

Material and Methods

Source of islets and tissues. All animal experimentation was approved by the Institutional Animal Care and Use Committee at the University of Iowa. Adult ferret islets were isolated as previous described (1). Non-diabetic human islets were obtained from the Integrated Islet Distribution Program.

Generation of recombinant adenoviruses to knockdown CFTR. To knockdown CFTR expression in WT ferret and human islets, two adenoviral vectors expressing a shRNA to human or ferret CFTR were generated to the same region of the *CFTR* cDNA (Exon 14; 2083-2111 bp of the transcript with the start codon ATG as +1). The coding sequence homologous to the 22 bp shRNA core (*italics*) was 5'-

AAAAAAGGAAGAATTCTATTCTCAATCC-3' for human *CFTR* and 5'-AAAAAAGGAAGAACTCT

ATTCTTAATCC-3' for ferret *CFTR*. For construction of the adenoviral vector expressing shRNA-CFTR,

double stranded oligonucleotides for the following complete shRNA sequences were synthesized and cloned downstream of the murine U6 promoter within the paced shuttle vector (Vector core, University of Iowa):

Human 5'-GAAGAATTCTATTCTCAATCTGTAAAGCCACAGATGGGATTGAGAATAG

AATCTTCCTTTTTT-3'; Ferret 5'-GAAGAACTCTATTCTTAATCTGTAAAGCCACAGATGGGA

TTAAGAATAGAGTTCTTCCTTTTTT-3'. Similarly, a scrambled shRNA was also generated with a core

sequence (*italics*) of 5'-TTTTTTATTGCGTTAGTTATACTAATCA-3' using synthetic oligonucleotides of 5'-

AACGCAATCAATGATTAGTCTGTAAAGCCACAGATGGGACTAAT

CATTGATTGCGTTATTTTTT-3'. Recombinant adenoviral vectors were generated by the co-transfection

of 293 cells with the linearized proviral plasmid containing the shRNA cassette in the E1 region and an

adenoviral genome plasmid containing an RSV-driven eGFP cassette in the E3 region, as previously

described (28). Viral stocks were amplified by CsCl banding and desalted prior to use. Viruses are

designated: Ad.fCFTR-shRNA (ferret *CFTR* targeting vector), Ad.hCFTR-shRNA (human *CFTR* targeting

vector), and Ad.Scr-shRNA (scrambled shRNA control vector).

CFTR knockdown in ferret and human islets using recombinant adenovirus. *CFTR* gene expression was

inhibited in isolated ferret and human islets using recombinant adenovirus (Ad.fCFTR-shRNA and

Ad.hCFTR-shRNA) encoding an shRNA against each *CFTR* species. Virus containing a scrambled shRNA

(Ad.Scr-shRNA) was used as a negative control. Approximately 100 adult human or ferret islets were cultured with recombinant adenoviruses (1.0×10^9 particles/ml) in RPMI1640 without serum for 4hrs and then supplemented with 10% fetal bovine serum thereafter. After 24hrs, the transduced islets were transferred into fresh culture medium for another 24hrs prior to using the islets for experiments.

CFTR RNA quantification in islets. CFTR RNA expression was measured by the QuantiGene Plex Assay Kit (Affymetrix). Bead-based oligonucleotide probe sets specific for ferret CFTR and peptidylprolyl isomerase B (PPIB) RNA were developed by Affymetrix, as detailed in the RNA probe map on the next page. Islet homogenates were prepared using the QuantiGene Sample Processing Kit (Affymetrix) and quantified as previously described (2), using PPIB as the internal control (housekeeping gene) by which to normalize expression for each sample.

Islet hormone secretion assays. Insulin secretion assays using isolated islets were performed in static culture as previously described (3). Briefly, five adult islets (150-200 μ m diameter) were equilibrated for 1hr in 1.67mM glucose Krebs-Ringers Bicarbonate Buffer (KRB: 120mM NaCl, 4.8mM KCl, 2.5mM CaCl₂, 1.2mM MgCl₂, 20mM NaHCO₃, 5mM HEPES, pH7.4) at 37°C (human islets) or 38.5°C (ferret islets). Each group of islets was then divided equally into either 1.67 or 16.7mM glucose KRB and cultured for 1hr. Insulin was measured using human insulin ELISA kit (Calbiotech, Cat#IS130D). Insulin release was calculated on a per islet basis as previously described (2).

1. Li DS, Yuan YH, Tu HJ, Liang QL, Dai LJ 2009 A protocol for islet isolation from mouse pancreas. *Nat Protoc* **4** 1649-1652.
2. Yi Y, Sun X, Gibson-Corley K, Xie W, Liang B, He N, Tyler SR, Uc A, Philipson LH, Wang K, Hara M, Ode KL, Norris AW, Engelhardt JF 2016 A Transient Metabolic Recovery from Early Life Glucose Intolerance in Cystic Fibrosis Ferrets Occurs During Pancreatic Remodeling. *Endocrinology* **157** 1852-1865.
3. Olivier AK, Yi Y, Sun X, Sui H, Liang B, Hu S, Xie W, Fisher JT, Keiser NW, Lei D, Zhou W, Yan Z, Li G, Evans TI, Meyerholz DK, Wang K, Stewart ZA, Norris AW, Engelhardt JF 2012 Abnormal endocrine pancreas function at birth in cystic fibrosis ferrets. *J Clin Invest* **122** 3755-3768

RNA Probe Map

Capture Extender Oligo (CE) is BLUE

Label Extender Oligo (LE) is RED

Blocking Probe Oligo (BL) is GREEN

>gi|859759950|ref|XM_004741906| PREDICTED: Mustela putorius furo cystic fibrosis transmembrane conductance regulator (ATP-binding cassette sub-family C, member 7) (CFTR), transcript variant X2, mRNA. [Mustela putorius furo] MAM 01-JUL-2015

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