA 2008 Vitamin D receptor activators can protect against vascular calcification. *Journal of the American Society of Nephrology* **19** 1509–1519. (doi:10.1681/ASN.2007080902)
- Mazuy C, Helleboid A, Stael B & Lefebvre P 2015 Nuclear bile acid signaling through the farnesoid X receptor. *Cellular and Molecular Life Sciences* **72** 1631–1650. (doi:10.1007/s00018-014-1805-y)
- McGarr SE, Ridlon JM & Hylemon PB 2005 Diet, anaerobic bacterial metabolism, and colon cancer: a review of the literature. *Journal of Clinical Gastroenterology* **39** 98–109.
- Modica S, Gadaleta RM & Moschetta A 2010 Deciphering the nuclear bile acid receptor FXR paradigm. *Nuclear Receptor Signaling* **8** e005. (doi:10.1621/nrs.08005)
- Mudaliar S, Henry RR, Sanyal AJ, Morrow L, Marschall HU, Kipnes M, Adorini L, Sciacca CI, Clopton P, Castelloe E *et al.* 2013 Efficacy and

- safety of the farnesoid X receptor agonist obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver disease. *Gastroenterology* **145** 574–582. (doi:10.1053/j.gastro.2013.05.042)
- Musri MM, Carmona MC, Hanzu FA, Kaliman P, Gomis R & Parrizas M 2010 Histone demethylase LSD1 regulates adipogenesis. *Journal of Biochemical Chemistry* **285** 30034–30041. (doi:10.1074/jbc.M110.151209)
- Myronovych A, Kirby M, Ryan KK, Zhang W, Jha P, Setchell KD, Dexheimer PJ, Aronow B, Seeley RJ & Kohli R 2014 Vertical sleeve gastrectomy reduces hepatic steatosis while increasing serum bile acids in a weight-loss-independent manner. *Obesity* **22** 390–400. (doi:10.1002/oby.20548)
- Nauck MA, Vardarli I, Deacon CF, Holst JJ & Meier JJ 2011 Secretion of glucagon-like peptide-1 (GLP-1) in type 2 diabetes: what is up, what is down? *Diabetologia* **54** 10–18. (doi:10.1007/s00125-010-1896-4)
- Neish AS 2009 Microbes in gastrointestinal health and disease. *Gastroenterology* **136** 65–80. (doi:10.1053/j.gastro.2008.10.080)
- Neuschwander-Tetri BA, Loomba R, Sanyal AJ, Lavine JE, Van Natta ML, Abdelmalek MF, Chalasani N, Dasarthy S, Diehl AM, Hameed B *et al.* 2015 Farnesoid X nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a multicentre, randomised, placebo-controlled trial. *Lancet* **385** 956–965. (doi:10.1016/S0140-6736(14)61933-4)
- O'Brien PE 2010 Bariatric surgery: mechanisms, indications and outcomes. *Journal of Gastroenterology and Hepatology* **25** 1358–1365. (doi:10.1111/j.1440-1746.2010.06391.x)
- Ockenga J, Valentini L, Schuetz T, Wohlgemuth F, Glaeser S, Omar A, Kasim E, duPlessis D, Featherstone K, Davis JR *et al.* 2012 Plasma bile acids are associated with energy expenditure and thyroid function in humans. *Journal of Clinical Endocrinology and Metabolism* **97** 535–542. (doi:10.1210/jc.2011-2329)
- Park JS, Seo JH & Youn HS 2013 Gut microbiota and clinical disease: obesity and nonalcoholic fatty liver disease. *Pediatric Gastroenterology, Hepatology and Nutrition* **16** 22–27. (doi:10.5223/pghn.2013.16.1.22)
- Parker HE, Wallis K, le Roux CW, Wong KY, Reimann F & Gribble FM 2012 Molecular mechanisms underlying bile acid-stimulated glucagon-like peptide-1 secretion. *British Journal of Pharmacology* **165** 414–423. (doi:10.1111/j.1476-5381.2011.01561.x)
- Parlesak A, Schaeckeler S, Moser L & Bode C 2007 Conjugated primary bile salts reduce permeability of endotoxin through intestinal epithelial cells and synergize with phosphatidylcholine in suppression of inflammatory cytokine production. *Critical Care Medicine* **35** 2367–2374. (doi:10.1097/01.CCM.0000284586.84952.FB)
- Patti ME, Houten SM, Bianco AC, Bernier R, Larsen PR, Holst JJ, Badman MK, Maratos-Flier E, Mun EC, Pihlajamaki J *et al.* 2009 Serum bile acids are higher in humans with prior gastric bypass: potential contribution to improved glucose and lipid metabolism. *Obesity* **17** 1671–1677. (doi:10.1038/oby.2009.102)
- Penney NC, Kinross J, Newton RC & Purkayastha S 2015 The role of bile acids in reducing the metabolic complications of obesity after bariatric surgery: a systematic review. *International Journal of Obesity* **39** 1565–1574. (doi:10.1038/ijo.2015.115)
- Perino A, Pols TW, Nomura M, Stein S, Pellicciari R & Schoonjans K 2014 TGR5 reduces macrophage migration through mTOR-induced C/EBP β differential translation. *Journal of Clinical Investigation* **124** 5424–5436. (doi:10.1172/JCI176289)
- Pories WJ 2008 Bariatric surgery: risks and rewards. *Journal of Clinical Endocrinology and Metabolism* **93** S89–S96. (doi:10.1210/jc.2008-1641)
- Potthoff MJ, Klier SA & Mangelsdorf DJ 2012 Endocrine fibroblast growth factors 15/19 and 21: from feast to famine. *Genes and Development* **26** 312–324. (doi:10.1101/gad.184788.111)
- Pournaras DJ, Glicksman C, Vincent RP, Kuganlipava S, Alaghband-Zadeh J, Mahon D, Bekker JH, Ghatei MA, Bloom SR, Walters JR *et al.* 2012 The role of bile after Roux-en-Y gastric bypass in promoting weight loss and improving glycaemic control. *Endocrinology* **153** 3613–3619. (doi:10.1210/en.2011-2145)
- Qi Y, Jiang C, Cheng J, Krausz KW, Li T, Ferrell JM, Gonzalez FJ & Chiang JY 2015 Bile acid signaling in lipid metabolism: metabolomic and lipidomic analysis of lipid and bile acid markers linked to anti-obesity and anti-diabetes in mice. *Biochimica Biophysica Acta* **1851** 19–29. (doi:10.1016/j.bbali.2014.04.008)
- Qiao L, Studer E, Leach K, McKinstry R, Gupta S, Decker R, Kukreja R, Valerie K, Nagarkatti P, El Deiry W *et al.* 2001 Deoxycholic acid (DCA) causes ligand-independent activation of epidermal growth factor receptor (EGFR) and FAS receptor in primary hepatocytes: Inhibition of EGFR/mitogen-activated protein kinase-signaling module enhances DCA-induced apoptosis. *Molecular Biology of the Cell* **12** 2629–2645. (doi:10.1091/mbc.12.9.2629)
- Raufman JP, Cheng K & Zimniak P 2003 Activation of muscarinic receptor signaling by bile acids: physiological and medical implications. *Digestive Diseases and Sciences* **48** 1431–1444. (doi:10.1023/A:1024733500950)
- Ren K, Tang ZL, Jiang Y, Tan YM & Yi GH 2015 Apolipoprotein M. *Clinical Chimica Acta* **446** 21–29. (doi:10.1016/j.cca.2015.03.038)
- Renga B, Mencarelli A, Vavassori P, Brancalone V & Fiorucci S 2010 The bile acid sensor FXR regulates insulin transcription and secretion. *Biochimica Biophysica Acta* **1802** 363–372. (doi:10.1016/j.bbadis.2010.01.002)
- Renga B, Mencarelli A, Cipriani S, D'Amore C, Carino A, Bruno A, Francisci D, Zampella A, Distrutti E & Fiorucci S 2013 The bile acid sensor FXR is required for immune-regulatory activities of TLR-9 in intestinal inflammation. *PLoS ONE* **8** e54472. (doi:10.1371/journal.pone.0054472)
- Ridlon JM, Kang DJ, Hylemon PB & Bajaj JS 2014 Bile acids and the gut microbiome. *Current Opinion in Gastroenterology* **30** 332–338. (doi:10.1097/MOG.0000000000000057)
- Rudling M, Camilleri M, Graffner H, Holst JJ & Rikner L 2015 Specific inhibition of bile acid transport alters plasma lipids and GLP-1. *BMC Cardiovascular Disorders* **15** 75. (doi:10.1186/s12872-015-0070-9)
- Ryan KK, Tremaroli V, Clemmensen C, Kovatcheva-Datchary P, Myronovych A, Karns R, Wilson-Perez HE, Sandoval DA, Kohli R, Backhed F *et al.* 2014 FXR is a molecular target for the effects of vertical sleeve gastrectomy. *Nature* **509** 183–188. (doi:10.1038/nature13135)
- Salmoirago-Blotcher E, Crawford S, Jackson E, Ockene J & Ockene I 2011 Constipation and risk of cardiovascular disease among postmenopausal women. *American Journal of Medicine* **124** 714–723. (doi:10.1016/j.amjmed.2011.03.026)
- Sayin SI, Wahlstrom A, Felin J, Jantti S, Marschall HU, Bamberg K, Angelin B, Hyotylainen T, Oresic M & Backhed F 2013 Gut microbiota regulates bile acid metabolism by reducing the levels of tauro- β -muricholic acid, a naturally occurring FXR antagonist. *Cell Metabolism* **17** 225–235. (doi:10.1016/j.cmet.2013.01.003)
- Schittenhelm B, Wagner R, Kahny V, Peter A, Krippeit-Drews P, Dufer M & Drews G 2015 Role of FXR in β -cells of lean and obese mice. *Endocrinology* **156** 1263–1271. (doi:10.1210/en.2014-1751)
- Sebastian-Ochoa A, Quesada-Charneco M, Fernandez-Garcia D, Reyes-Garcia R, Rozas-Moreno P & Escobar-Jimenez F 2008 Dramatic response to cholestyramine in a patient with Graves' disease resistant to conventional therapy. *Thyroid* **18** 1115–1117. (doi:10.1089/thy.2008.0094)
- Seok S, Kanamaluru D, Xiao Z, Ryerson D, Choi SE, Suino-Powell K, Xu HE, Veenstra TD & Kemper JK 2013 Bile acid signal-induced phosphorylation of small heterodimer partner by protein kinase C ζ is critical for epigenomic regulation of liver metabolic genes. *Journal of Biochemical Chemistry* **288** 23252–23263. (doi:10.1074/jbc.M113.452037)
- Seyer P, Vallois D, Poitry-Yamate C, Schutz F, Metref S, Tarussio D, Maechler P, Staels B, Lanz B, Grueter R *et al.* 2013 Hepatic glucose sensing is required to preserve β cell glucose competence. *Journal of Clinical Investigation* **123** 1662–1676. (doi:10.1172/JCI65538)
- Shakir KM, Michaels RD, Hays JH & Potter BB 1993 The use of bile acid sequestrants to lower serum thyroid hormones in iatrogenic hyperthyroidism. *Annals of Internal Medicine* **118** 112–113. (doi:10.7326/0003-4819-118-2-199301150-00006)

- Shakir YA, Samsioe G, Khatibi EA, Nyberg P, Lidfeldt J, Agardh CD & Nerbrand C 2007 Health hazards in middle-aged women with cardiovascular disease: a case-control study of Swedish women. The women's health in the Lund area (WHILA) study. *Journal of Women's Health* **16** 406–414. (doi:10.1089/jwh.2006.0056)
- Shen H, Zhang Y, Ding H, Wang X, Chen L, Jiang H & Shen X 2008 Farnesoid X receptor induces GLUT4 expression through FXR response element in the GLUT4 promoter. *Cellular Physiology and Biochemistry* **22** 1–14. (doi:10.1159/000149779)
- da-Silva WS, Ribich S, Arrojo e Drigo R, Castillo M, Patti ME & Bianco AC 2011 The chemical chaperones tauroursodeoxycholic and 4-phenylbutyric acid accelerate thyroid hormone activation and energy expenditure. *FEBS Letter* **585** 539–544. (doi:10.1016/j.febslet.2010.12.044)
- Sipka S & Bruckner G 2014 The immunomodulatory role of bile acids. *International Archives of Allergy and Immunology* **165** 1–8. (doi:10.1159/000366100)
- Smith Z, Ryerson D & Kemper JK 2013 Epigenomic regulation of bile acid metabolism: emerging role of transcriptional cofactors. *Molecular Cell Endocrinology* **368** 59–70. (doi:10.1016/j.mce.2012.04.008)
- Song Y, Xu C, Shao S, Liu J, Xing W, Xu J, Qin C, Li C, Hu B, Yi S *et al.* 2015 Thyroid-stimulating hormone regulates hepatic bile acid homeostasis via SREBP-2/HNF-4a/CYP7A1 axis. *Journal of Hepatology* **62** 1171–1179. (doi:10.1016/j.jhep.2014.12.006)
- Staels B & Kuipers F 2007 Bile acid sequestrants and the treatment of type 2 diabetes mellitus. *Drugs* **67** 1383–1392. (doi:10.2165/00003495-200767100-00001)
- Suzuki T, Oba K, Futami-Suda S, Suzuki K, Ouchi M, Igari Y, Matsumura N, Watanabe K, Kigawa Y & Nakano H 2007a Effects of colestimide on blood glucose-lowering activity and body weight in patients with type 2 diabetes and hypercholesterolemia. *Journal of Nippon Medical School* **74** 81–84. (doi:10.1272/jnms.74.81)
- Suzuki T, Oba K, Igari Y, Matsumura N, Watanabe K, Futami-Suda S, Yasuoka H, Ouchi M, Suzuki K, Kigawa Y *et al.* 2007b Colestimide lowers plasma glucose levels and increases plasma glucagon-like peptide-1 (7-36) levels in patients with type 2 diabetes mellitus complicated by hypercholesterolemia. *Journal of Nippon Medical School* **74** 338–343. (doi:10.1272/jnms.74.338)
- Talley NJ, Jones M, Nuyts G & Dubois D 2003 Risk factors for chronic constipation based on a general practice sample. *American Journal of Gastroenterology* **98** 1107–1111. (doi:10.1111/j.1572-0241.2003.07465.x)
- Thomas C, Gioiello A, Noriega L, Strehle A, Oury J, Rizzo G, Macchiarulo A, Yamamoto H, Matakic C, Pruzanski M *et al.* 2009 TGR5-mediated bile acid sensing controls glucose homeostasis. *Cell Metabolism* **10** 167–177. (doi:10.1016/j.cmet.2009.08.001)
- Tilg H & Moschen AR 2014 Microbiota and diabetes: an evolving relationship. *Gut* **63** 1513–1521. (doi:10.1136/gutjnl-2014-306928)
- Tomlinson E, Fu L, John L, Hultgren B, Huang X, Renz M, Stephan JP, Tsai SP, Powell-Braxton L, French D *et al.* 2002 Transgenic mice expressing human fibroblast growth factor-19 display increased metabolic rate and decreased adiposity. *Endocrinology* **143** 1741–1747. (doi:10.1210/endo.143.5.8850)
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER & Gordon JI 2006 An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **444** 1027–1031. (doi:10.1038/nature05414)
- Vavassori P, Mencarelli A, Renga B, Distrutti E & Fiorucci S 2009 The bile acid receptor FXR is a modulator of intestinal innate immunity. *Journal of Immunology* **183** 6251–6261. (doi:10.4049/jimmunol.0803978)
- Vinik AI, Maser RE, Mitchell BD & Freeman R 2003 Diabetic autonomic neuropathy. *Diabetes Care* **26** 1553–1579. (doi:10.2337/diacare.26.5.1553)
- Vítek L 2015 Bile acid malabsorption in inflammatory bowel disease. *Inflammatory Bowel Diseases* **21** 476–483. (doi:10.1097/MIB.000000000000193)
- Vrieze A, Schopman JE, Admiraal WM, Soeters MR, Nieuwdorp M, Verberne HJ & Holleman F 2012 Fasting and postprandial activity of brown adipose tissue in healthy men. *Journal of Nuclear Medicine* **53** 1407–1410. (doi:10.2967/jnumed.111.100701)
- Wang X, Fang X, Zhou J, Chen Z, Zhao B, Xiao L, Liu A, Li YS, Shyy JY, Guan Y *et al.* 2013 Shear stress activation of nuclear receptor PXR in endothelial detoxification. *PNAS* **110** 13174–13179. (doi:10.1073/pnas.1312065110)
- Wang S, Lei T, Zhang K, Zhao W, Fang L, Lai B, Han J, Xiao L & Wang N 2014 Xenobiotic pregnane X receptor (PXR) regulates innate immunity via activation of NLRP3 inflammasome in vascular endothelial cells. *Journal of Biochemical Chemistry* **289** 30075–30081. (doi:10.1074/jbc.M114.578781)
- Watanabe M, Houten SM, Matakic C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney JW, Ezaki O, Kodama T *et al.* 2006 Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. *Nature* **439** 484–489. (doi:10.1038/nature04330)
- Wu W, Liu X, Peng X, Xue R, Ji L, Shen X, Chen S, Gu J & Zhang S 2014 Bile acids override steatosis in farnesoid X receptor deficient mice in a model of non-alcoholic steatohepatitis. *Biochemical and Biophysical Research Communications* **448** 50–55. (doi:10.1016/j.bbrc.2014.04.048)
- Yang TT, Chang CK, Tsao CW, Hsu YM, Hsu CT & Cheng JT 2009 Activation of muscarinic M-3 receptor may decrease glucose uptake and lipolysis in adipose tissue of rats. *Neuroscience Letter* **451** 57–59. (doi:10.1016/j.neulet.2008.12.029)
- Yang Y, Hwang S, Kim M, Lim Y, Kim MH, Lee S, Lim DJ, Kang MI & Cha BY 2015 Refractory Graves' disease successfully cured by adjunctive cholestyramine and subsequent total thyroidectomy. *Endocrinology and Metabolism* [in press].
- Yusta B, Holland D, Waschek JA & Drucker DJ 2012 Intestintrophic glucagon-like peptide-2 (GLP-2) activates intestinal gene expression and growth factor-dependent pathways independent of the vasoactive intestinal peptide gene in mice. *Endocrinology* **153** 2623–2632. (doi:10.1210/en.2012-1069)
- Zhang J, Huang W, Qatanani M, Evans RM & Moore DD 2004 The constitutive androstane receptor and pregnane X receptor function coordinately to prevent bile acid-induced hepatotoxicity. *Journal of Biochemical Chemistry* **279** 49517–49522. (doi:10.1074/jbc.M409041200)
- Zhang Y, Lee FY, Barrera G, Lee H, Vales C, Gonzalez FJ, Willson TM & Edwards PA 2006 Activation of the nuclear receptor FXR improves hyperglycemia and hyperlipidemia in diabetic mice. *PNAS* **103** 1006–1011. (doi:10.1073/pnas.0506982103)
- Zhang H, DiBaise JK, Zuccolo A, Kudrna D, Braidotti M, Yu Y, Parameswaran P, Crowell MD, Wing R, Rittmann BE *et al.* 2009 Human gut microbiota in obesity and after gastric bypass. *PNAS* **106** 2365–2370. (doi:10.1073/pnas.0812600106)
- Zhou H & Hylemon PB 2014 Bile acids are nutrient signaling hormones. *Steroids* **86C** 62–68. (doi:10.1016/j.steroids.2014.04.016)
- Zhou JH, Pan ZY, Zhang YF, Cui WY, Long CL & Wang H 2014 Stimulation of endothelial non-neuronal muscarinic receptor attenuates the progression of atherosclerosis via inhibiting endothelial cells activation. *Zhongguo Ying Yong Sheng Li Xue Za Zhi* **30** 549–559.
- Zwicker BL & Agellon LB 2013 Transport and biological activities of bile acids. *International Journal of Biochemistry & Cell Biology* **45** 1389–1399. (doi:10.1016/j.biocel.2013.04.012)

Received in final form 30 December 2015

Accepted 5 January 2016

Accepted Preprint published online 5 January 2016